

This author's PDF version corresponds to the article as it appeared upon acceptance. Fully formatted PDF versions will be made available soon.

Tart cherry intake and serum uric acid: Meta-analysis of randomized controlled trials and evidence from network pharmacology

doi: 10.6133/apjcn.202603/PP.0007

Published online: March 2026

Running title: Tart cherry intake and serum uric acid

Zhenzhen Zhang MSc, Zhiyuan Feng PhD, Wei Yan PhD, Tianyu Wu PhD, Jiayue Xia PhD, Junhui Yu PhD, Jingyi Yang MSc, Yuanyuan Wang PhD, Guiju Sun PhD

Key Laboratory of Environmental Medicine and Engineering of Ministry of Education, Department of Nutrition and Food Hygiene School of Public Health, Southeast University, Nanjing, China

Authors' email addresses and contributions:

Zhenzhen Zhang MSc, email: 220234030@seu.edu.cn

Zhiyuan Feng PhD, email: 230248567@seu.edu.cn

Wei Yan PhD, email: 230259489@seu.edu.cn

Tianyu Wu PhD, email: tianyu_w@seu.edu.cn

Jiayue Xia PhD, email: 230229010@seu.edu.cn

Junhui Yu PhD, email: 230239086@seu.edu.cn

Jingyi Yang MSc, email: jingyiyang@seu.edu.cn

Yuanyuan Wang PhD, email: 230218460@seu.edu.cn

Guiju Sun PhD, email: gjsun@seu.edu.cn.

Conception and design: Zhenzhen Zhang, Yuanyuan Wang,

Fieldwork/data collection: Zhenzhen Zhang, Zhiyuan Feng, Wei Yan, Tianyu Wu, Jiayue Xia, Junhui Yu, Jingyi Yang

Data analysis and interpretation: Zhenzhen Zhang, Zhiyuan Feng, Yuanyuan Wang,

Drafting of manuscript: Zhenzhen Zhang, Zhiyuan Feng, Wei Yan, Yuanyuan Wang

Critical revision of manuscript: Junhui Yu, Jingyi Yang, Yuanyuan Wang, Guiju Sun

Overall scientific management: Guiju Sun.

Corresponding Author: Prof. Guiju Sun, Key Laboratory of Environmental Medicine and Engineering of Ministry of Education, Department of Nutrition and Food Hygiene, School of Public Health, Southeast University, Gulou District, Dingjiaqiao 87#, Nanjing, Jiangsu 210009, China. Tel: 86-13951928860. Email: gjsun@seu.edu.cn

ABSTRACT

Background and Objectives: Tart cherry products have been proposed to lower serum uric acid and reduce gout risk, but clinical findings are inconsistent. This study evaluated the effect of tart cherry intake on serum uric acid concentration and explored possible mechanisms. **Methods and Study Design:** Randomized controlled trials comparing tart cherry intake with placebo or usual diet and reporting serum uric acid concentration were identified from electronic databases. A random effects meta analysis was used to pool standardized mean differences between groups. In parallel, bioactive compounds in tart cherry were retrieved from public databases, and their potential targets related to serum uric acid and gout were investigated using network based analyses and molecular docking. **Results:** Four randomized trials (eight comparisons; 392 participants) were included. Tart cherry intake was associated with a modest reduction in serum uric acid concentration compared with control (standardized mean difference -0.22 ; 95% confidence interval -0.43 to -0.01), with substantial heterogeneity. Network and docking analyses suggested that anthocyanins and other polyphenols may act on inflammatory, oxidative stress and metabolic pathways. **Conclusions:** Tart cherry intake may modestly lower serum uric acid concentration, but evidence is limited in quantity and consistency. Larger, rigorously designed randomized trials are needed.

Key Words: tart cherry intake, serum uric acid (sUA), gout, meta-analysis, network pharmacology

INTRODUCTION

Uric acid derives from purine catabolism and may play multiple physiological roles, including functioning as an antioxidant and neuroprotective factor, helping maintain blood pressure under low-salt dietary conditions, and participating in immune regulation.¹ As an important non-enzymatic antioxidant in human plasma, uric acid scavenges reactive oxygen and nitrogen species and is involved in multiple redox reactions, thereby contributing to the maintenance of internal homeostasis.² However, serum uric acid (sUA) may remain persistently elevated and exceed the physiological saturation threshold. In such cases, monosodium urate can deposit in the joints and periarticular tissues. This leads to the activation of innate immunity and local inflammation mediated by crystals. Clinically, this presents as a complex inflammatory arthritis with erythema, warmth, marked tenderness, and severe pain—namely, gout.³⁻⁵ Epidemiologically, the global prevalence and incidence of gout

have continued to rise. Rates differ across regions and populations. Overlapping metabolic comorbidities and lifestyle factors further complicate management.^{4, 6, 7}

For recurrent gout, the effectiveness of urate-lowering therapy (ULT) and anti-inflammatory treatment has been established. International and regional guidelines also emphasize a treat-to-target approach to reduce the risk of recurrence and complications.⁷⁻¹² However, in real-world settings, long-term adherence to prescribed ULT is only about 20%–70%. This undermines sustained efficacy and prevention of recurrence.⁴ Against this backdrop, adjunctive strategies that fit daily life alongside pharmacotherapy are of practical relevance. Dietary interventions, such as tart cherry products, have attracted attention for their safety and accessibility and are increasingly in the spotlight.¹ Prior short-term interventional and observational studies suggest that intake of tart cherry juice or its concentrates is associated with reductions in sUA, downregulation of inflammatory biomarkers, and a decreased risk of gout flares. However, effect sizes vary with population characteristics (weight, comorbidities), formulation, anthocyanin dose, and intervention duration.^{3, 13-16} Given suboptimal adherence and a rising disease burden, dietary interventions can act as useful adjuncts to lifestyle management. They are most effective when combined with purine restriction, weight control, limited alcohol intake, and urinary alkalization for additive benefits.^{7, 11, 12}

From a mechanistic perspective, anthocyanins and related polyphenols are the main active constituents of tart cherries. They can inhibit cyclooxygenase (COX) activity, scavenge reactive nitrogen and oxygen species, and modulate inflammatory and oxidative stress pathways. These actions provide biological plausibility for anti-inflammatory and antioxidant effects.¹⁷⁻²⁰ When adherence falls short and the burden keeps climbing, these interventions can complement routine care.³

In vitro and animal studies suggest that tart cherry extracts or principal anthocyanins may inhibit xanthine oxidase (XO). They may also influence uric acid homeostasis by modulating the redox microenvironment and affecting renal and intestinal urate excretion.^{1, 19} Genetic and transport pathway studies indicate that urate acid transporter 1 (URAT1), glucose transporter 9 (GLUT9), and ATP binding cassette subfamily G member 2 (ABCG2) are central to renal and intestinal uric acid handling. Altered function of these transporters is closely associated with hyperuricemia and gout risk. This provides a theoretical basis for dietary effects on the transport–excretion axis.^{1, 21, 22} Although the biological rationale is robust, human evidence remains heterogeneous. Differences span study populations, sample sizes, product sources, anthocyanin quantification, intervention duration, control design, and outcome measures

including sUA, inflammatory biomarkers, and clinical flares. These factors limit the consistency and generalizability of the conclusions drawn from the studies.^{3, 13, 15, 16}

Based on these considerations, a systematic review, meta-analysis, and network pharmacology analysis were conducted. Randomized controlled trials were systematically integrated to quantify the effect of tart cherry products on sUA level, and observational and mechanistic evidence was qualitatively summarized. Subgroup analyses were prespecified by population characteristics, formulation and anthocyanin dose, intervention duration, and concurrent lifestyle measures to explore dose–response relationships and heterogeneity. Safety outcomes and long-term sustainability were also evaluated. Our aim was to provide clearer, actionable evidence for clinical practice and for subsequent standardized human trials.

MATERIALS AND METHODS

Study design

Randomized controlled trials (RCTs) served as the primary evidence for a systematic review and meta-analysis, and cohort and case–control studies were included for qualitative reference. Network pharmacology and molecular docking were integrated to explore mechanisms. The study workflow followed Preferred Reporting Items for Systematic Reviews and Meta Analyses (PRISMA) guidelines. We placed no restriction on year of publication. The study was registered in PROSPERO under the registration number CRD420261347406.

Search strategy

We conducted in-depth searches in eight databases (Cochrane Library, PubMed, MEDLINE, Embase, EBSCO, Ovid, ScienceDirect, and Web of Science) to identify relevant studies. Medical Subject Headings (MeSH) terms were incorporated to cover all eligible reports from database inception through July 2025. Full search strategies and database query strings are provided in Supplementary Table 1.

Only human studies were included, and English was used as the primary language. After deduplication, titles and abstracts were screened, followed by full-text review to assess eligibility. Publication date was not used as a restriction for searching or inclusion.

Inclusion and exclusion criteria

Inclusion criteria: (1) randomized controlled trials (parallel or crossover) in adults (≥ 18 years); (2) interventions involving tart cherry products (juice, concentrate, powder, or standardized extract derived from *Prunus cerasus L.*); (3) a control condition (e.g., placebo, isoenergetic

beverage, or no intervention); and (4) usable arm-level sUA data either pre-/post-intervention or between groups at the end of treatment. Cohort and case-control studies addressing tart cherry intake and urate-related outcomes were included only for qualitative context and not pooled quantitatively.

Exclusion criteria: (1) opinions, reviews, case reports, abstracts, or in vitro studies; (2) interventions using sweet cherries or studies that could not distinguish tart cherries; (3) no control group or missing key outcome data; and (4) duplicate data or missing sample-size information that rendered data unusable.

Data extraction, synthesis, and analysis

Methods included extraction of the following information: author and publication year; country or region; study design and blinding; sample size and population characteristics; baseline sUA; intervention preparation (formulation, dose, frequency, duration); type of control; measurement time points; and statistics related to sUA. Where unit inconsistency was present, sUA values were standardized to mg/dL ($1 \text{ mg/dL} \approx 59.5 \text{ } \mu \text{ mol/L}$). As needed, authors were contacted or estimates were derived using handbook methods. For crossover designs, measurements after an adequate washout were prioritized. For multi-arm studies, arm-specific doses or formulations were recorded, and pooled data were processed under prespecified rules.

Study characteristics and outcome definitions were summarized in Supplementary Table 2.

Quality assessment and Risk of Bias (RoB)

Risk of bias for randomized controlled trials included in the quantitative synthesis was assessed using the Cochrane Risk of Bias tool, version 2 (RoB 2). Two reviewers independently evaluated each domain (randomization process, deviations from intended interventions, missing outcome data, measurement of the outcome, and selection of the reported result). Disagreements were resolved through discussion or consultation with a third reviewer. Study-level RoB judgments are summarized in Table 1, and domain-level details are provided in the Supplementary Materials.

Certainty of evidence Grading of Recommendations Assessment, Development and Evaluation (GRADE) assessment

The GRADE framework was used to assess the certainty of prespecified outcomes. In randomized evidence, certainty starts at a high level and may be downgraded across five domains: risk of bias, inconsistency, indirectness, imprecision, and publication bias (or other concerns). Downgrading decisions follow standardized GRADE criteria and are accompanied by explicit justification for each downgrade. Where appropriate, potential upgrading considerations may also be discussed, but upgrading is uncommon in typical randomized evidence.

Statistical analysis

We pooled effects using random-effects models with restricted maximum likelihood (REML) to estimate between-study variance (τ^2) and applied Hartung–Knapp–Sidik–Jonkman adjustments for confidence intervals. Because trials reported sUA using different summary formats (change-from-baseline versus end-of-treatment values), the primary effect measure was the standardized mean difference (SMD). We report pooled SMDs with 95% confidence intervals (CIs), along with τ^2 and I^2 as measures of between-study heterogeneity. Change-from-baseline values were preferentially used; when unavailable, end-of-treatment between-group differences were analyzed.

Crossover trials were analyzed as paired designs assuming a within-person correlation $r = 0.5$. For multi-arm trials, we derived separate randomized comparisons by either equally splitting shared control groups or combining similar intervention arms, following Cochrane Handbook guidance, to ensure that each participant contributed only once to each comparison.

Potential small-study effects and publication bias were explored using funnel plots and Egger’s regression test, recognizing that these methods are difficult to interpret when fewer than 10 comparisons are available. All meta-analyses were conducted using Stata (meta suite) and cross-checked in RevMan.

Network pharmacology database

Key constituents of tart cherries, primarily anthocyanins along with chlorogenic acid, p-coumaric acid, and quercetin were identified using Simplified Molecular Input Line Entry System (SMILES) strings retrieved from PubChem. Target prediction was conducted with SwissTargetPrediction, Similarity Ensemble Approach(SEA), and TargetNET, followed by deduplication and standardization to UniProt identifiers. Gene sets related to uric acid and gout were retrieved from GeneCards and intersected with the tart cherry–related targets to identify candidate targets associated with tart cherries. We constructed a protein-protein

interaction (PPI) network using Search Tool for the Retrieval of Interacting Genes/Proteins (STRING) (human; confidence 70) and visualized it in Cytoscape 3.10.0, where hub genes were identified using CytoHubba. Protein 3D structures were obtained from the Protein Data Bank (PDB), and docking was performed using AutoDock Vina to evaluate binding affinities based on binding energy.

Protocol registration

This review was not prospectively registered on PROSPERO or OSF, but the study followed PRISMA 2020 recommendations.

RESULTS

Study selection

The flow diagram of study selection is shown in Figure 1. According to the inclusion and exclusion criteria, we searched eight English-language databases: the Cochrane Library, PubMed, MEDLINE, Embase, EBSCO, Ovid, ScienceDirect, and Web of Science, identifying a total of 101 records. 36 duplicates were removed, leaving 65 studies for screening. A further 46 studies were excluded after screening titles and abstracts. The remaining 19 studies underwent full-text review, of which 11 were excluded for the following reasons: not retrieved, the intervention involved sweet cherries, non-human study, and the assessed outcome was not sUA level.

Finally, four RCTs met the inclusion criteria and were included in the meta-analysis, contributing eight randomized comparisons in total because some trials had multiple intervention or control arms. Among these four RCTs, three were published within the past 5 years, comprising a total of 392 participants. All studies measured uric acid levels using routine blood sampling. Two trials were double-blind, one was a parallel-group trial, and one used a crossover design. Martin KR used a double-blind, placebo-controlled, two-by-two crossover design in which placebo beverages were masked in color and were equivalent in sugar and energy to the real beverage to allow for the potential for blinding, with crossover through automated online randomization. Gonzalez DE achieved double-blinding by matching cherry powder capsules exactly in size and color to placebo in identically labeled bottles in a randomized, double-blind, placebo-controlled, counter-balanced, cross-over study with washout periods. Wang was a parallel controlled open-label randomized controlled trial with a three-arm design. Stamp is also an open-label study. Detailed study characteristics are provided in Supplementary Table 2.

Quality RoB

We used Review Manager 5.4 to implement the RoB 2 tool for the four included RCTs and to summarize study quality.

As shown in Table 1, the RoB assessment was conducted in accordance with the RoB framework. With regard to the randomization process, four studies were at low RoB. Concerning deviations from intended interventions, all four studies were at low RoB, primarily due to minimal and non-differential loss to follow-up. In terms of outcome assessment, four studies were likewise judged to be at low RoB. Nevertheless, several studies did not provide sufficient details regarding blinding of outcome assessors. All included studies demonstrated consistency with prespecified outcomes, with no indication of selective outcome reporting.

Certainty of evidence (GRADE)

GRADE judgments are shown in Table 2. Overall certainty varied by dose and outcome, with notable imprecision at several dose levels. The 7.5 mL dose group had a small sample size and large within-group variation, contributing to imprecise effect estimates and wide CIs. Small sample sizes and mid-range doses increase random error; these uncertainties should be highlighted in the discussion and considered in sensitivity analyses. When dose differences are substantial and samples are small, extrapolation to unstudied doses is limited; dose-subgroup analyses should be presented separately for the highest dose, with explicit emphasis on uncertainty. Some individual studies provided higher certainty for their specific outcomes, but overall certainty remains limited due to imprecision and inconsistency.

Overall effect size

Among the four included RCTs, some trials had multiple intervention or control arms. For the purposes of meta-analysis, we conducted group-wise comparisons within these multi-arm designs, resulting in eight randomized comparisons derived from the four parent trials.

The forest plot indicated statistical significance at the 5% level. Under a random-effects model, the overall SMD for uric acid levels comparing tart cherry intake with control was -0.22 (95% CI: -0.43 to -0.01), suggesting a modest reduction in sUA associated with the intervention (Figure 2). Between-study heterogeneity was substantial ($I^2=88\%$; $\text{Chi}^2 = 59.1$, $\text{df} = 7$, $p < 0.01$), indicating marked variability in effect estimates across comparisons. Given the small number of trials and the high heterogeneity, the magnitude of the pooled effect should be interpreted with caution. Funnel-plot inspection and Egger's test ($p < 0.01$)

suggested some asymmetry; however, with fewer than 10 randomized comparisons, these findings are exploratory and cannot be considered definitive evidence of publication bias (Figure 3).

Dots represent individual studies; the dashed triangle denotes the 95% confidence limits, and the vertical dashed line indicates the pooled effect (SMD: -0.22). A slight right shift and mild asymmetry suggest the presence of small-study effects or publication bias, although most points fall within the contours.

Testing of sensitivity

In the leave-one-out analysis, each study was sequentially omitted and the remainder re-pooled. The pooled effect did not reverse direction, and statistical significance was largely maintained. No single study with a decisive influence on the overall result was identified (Figure 4A). The funnel plot showed mild asymmetry consistent with Egger's test.

Network pharmacological analysis

All putative targets collected from each platform were subsequently deduplicated and merged, ultimately yielding 131 high-confidence targets. Finally, gene symbols of the predicted targets were retrieved from the UniProt database. Meanwhile, sUA-related targets were identified using the GeneCards database and were intersected with the tart cherry-related targets, yielding 78 common targets. These targets were defined as those related to tart cherry's effect on lowering sUA level. To further identify key functional regulatory targets, the aforementioned sUA-lowering tart cherry-related targets were imported into the STRING database to construct a PPI network (Figure 5A).

After construction, the network was imported into Cytoscape for visualization, and the CytoHubba plugin was used to perform topological analysis of the PPI network. Using algorithms such as Degree and Maximal Clique Centrality (MCC) to score the nodes, top 7 genes were ultimately identified as hub genes, namely interleukin 6 (IL6), tumor necrosis factor (TNF), AKT serine/threonine kinase 1 (AKT1), peroxisome proliferator activated receptor gamma (PPARG), tumor protein p53 (TP53), insulin (INS), and adiponectin (ADIPOQ) (Figure 5B).

Finally, we performed docking between the above 7 hub targets—IL6, INS, PPARG, AKT1, TNF, TP53, and ADIPOQ and anthocyanins, chlorogenic acid, p-coumaric acid, and quercetin (Figure 6). The molecular docking results showed binding energies below -5.0 kcal/mol between the four classes of compounds and the selected hub targets. Among them,

anthocyanins with ADIPOQ exhibited the strongest docking ability, followed by quercetin with IL6, and anthocyanins with both IL6 and PPARG (Figure 7). This suggests that tart cherry compounds may target these hubs to modulate sUA level in humans.

DISCUSSION

Prespecified dose-group analyses did not eliminate substantial between-study heterogeneity ($I^2 \approx 88\%$). Across eight randomized controlled trials, tart cherry products were associated with a small and unstable reduction in sUA.³ Variation in baseline characteristics, product formulation/anthocyanin content, dosing and timing, control design, and outcome definitions likely underlie this inconsistency.²³ Future work should standardize exposure, increase sample sizes, and harmonize outcomes to identify sources of heterogeneity and strengthen the evidence base.²⁴ Nevertheless, integrating RCT data with network pharmacology and molecular docking analyses provides a framework to explore potential mechanisms and assess concordance and discordance across evidence streams.⁵

Based on a systematic integration of four randomized controlled trials contributing eight randomized comparisons,^{3, 25-27} this study incorporated network pharmacology and molecular docking results,^{28, 29} to re-evaluate the relationship between tart cherry intake and serum uric acid. Overall, tart cherry intake was associated with lower sUA in our pooled analysis (SMD: -0.22 ; 95% CI: -0.43 to -0.01), but between-study heterogeneity was very high ($I^2 = 88\%$). Egger's test and slight funnel-plot asymmetry suggested possible small-study effects and publication bias.³⁰ though such tests are difficult to interpret with so few comparisons. While most comparisons trend toward lower sUA with tart cherry intake, wide confidence intervals and marked heterogeneity mean that both the magnitude and the consistency of the effect remain uncertain.³¹

Although tart cherry intake was associated with lower sUA, the evidence varied by dose and was highly heterogeneous across studies.²⁵ Thus, conclusions should be interpreted with caution, and tart cherry should be viewed as an adjunct to lifestyle management rather than a replacement for therapy.³² Future studies should standardize dose/formulation, increase sample sizes, and include mechanistic endpoints to clarify dose-response relationships and long-term safety.¹²

When compared to existing literature, two noteworthy trends emerge for interpretation.^{16, 33} Initially, real-world observational studies indicate a correlation between cherry consumption and a reduced risk of gout recurrence.³⁴ Conversely, the findings from rigorously randomized trials exhibit inconsistencies, as short-term interventions result in decreased sUA level in

healthy individuals or those with metabolic abnormalities.^{3,35} However, among gout patients who are particularly undergoing standard ULT or following strict dietary guidelines, the results often demonstrate a diminished effect.³⁵ The observed discrepancies arise from systematic differences across several factors, including study populations, intervention preparations, and measurement protocols.^{33,36} Population characteristics, including healthy individuals, asymptomatic hyperuricemia, and various phases of gout both intercritical and acute, influence baseline sUA, renal, and intestinal clearance capacities, body weight, insulin sensitivity, and concurrent medications (e.g., allopurinol/febuxostat, uricosurics, urinary alkalinizers), thereby affecting the response magnitude.^{7,37-39} Likewise, significant differences exist at the preparation level, as juices, concentrates, and capsules vary in terms of anthocyanin/polyphenol equivalents, bioavailability, and matrix components (including sugars, organic acids, and minerals).⁴⁰ Moreover, at the design and measurement level, studies often employ short-duration or acute observations. These can be influenced by factors such as blood sampling timing, fasting status, control of purine and alcohol intake, physical activity, and circadian rhythm. Additionally, crossover designs are susceptible to carryover effects if the washout period is insufficient.⁴¹ It is important to note that the fructose content in juice formulations may elevate uric acid levels.^{34,42} In contrast, components like citrate or malate can aid in uric acid excretion by raising urinary pH.⁴³⁻⁴⁵ The interaction of these opposing factors varies across studies and affects the overall results.³⁵ This combination of factors largely accounts for the considerable variability and slight signs of publication bias in the pooled analysis.^{30, 46} Future studies are likely to benefit from including healthy or hyperuricemic individuals without gout. Utilizing low-sugar standardized preparations with clearly defined anthocyanin levels and consistent sampling methods will likely reduce variability and improve the relevance of the findings.

The network pharmacology results provide biological support for “why it may be effective”.^{24,47,48} Beginning with the main active components of tart cherry, primarily anthocyanins, along with chlorogenic acid, p-coumaric acid, and quercetin, we identified overlapping targets related to uric acid metabolism and the inflammation–metabolism network. IL6, TNF, AKT1, PPARG, TP53, INS, and ADIPOQ are located at the hubs of the PPI network.^{47,49,50} Furthermore, molecular docking suggests that most compounds have a strong affinity for these targets, with binding energies all less than -5 kcal/mol.^{51,52} These signals are highly consistent with the pathophysiology of gout/hyperuricemia: on the one hand, inflammation amplification mediated by the nuclear factor κ B (NF κ B)/ NLR family pyrin domain containing 3 (NLRP3) axis and oxidative stress can drive purine catabolism and

enhance XO activity.^{53,54} On the other hand, insulin resistance and disordered lipid metabolism can affect the expression or function of proximal tubule transporters (URAT1/SLC22A12, GLUT9/SLC2A9, ABCG2), thereby altering uric acid reabsorption and excretion.^{55,56} Existing *in vitro* and animal studies indicate that anthocyanins possess COX-inhibitory and antioxidant activities. Quercetin and chlorogenic acid can inhibit XO under certain conditions.^{57,58} If they act together with citrate salts in raising urinary pH, the clinically observed “modest decrease” becomes explicable.⁴³ Molecular docking provides affinity-level evidence but cannot substitute for *in vivo* efficacy. These mechanistic findings are inherently hypothesis-generating and do not compensate for the limitations of the clinical evidence base. The parent forms of anthocyanins have low exposure in circulation, and their metabolites, such as protocatechuic acid, are more likely the actual effector molecules.⁵⁹ Future work is needed within a pharmacokinetics–metabolomics framework, combining functional experiments to validate key circuits, tissue specificity, and dose thresholds.^{60,61}

From a practical standpoint, tart cherry should be positioned as an adjunct. It reinforces lifestyle management rather than replacing first-line urate-lowering medications.^{3,11,32} In individuals who are overweight or insulin resistant, have higher purine or alcohol exposure, elevated inflammatory burden, or limited tolerability to conventional therapy, clinically perceptible marginal benefits are more likely to be observed.^{36,57,62} In patients who have already achieved targets with medication or have good dietary control, the net effect may be smaller.³ Practical recommendations include prioritizing preparations with transparent composition, labeled anthocyanin equivalents, and controllable sugar content. Capsules or low-sugar concentrates are preferable when needed.⁶³ It is important to coordinate intake with weight management, restrict purines and alcohol, ensure adequate hydration, and promote urinary alkalinization.⁶⁴ For patients with chronic kidney disease or glycemic concerns, monitoring potassium and sugar loads and regularly checking sUA and renal function is essential.^{63,65,66} Furthermore, patients using allopurinol or febuxostat should not reduce or discontinue these medications because of increasing tart cherry intake or when adding tart cherry products to the diet.^{11,12} When communicating expectations, it should be emphasized that the “average reduction is limited and inter-individual variability is pronounced” to avoid misinterpreting it as a substitute for medication.^{3,57}

This study has several notable strengths. First, it effectively integrates recent randomized evidence and conducts robustness checks, including leave-one-out analyses and bias assessments.^{23,67-69} Second, it aligns clinical observations with network pharmacology and docking results, creating a feedback loop between clinical signals and molecular

mechanisms.^{51,70} Furthermore, it differentiates between “stability in direction” and “uncertainty in magnitude” to avoid over-extrapolation in the presence of substantial statistical heterogeneity.^{69,71} However, there are also significant limitations. Most of the trials included in this study have small sample sizes and short durations, and they lack supporting dose–response and exposure–response evidence.⁷²⁻⁷⁴ Additionally, there is a lack of uniformity in formulations and dosing conventions. The reporting of anthocyanin equivalents and raw material profiles, such as high performance liquid chromatography (HPLC) polyphenol profiles, is insufficient.^{40,75,76} Moreover, inconsistencies in dietary control, concomitant medications, and adherence records may introduce noise that does not differ between groups.⁷⁷ Lastly, the reliance on database inference in network pharmacology results in a lack of direct evidence regarding the in vivo regulation of key transporters and XO in human subjects.^{47,56,60} Overall, these factors suggest that the certainty of the current evidence falls within a "limited to moderate" range.⁷⁸

Overall, tart cherry intake (including derived products) is associated with reductions in sUA, showing a generally consistent direction across most comparisons but a modest magnitude and considerable uncertainty. The effect size is markedly influenced by population characteristics, formulation differences, and intervention protocols.^{3,36,79,80} Its biological basis includes anti-inflammatory and antioxidant actions mediated by anthocyanins/polyphenols, potential inhibition of XO, and indirect modulation of urate transport processes and urinary pH.^{53,55} Clinically, it should be used as a complement to lifestyle management, in combination with a purine-restricted diet, weight management, and urine alkalinization.^{11,12,64} Future research should, on the basis of standardized dose and composition, ensuring adequate follow-up and include mechanistic endpoints to define actionable dosing, responder populations, and long-term safety.^{36,67,80,81}

Conclusion

In summary, this study integrated a meta-analysis of 4 RCTs with network pharmacology findings. Tart cherry intake (and its preparations) was associated with reductions in sUA. The leave-one-out analysis supported a consistent direction. Heterogeneity was high ($I^2 = 88\%$), and Egger’s test and the funnel plot indicated possible small study or publication bias. Anthocyanin-dominant polyphenols may regulate inflammation and metabolism via pathways involving IL6, TNF, AKT1, PPARG, and ADIPOQ, inhibit XO, and influence URAT1, GLUT9, ABCG2, and urinary pH, constituting multi-target modulation. Given the small number and heterogeneity of available RCTs, the overall certainty of evidence remains

limited. Tart cherry intake is therefore best used as a complement to lifestyle management rather than a replacement for first-line urate-lowering medications. Individuals with a higher metabolic burden may derive greater marginal benefit. Future work should include well-designed randomized studies with standardized tart cherry formulations and dosing, adequate follow-up, and mechanistic endpoints to define actionable dosing, responder populations, and long-term safety.

SUPPLEMENTARY MATERIALS

All supplementary tables and figures are available upon request from the editorial office, and are also accessible on the journal's webpage (apjcn.qdu.edu.cn).

CONFLICT OF INTEREST AND FUNDING DISCLOSURE

The authors declare no conflict of interest.

This study was supported by grants from the National Key R&D Program of China (Grant numbers: 2023YFF1104402).

REFERENCES

- 1.
1. Yang SL, Liu HM, Fang XM, Yan FM, Zhang YX. Signaling pathways in uric acid homeostasis and gout: From pathogenesis to therapeutic interventions. *Int Immunopharm.* 2024;132:doi:10.1016/j.intimp.2024.111932.
2. Ames BN, Cathcart R, Schwiers E, Hochstein P. Uric acid provides an antioxidant defense in humans against oxidant- and radical-caused aging and cancer: a hypothesis. *Proc Natl Acad Sci U S A.* 1981;78:6858-62. doi:10.1073/pnas.78.11.6858.
3. Martin KR, Coles KM. Consumption of 100% Tart Cherry Juice Reduces Serum Urate in Overweight and Obese Adults. *Current developments in nutrition.* 2019;3:nzz011-nzz. doi:10.1093/cdn/nzz011.
4. Perez-Ruiz F, Desideri G. Improving adherence to gout therapy: an expert review. *Therapeutics and Clinical Risk Management.* 2018;14:793-802. doi:10.2147/tcrm.S162956.
5. Martinon F, Pétrilli V, Mayor A, Tardivel A, Tschopp J. Gout-associated uric acid crystals activate the NALP3 inflammasome. *Nature.* 2006;440:237-41. doi:10.1038/nature04516.
6. Dehlin M, Jacobsson L, Roddy E. Global epidemiology of gout: prevalence, incidence, treatment patterns and risk factors. *Nat Rev Rheumatol.* 2020;16:380-90. doi:10.1038/s41584-020-0441-1.
7. Dalbeth N, Gosling AL, Gaffo A, Abhishek A. Gout. *Lancet.* 2021;397:1843-55. doi:10.1016/s0140-6736(21)00569-9.

8. Sattui SE, Gaffo AL. Treatment of hyperuricemia in gout: current therapeutic options, latest developments and clinical implications. *Therapeutic Advances in Musculoskeletal Disease*. 2016;8:145-59. doi:10.1177/1759720x16646703.
9. Soskind R, Abazia DT, Bridgeman MB. Updates on the treatment of gout, including a review of updated treatment guidelines and use of small molecule therapies for difficult-to-treat gout and gout flares. *Expert Opinion on Pharmacotherapy*. 2017;18:1115-25. doi:10.1080/14656566.2017.1349099.
10. Keenan RT, Schlesinger N. New and Pipeline Drugs for Gout. *Curr Rheumatol Rep*. 2016;18:32. doi:10.1007/s11926-016-0579-7.
11. FitzGerald JD, Dalbeth N, Mikuls T, Brignardello-Petersen R, Guyatt G, Abeles AM, et al. 2020 American College of Rheumatology Guideline for the Management of Gout. *Arthritis Care Res (Hoboken)*. 2020;72:744-60. doi:10.1002/acr.24180.
12. Richette P, Doherty M, Pascual E, Barskova V, Becce F, Castañeda-Sanabria J, et al. 2016 updated EULAR evidence-based recommendations for the management of gout. *Ann Rheum Dis*. 2017;76:29-42. doi:10.1136/annrheumdis-2016-209707.
13. Jacob RA, Spinozzi GM, Simon VA, Kelley DS, Prior RL, Hess-Pierce B, et al. Consumption of cherries lowers plasma urate in healthy women. *Journal of Nutrition*. 2003;133:1826-9. doi:10.1093/jn/133.6.1826.
14. Golovinskaia O, Wang CK. Review of Functional and Pharmacological Activities of Berries. *Molecules*. 2021;26:doi:10.3390/molecules26133904.
15. Martin KR, Burrell L, Bopp J. Authentic tart cherry juice reduces markers of inflammation in overweight and obese subjects: a randomized, crossover pilot study. *Food & Function*. 2018;9:5290-300. doi:10.1039/c8fo01492b.
16. Zhang YQ, Neogi T, Chen C, Chaisson C, Hunter DJ, Choi HK. Cherry Consumption and Decreased Risk of Recurrent Gout Attacks. *Arthritis and Rheumatism*. 2012;64:4004-11. doi:10.1002/art.34677.
17. Seeram NP, Momin RA, Nair MG, Bourquin LD. Cyclooxygenase inhibitory and antioxidant cyanidin glycosides in cherries and berries. *Phytomedicine*. 2001;8:362-9. doi:10.1078/0944-7113-00053.
18. Wang H, Nair MG, Strasburg GM, Chang YC, Booren AM, Gray JI, et al. Antioxidant and antiinflammatory activities of anthocyanins and their aglycon, cyanidin, from tart cherries. *Journal of Natural Products*. 1999;62:294-6. doi:10.1021/np980501m.
19. Kirakosyan A, Gutierrez E, Ramos Solano B, Seymour EM, Bolling SF. The inhibitory potential of Montmorency tart cherry on key enzymes relevant to type 2 diabetes and cardiovascular disease. *Food Chemistry*. 2018;252:142-6. doi:10.1016/j.foodchem.2018.01.084.
20. He Y, Hu YF, Jiang XW, Chen TF, Ma YT, Wu S, et al. Cyanidin-3-O-glucoside inhibits the UVB-induced ROS/COX-2 pathway in HaCaT cells. *Journal of Photochemistry and Photobiology B-Biology*. 2017;177:24-31. doi:10.1016/j.jphotobiol.2017.10.006.
21. Dehghan A, Kottgen A, Yang Q, Hwang S, Kao WL, Rivadeneira F, et al. Association of three genetic loci with uric acid concentration and risk of gout: a genome-wide association study. *Lancet*. 2008;372:1953-61. doi:10.1016/s0140-6736(08)61343-4.

22. Ichida K, Matsuo H, Takada T, Nakayama A, Murakami K, Shimizu T, et al. Decreased extra-renal urate excretion is a common cause of hyperuricemia. *Nat Commun.* 2012;3:764. doi:10.1038/ncomms1756.
23. Higgins JPT, Thompson SG, Spiegelhalter DJ. A re-evaluation of random-effects meta-analysis. *Journal of the Royal Statistical Society Series a-Statistics in Society.* 2009;172:137-59. doi:10.1111/j.1467-985X.2008.00552.x.
24. Hao DC, Xiao PG. Network pharmacology: a Rosetta Stone for traditional Chinese medicine. *Drug Dev Res.* 2014;75:299-312. doi:10.1002/ddr.21214.
25. Stamp LK, Chapman P, Frampton C, Duffull SB, Drake J, Zhang YQ, et al. Lack of effect of tart cherry concentrate dose on serum urate in people with gout. *Rheumatology.* 2020;59:2374-80. doi:10.1093/rheumatology/kez606.
26. Gonzalez DE, Kendra JA, Dickerson B, Yoo C, Ko J, McAngus K, et al. Effects of Acute and One-Week Supplementation with Montmorency Tart Cherry Powder on Food-Induced Uremic Response and Markers of Health: A Proof-of-Concept Study. *Nutrients.* 2024;16:doi:10.3390/nu16193391.
27. Wang CK, Sun WY, Dalbeth N, Wang ZJ, Wang XF, Ji XP, et al. Efficacy and safety of tart cherry supplementary citrate mixture on gout patients: a prospective, randomized, controlled study. *Arthritis Research & Therapy.* 2023;25:doi:10.1186/s13075-023-03152-1.
28. Chen PE, Liu CY, Chien WH, Chien CW, Tung TH. Effectiveness of Cherries in Reducing Uric Acid and Gout: A Systematic Review. *Evidence-Based Complementary and Alternative Medicine.* 2019;2019:doi:10.1155/2019/9896757.
29. Scanu A, Luisetto R, Ramonda R, Spinella P, Sfriso P, Galozzi P, et al. Anti-Inflammatory and Hypouricemic Effect of Bioactive Compounds: Molecular Evidence and Potential Application in the Management of Gout. *Current Issues in Molecular Biology.* 2022;44:5173-90. doi:10.3390/cimb44110352.
30. Egger M, Smith GD, Schneider M, Minder C. Bias in meta-analysis detected by a simple, graphical test. *Bmj-British Medical Journal.* 1997;315:629-34. doi:10.1136/bmj.315.7109.629.
31. Hillman AR, Uhranowsky K. Acute Ingestion of Montmorency Tart Cherry Reduces Serum Uric Acid but Has no Impact on High Sensitivity C-Reactive Protein or Oxidative Capacity. *Plant Foods for Human Nutrition.* 2021;76:83-9. doi:10.1007/s11130-021-00879-7.
32. Khanna D, Fitzgerald JD, Khanna PP, Bae S, Singh MK, Neogi T, et al. 2012 American College of Rheumatology guidelines for management of gout. Part 1: systematic nonpharmacologic and pharmacologic therapeutic approaches to hyperuricemia. *Arthritis Care Res (Hoboken).* 2012;64:1431-46. doi:10.1002/acr.21772.
33. Mulabagal V, Lang GA, Dewitt DL, Dalavoy SS, Nair MG. Anthocyanin Content, Lipid Peroxidation and Cyclooxygenase Enzyme Inhibitory Activities of Sweet and Sour Cherries. *Journal of Agricultural and Food Chemistry.* 2009;57:1239-46. doi:10.1021/jf8032039.
34. Choi HK, Curhan G. Soft drinks, fructose consumption, and the risk of gout in men: prospective cohort study. *Bmj-British Medical Journal.* 2008;336:309-+. doi:10.1136/bmj.39449.819271.BE.

35. Lamb KL, Lynn A, Russell J, Barker ME. Effect of tart cherry juice on risk of gout attacks: protocol for a randomised controlled trial. *Bmj Open*. 2020;10:doi:10.1136/bmjopen-2019-035108.
36. Kelley DS, Adkins Y, Laugero KD. A Review of the Health Benefits of Cherries. *Nutrients*. 2018;10:doi:10.3390/nu10030368.
37. Choi HK, Ford ES. Prevalence of the metabolic syndrome in individuals with hyperuricemia. *American Journal of Medicine*. 2007;120:442-7. doi:10.1016/j.amjmed.2006.06.040.
38. Dehghan A, van Hoek M, Sijbrands EJG, Hofman A, Witteman JCM. High Serum Uric Acid as a Novel Risk Factor for Type 2 Diabetes. *Diabetes Care*. 2008;31:361-2. doi:10.2337/dc07-1276.
39. Becker MA, Schumacher HRJ, Wortmann RL, MacDonald PA, Eustace D, Palo WA, et al. Febuxostat compared with allopurinol in patients with hyperuricemia and gout. *New England Journal of Medicine*. 2005;353:2450-61. doi:10.1056/NEJMoa050373.
40. Kirakosyan A, Seymour EM, Llanes DEU, Kaufman PB, Bolling SF. Chemical profile and antioxidant capacities of tart cherry products. *Food Chemistry*. 2009;115:20-5. doi:10.1016/j.foodchem.2008.11.042.
41. Atkinson G, Batterham AM. True and false interindividual differences in the physiological response to an intervention. *Experimental Physiology*. 2015;100:577-88. doi:10.1113/ep085070.
42. Johnson RJ, Segal MS, Sautin YS, Nakagawa T, Feig DI, Kang D, et al. Potential role of sugar (fructose) in the epidemic of hypertension, obesity and the metabolic syndrome, diabetes, kidney disease, and cardiovascular disease. *Am J Clin Nutr*. 2007;86:899-906. doi:10.1093/ajcn/86.4.899.
43. Sakhaee K, Nicar M, Hill K, Pak CY. Contrasting effects of potassium citrate and sodium citrate therapies on urinary chemistries and crystallization of stone-forming salts. *Kidney Int*. 1983;24:348-52. doi:10.1038/ki.1983.165.
44. Ettinger B, Pak CY, Citron JT, Thomas C, AdamsHuet B, Vangessel A. Potassium-magnesium citrate is an effective prophylaxis against recurrent calcium oxalate nephrolithiasis. *Journal of Urology*. 1997;158:2069-73. doi:10.1016/s0022-5347(01)68155-2.
45. Shoag J, Tasian GE, Goldfarb DS, Eisner BH. The new epidemiology of nephrolithiasis. *Advances in Chronic Kidney Disease*. 2015;22:273-8. doi:10.1053/j.ackd.2015.04.004.
46. Sterne JAC, Sutton AJ, Ioannidis JPA, Terrin N, Jones DR, Lau J, et al. Recommendations for examining and interpreting funnel plot asymmetry in meta-analyses of randomised controlled trials. *Bmj-British Medical Journal*. 2011;343:doi:10.1136/bmj.d4002.
47. Zhou YY, Zhou B, Pache L, Chang M, Khodabakhshi AH, Tanaseichuk O, et al. Metascape provides a biologist-oriented resource for the analysis of systems-level datasets. *Nature Communications*. 2019;10:doi:10.1038/s41467-019-09234-6.
48. Kibble M, Saarinen N, Tang J, Wennerberg K, Mäkelä S, Aittokallio T. Network pharmacology applications to map the unexplored target space and therapeutic potential of natural products. *Nat Prod Rep*. 2015;32:1249-66. doi:10.1039/c5np00005j.

49. Szklarczyk D, Gable AL, Nastou KC, Lyon D, Kirsch R, Pyysalo S, et al. The STRING database in 2021: customizable protein-protein networks, and functional characterization of user-uploaded gene/measurement sets. *Nucleic Acids Res.* 2021;49:D605-d12. doi:10.1093/nar/gkaa1074.
50. Stelzer G, Rosen N, Plaschkes I, Zimmerman S, Twik M, Fishilevich S, et al. The GeneCards Suite: From Gene Data Mining to Disease Genome Sequence Analyses. *Curr Protoc Bioinformatics.* 2016;54:1.30.1-1..3. doi:10.1002/cpbi.5.
51. Eberhardt J, Santos-Martins D, Tillack AF, Forli S. AutoDock Vina 1.2.0: New Docking Methods, Expanded Force Field, and Python Bindings. *J Chem Inf Model.* 2021;61:3891-8. doi:10.1021/acs.jcim.1c00203.
52. Pagadala NS, Syed K, Tuszynski J. Software for molecular docking: a review. *Biophys Rev.* 2017;9:91-102. doi:10.1007/s12551-016-0247-1.
53. Kelley EE. A new paradigm for XOR-catalyzed reactive species generation in the endothelium. *Pharmacol Rep.* 2015;67:669-74. doi:10.1016/j.pharep.2015.05.004.
54. He Y, Hara H, Núñez G. Mechanism and Regulation of NLRP3 Inflammasome Activation. *Trends Biochem Sci.* 2016;41:1012-21. doi:10.1016/j.tibs.2016.09.002.
55. Nakayama A, Matsuo H, Takada T, Ichida K, Nakamura T, Ikebuchi Y, et al. ABCG2 is a high-capacity urate transporter and its genetic impairment increases serum uric acid levels in humans. *Nucleosides Nucleotides Nucleic Acids.* 2011;30:1091-7. doi:10.1080/15257770.2011.633953.
56. Bobulescu IA, Moe OW. Renal transport of uric acid: evolving concepts and uncertainties. *Adv Chronic Kidney Dis.* 2012;19:358-71. doi:10.1053/j.ackd.2012.07.009.
57. Kelley DS, Adkins Y, Reddy A, Woodhouse LR, Mackey BE, Erickson KL. Sweet Bing Cherries Lower Circulating Concentrations of Markers for Chronic Inflammatory Diseases in Healthy Humans. *Journal of Nutrition.* 2013;143:340-4. doi:10.3945/jn.112.171371.
58. Khoo HE, Azlan A, Tang ST, Lim SM. Anthocyanidins and anthocyanins: colored pigments as food, pharmaceutical ingredients, and the potential health benefits. *Food Nutr Res.* 2017;61:1361779. doi:10.1080/16546628.2017.1361779.
59. Czank C, Cassidy A, Zhang QZ, Morrison DJ, Preston T, Kroon PA, et al. Human metabolism and elimination of the anthocyanin, cyanidin-3-glucoside: a (13)C-tracer study. *Am J Clin Nutr.* 2013;97:995-1003. doi:10.3945/ajcn.112.049247.
60. Wishart DS. Metabolomics for Investigating Physiological and Pathophysiological Processes. *Physiol Rev.* 2019;99:1819-75. doi:10.1152/physrev.00035.2018.
61. Rinschen MM, Ivanisevic J, Giera M, Siuzdak G. Identification of bioactive metabolites using activity metabolomics. *Nat Rev Mol Cell Biol.* 2019;20:353-67. doi:10.1038/s41580-019-0108-4.
62. Keane KM, George TW, Constantinou CL, Brown MA, Clifford T, Howatson G. Effects of Montmorency tart cherry (*Prunus Cerasus L.*) consumption on vascular function in men with early hypertension. *American Journal of Clinical Nutrition.* 2016;103:1531-9. doi:10.3945/ajcn.115.123869.

63. Te Morenga L, Mallard S, Mann J. Dietary sugars and body weight: systematic review and meta-analyses of randomised controlled trials and cohort studies. *Bmj*. 2012;346:e7492. doi:10.1136/bmj.e7492.
64. Kanbara A, Miura Y, Hyogo H, Chayama K, Seyama I. Effect of urine pH changed by dietary intervention on uric acid clearance mechanism of pH-dependent excretion of urinary uric acid. *Nutr J*. 2012;11:39. doi:10.1186/1475-2891-11-39.
65. Shlipak MG, Tummalapalli SL, Boulware LE, Grams ME, Ix JH, Jha V, et al. The case for early identification and intervention of chronic kidney disease: conclusions from a Kidney Disease: Improving Global Outcomes (KDIGO) Controversies Conference. *Kidney Int*. 2021;99:34-47. doi:10.1016/j.kint.2020.10.012.
66. Cheungpasitporn W, Thongprayoon C, O'Corragain OA, Edmonds PJ, Kittanamongkolchai W, Erickson SB. Associations of sugar-sweetened and artificially sweetened soda with chronic kidney disease: a systematic review and meta-analysis. *Nephrology (Carlton)*. 2014;19:791-7. doi:10.1111/nep.12343.
67. Page MJ, McKenzie JE, Bossuyt PM, Boutron I, Hoffmann TC, Mulrow CD, et al. The PRISMA 2020 statement: an updated guideline for reporting systematic reviews. *Bmj*. 2021;372:n71. doi:10.1136/bmj.n71.
68. Cumpston M, Li T, Page MJ, Chandler J, Welch VA, Higgins JP, et al. Updated guidance for trusted systematic reviews: a new edition of the Cochrane Handbook for Systematic Reviews of Interventions. *Cochrane Database Syst Rev*. 2019;10:Ed000142. doi:10.1002/14651858.Ed000142.
69. Sterne JAC, Savović J, Page MJ, Elbers RG, Blencowe NS, Boutron I, et al. RoB 2: a revised tool for assessing risk of bias in randomised trials. *Bmj*. 2019;366:l4898. doi:10.1136/bmj.l4898.
70. Zhang RZ, Zhu X, Bai H, Ning K. Network Pharmacology Databases for Traditional Chinese Medicine: Review and Assessment. *Front Pharmacol*. 2019;10:123. doi:10.3389/fphar.2019.00123.
71. IntHout J, Ioannidis JPA, Borm GF. The Hartung-Knapp-Sidik-Jonkman method for random effects meta-analysis is straightforward and considerably outperforms the standard DerSimonian-Laird method. *BMC Med Res Methodol*. 2014;14:25. doi:10.1186/1471-2288-14-25.
72. Dechartres A, Trinquart L, Boutron I, Ravaud P. Influence of trial sample size on treatment effect estimates: meta-epidemiological study. *Bmj*. 2013;346:f2304. doi:10.1136/bmj.f2304.
73. IntHout J, Ioannidis JP, Rovers MM, Goeman JJ. Plea for routinely presenting prediction intervals in meta-analysis. *BMJ Open*. 2016;6:e010247. doi:10.1136/bmjopen-2015-010247.
74. Greenland S, Longnecker MP. Methods for trend estimation from summarized dose-response data, with applications to meta-analysis. *Am J Epidemiol*. 1992;135:1301-9. doi:10.1093/oxfordjournals.aje.a116237.
75. Manach C, Williamson G, Morand C, Scalbert A, Rémésy C. Bioavailability and bioefficacy of polyphenols in humans. I. Review of 97 bioavailability studies. *Am J Clin Nutr*. 2005;81:230s-42s. doi:10.1093/ajcn/81.1.230S.

76. Rothwell JA, Perez-Jimenez J, Neveu V, Medina-Remón A, M'Hiri N, García-Lobato P, et al. Phenol-Explorer 3.0: a major update of the Phenol-Explorer database to incorporate data on the effects of food processing on polyphenol content. *Database* (Oxford). 2013;2013:bat070. doi:10.1093/database/bat070.
77. Mirmiran P, Bahadoran Z, Gaeini Z. Common Limitations and Challenges of Dietary Clinical Trials for Translation into Clinical Practices. *International Journal of Endocrinology and Metabolism*. 2021;19:doi:10.5812/ijem.108170.
78. Murad MH, Mustafa RA, Schünemann HJ, Sultan S, Santesso N. Rating the certainty in evidence in the absence of a single estimate of effect. *Evid Based Med*. 2017;22:85-7. doi:10.1136/ebmed-2017-110668.
79. Keane KM, George TW, Constantinou CL, Brown MA, Clifford T, Howatson G. Effects of Montmorency tart cherry (*Prunus Cerasus* L.) consumption on vascular function in men with early hypertension. *Am J Clin Nutr*. 2016;103:1531-9. doi:10.3945/ajcn.115.123869.
80. Hill JA, Keane KM, Quinlan R, Howatson G. Tart Cherry Supplementation and Recovery From Strenuous Exercise: A Systematic Review and Meta-Analysis. *Int J Sport Nutr Exerc Metab*. 2021;31:154-67. doi:10.1123/ijsnem.2020-0145.
81. Xu C, Zhang YG, Han FF, Niu YM, Kuang XY, Zhang C. How to perform dose-response meta-analysis: A brief Introduction of methodology. *Chinese Journal of Evidence-Based Medicine*. 2015;15:1236-40. doi:10.7507/1672-2531.20150204.

Table 1. RoB 2 assessment for included RCTs

RCT Author Name Tear [Ref.]	Bias					
	Selection	Performance	Detection	Attrition	Reporting	Other
Gonzalez, 2024	+	+	+	+	+	+
Martin, 2019	+	+	+	+	+	+
Stamp, 2020	+	+	?	+	+	+
Wang, 2022	+	+	+	?	+	+

Risk of bias levels; low RoB (+), high RoB (-), and unknown (?).

Not Proof Read

Table 2. GRADE certainty of prespecified outcomes in RCTs

	Num. of studies	Certainty assessment						Num of patients	
		Study design	Risk of bias	Inconsistency	Indirectness	Imprecision	Other considerations	Walnut	Control
Stamp, 2020 (7.5mL)	1	randomised trials	not serious	not serious	not serious	serious ^a	none	9	9
Stamp, 2020 (15 mL)	1	randomised trials	not serious	not serious	not serious	serious ^b	none	9	9
Stamp, 2020 (22.5 mL)	1	randomised trials	not serious	not serious	not serious	serious ^c	none	9	9
Stamp, 2020 (30 mL)	1	randomised trials	not serious	not serious	serious ^d	serious ^e	none	9	9
Gonzalez, 2024 (acute)	1	randomised trials	not serious	not serious	not serious	not serious	none	25	25
Gonzalez, 2024 (A week)	1	randomised trials	not serious	not serious	not serious	not serious	none	25	25
Wang, 2022	1	randomised trials	not serious	not serious	not serious	not serious	none	26	26
Martin,2019	1	randomised trials	not serious	not serious	not serious	not serious	none	82	86

	Num. of studies	Effect		Certainty	Importance
		Relative (95% CI)	Absolute (95% CI)		
Stamp, 2020 (7.5mL)	1	-	SMD 0.32 lower (1.25 lower to 0.61 higher)	⊕⊕⊕○ Moderate ^a	
Stamp, 2020 (15 mL)	1	-	SMD 0.35 SD higher (0.59 lower to 1.28 higher)	⊕⊕⊕○ Moderate ^b	
Stamp, 2020 (22.5 mL)	1	-	SMD 0.09 SD lower (1.01 lower to 0.84 higher)	⊕⊕⊕○ Moderate ^c	
Stamp, 2020 (30 mL)	1	-	SMD 0.1 lower (1.03 lower to 0.82 higher)	⊕⊕○○ Low ^{d,e}	
Gonzalez, 2024 (acute)	1	-	SMD 0.18 lower (0.74 lower to 0.37 higher)	⊕⊕⊕⊕ High	
Gonzalez, 2024 (A week)	1	-	SMD 0.14 lower (0.69 lower to 0.42 higher)	⊕⊕⊕⊕ High	
Wang, 2022	1	-	SMD 3.63 lower (4.54 lower to 2.72 lower)	⊕⊕⊕⊕ High	
Martin,2019	1	-	SMD 0.05 higher (0.25 lower to 0.35 higher)	⊕⊕⊕⊕ High	

[†]Items involving the 7.5 mL dose had a relatively small sample size and differences in within-group variation compared with the other doses. Together, these factors reduce the precision of the effect estimates, and confidence intervals can be wide; caution is needed in interpreting the direction and magnitude of the overall effect.

[‡]If some comparisons involve both small-sample and moderate-dose groups, there is an increased risk of random error that may amplify or attenuate the observed effect and reduce the stability and reproducibility of the results. Consider inclusion in sensitivity analyses or subgroup analyses.

[§]In the presence of significant dose differences and small sample sizes, additional indirect effects may occur when extrapolated to unstudied dose intervals, thereby reducing direct applicability to the target population. Define this clearly in the discussion and limit clinical decision making to stratified dose-subgroup analyses if necessary.

[¶]The high-dose group may carry a higher indirect risk if the study's dose interval does not overlap with the target interval. Interpret the dose-response relationship with caution; perform dose-group independent analyses as needed; report the strength of evidence for each subgroup.

^{††}Highest-dose groups tend to have minimal sample sizes; random fluctuations may be large. Present separate effects and CIs for the highest-dose subgroup and emphasize strength of evidence and potential bias in the discussion.

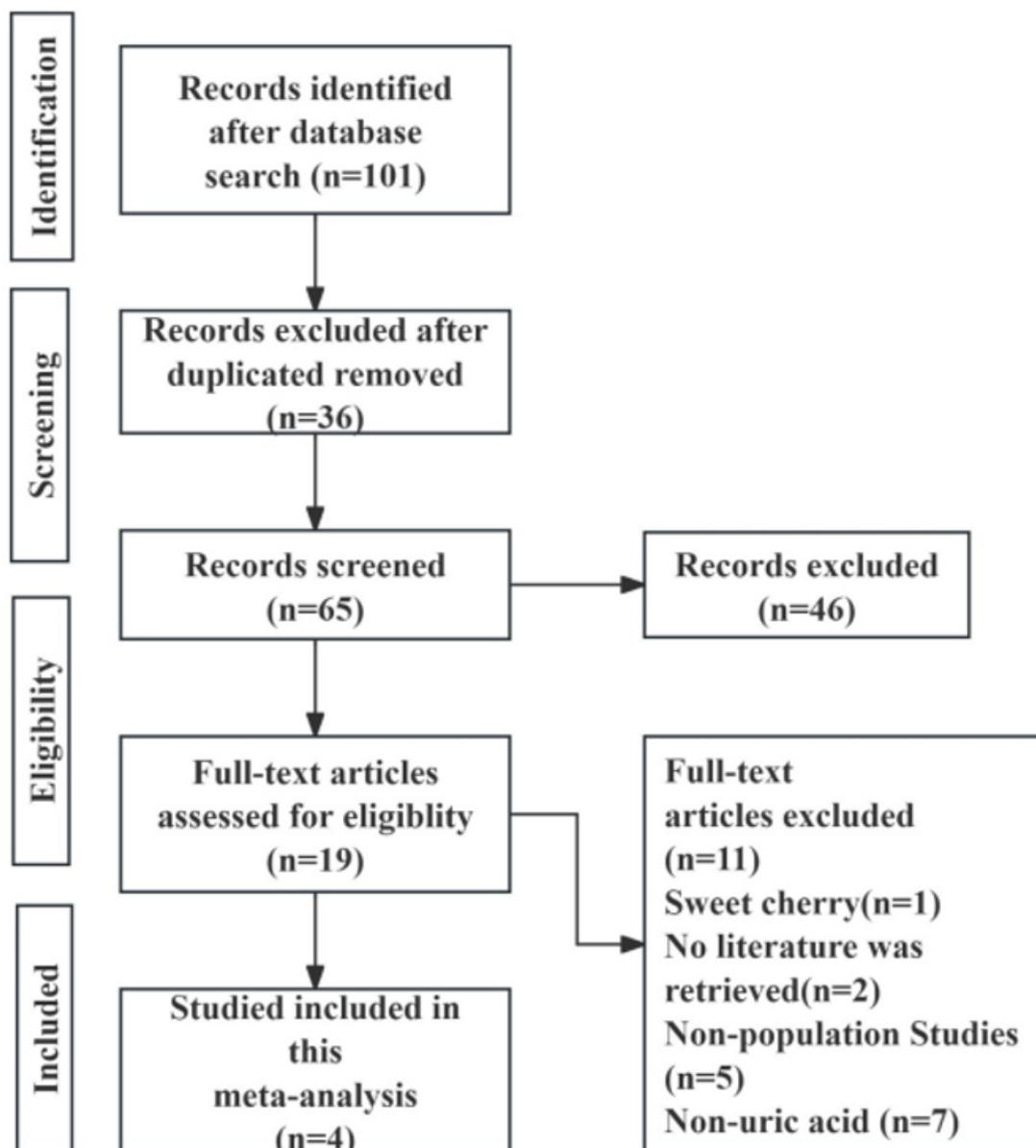


Figure 1. PRISMA flow diagram of study identification, screening, eligibility, and inclusion.

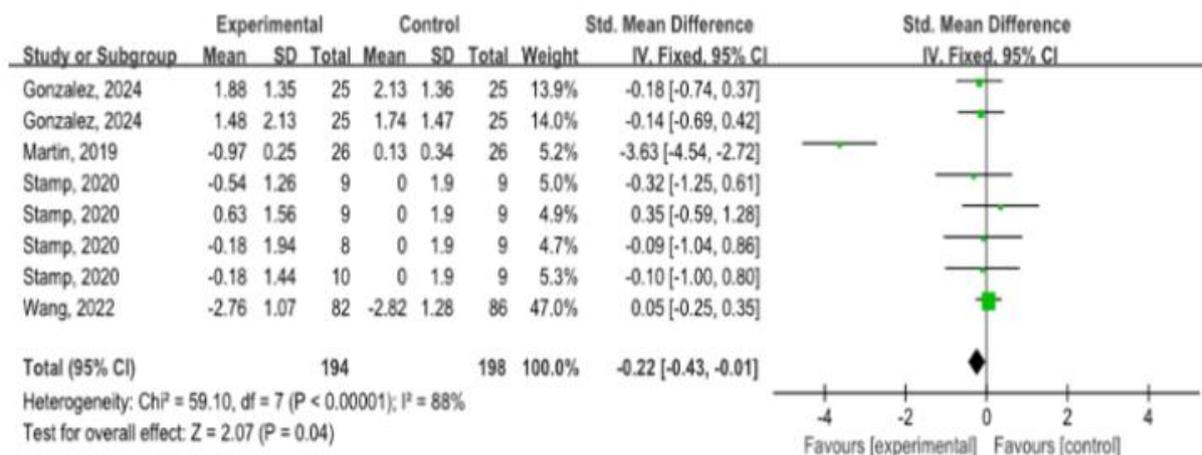


Figure 2. Forest plot of the effect of tart cherry intake on sUA levels

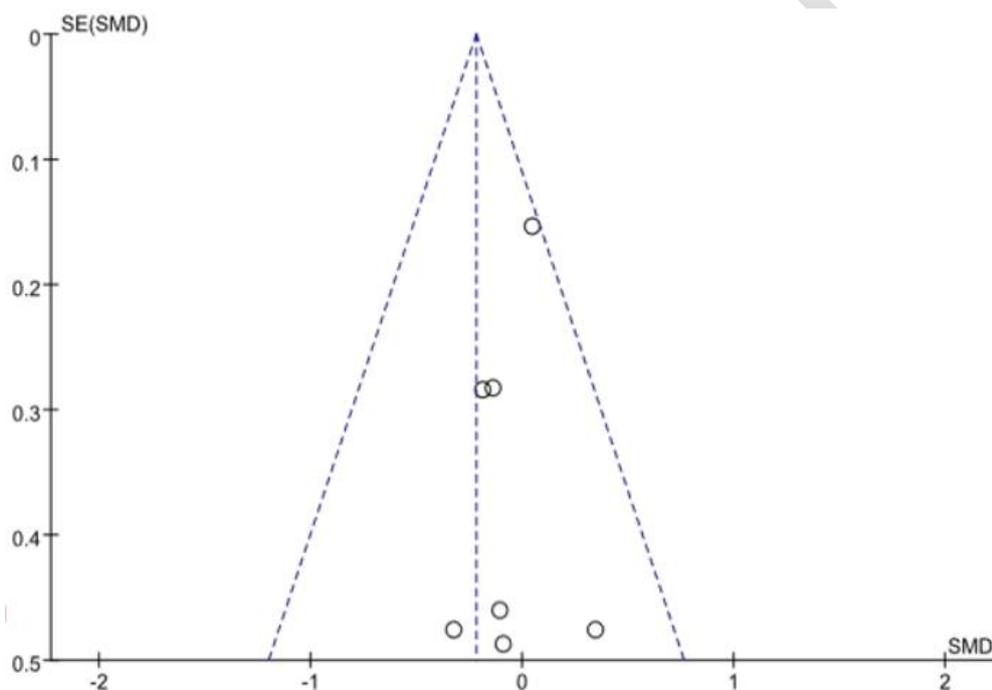


Figure 3. Funnel plot of SMD for serum uric acid levels

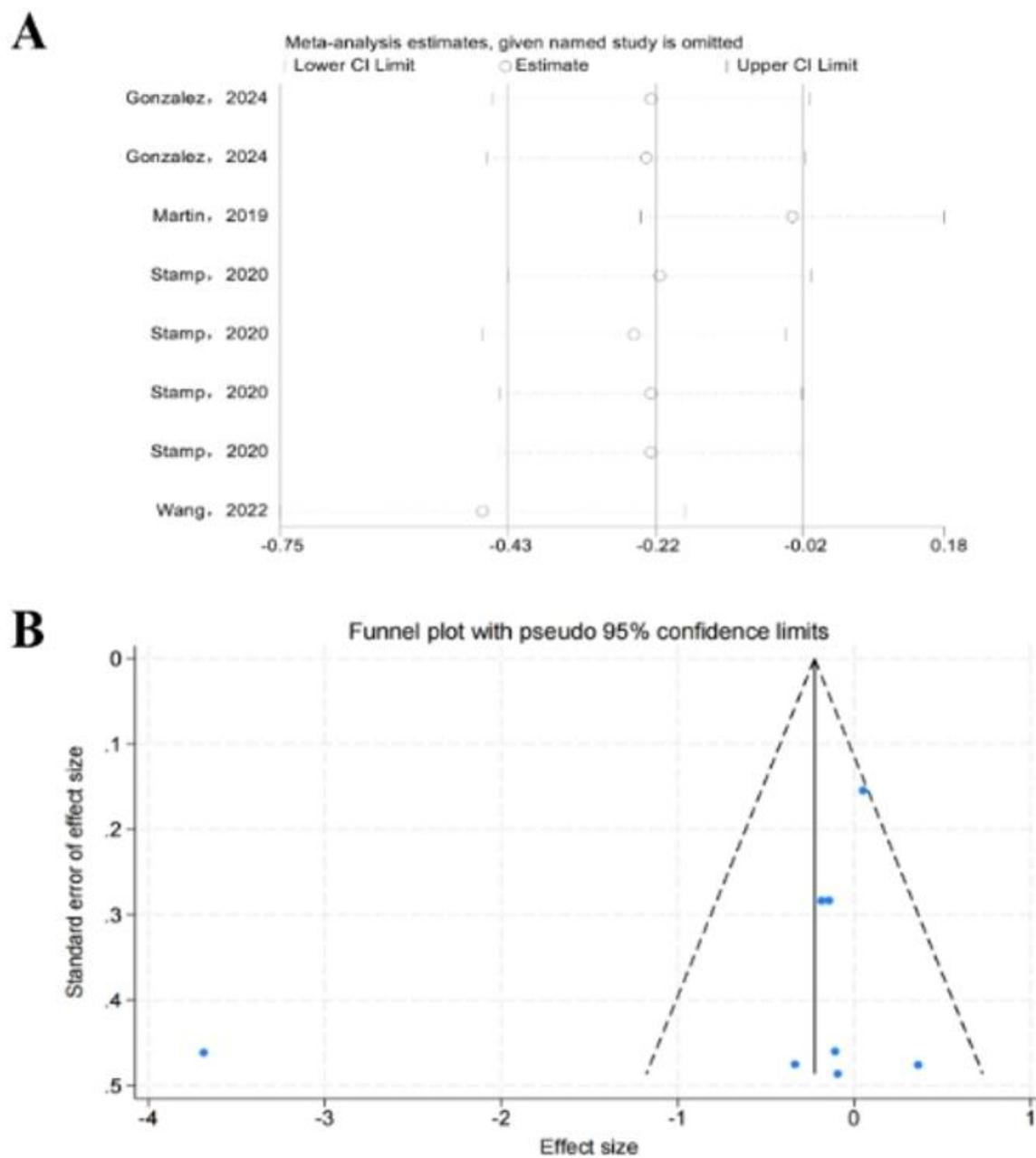


Figure 4. Sensitivity analysis and publication bias assessment. (A) Sensitivity analysis was performed by excluding studies one by one, (B) Funnel plots and their pseudo 95% confidence limits

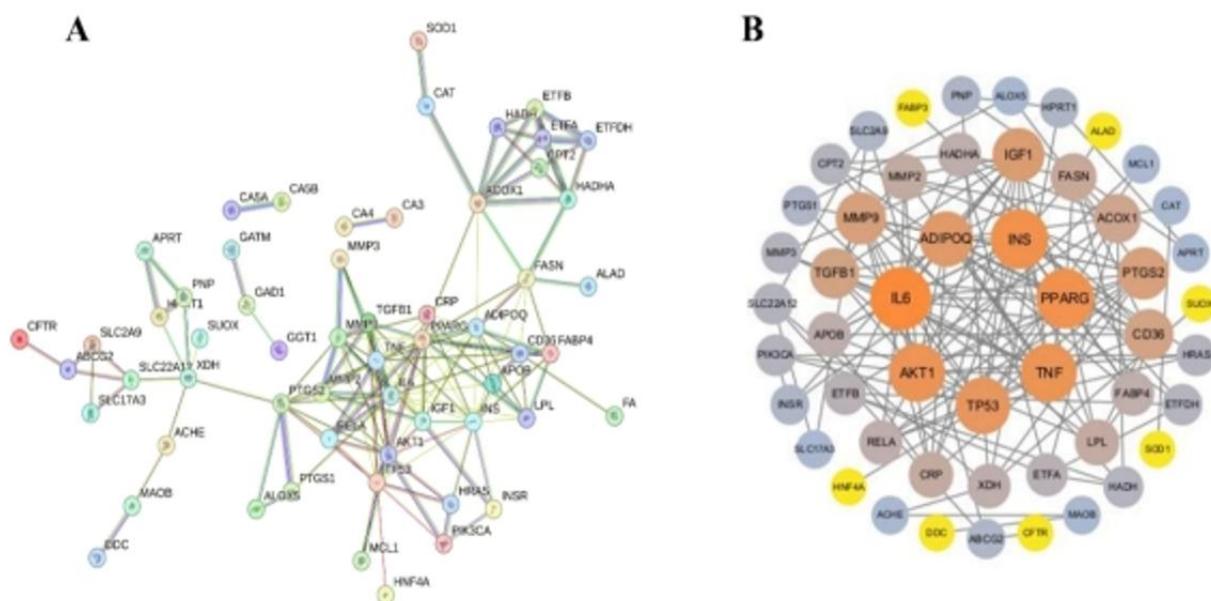


Figure 5. Network pharmacology of tart cherry for sUA reduction: (A) PPI network (STRING); (B) Hub genes (Cytoscape/CytoHubba)

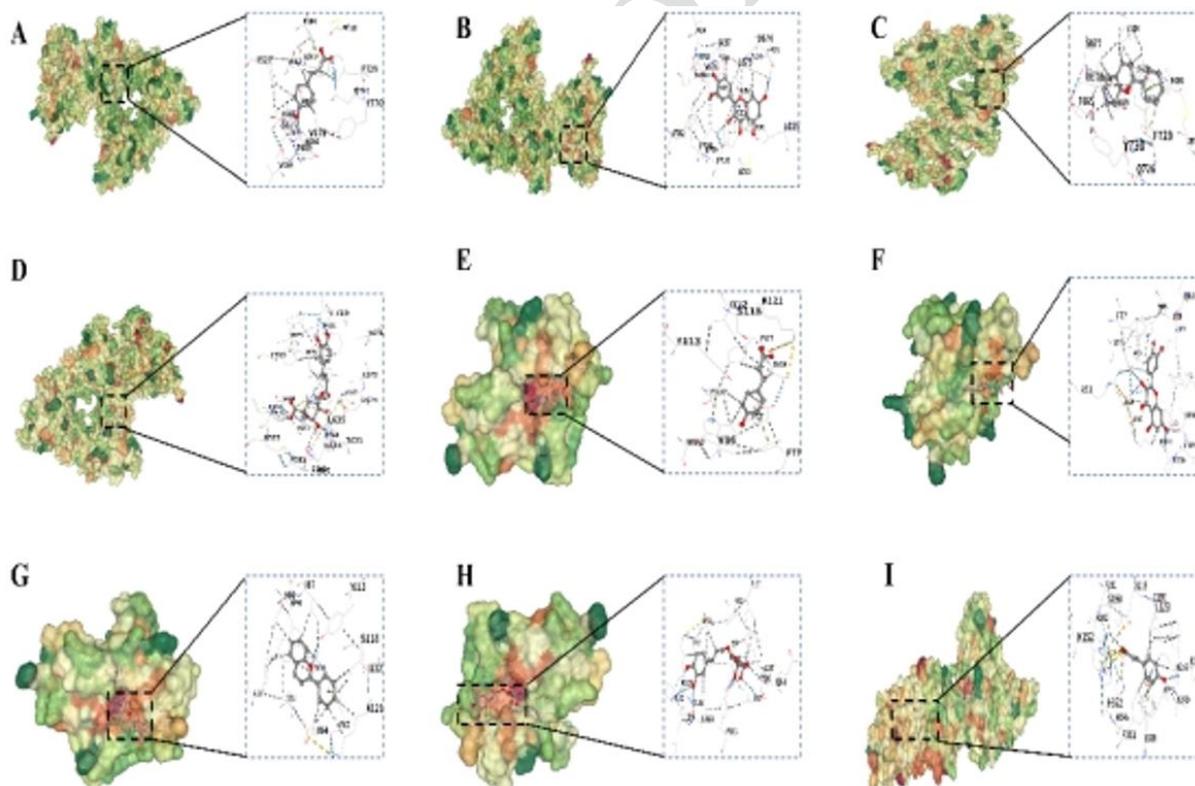


Figure 6. Molecular docking results. IL6 and anthocyanin (A), IL6 and chlorogenic acid (B), IL6 and p-coumaric acid (C), IL6 and quercetin (D), AKT1 and anthocyanin (E), AKT1 and chlorogenic acid (F), AKT1 and p-coumaric acid (G), AKT1 and quercetin (H), ADIPOQ and p-coumaric acid (I). The remaining docking results are presented in the Supplementary material

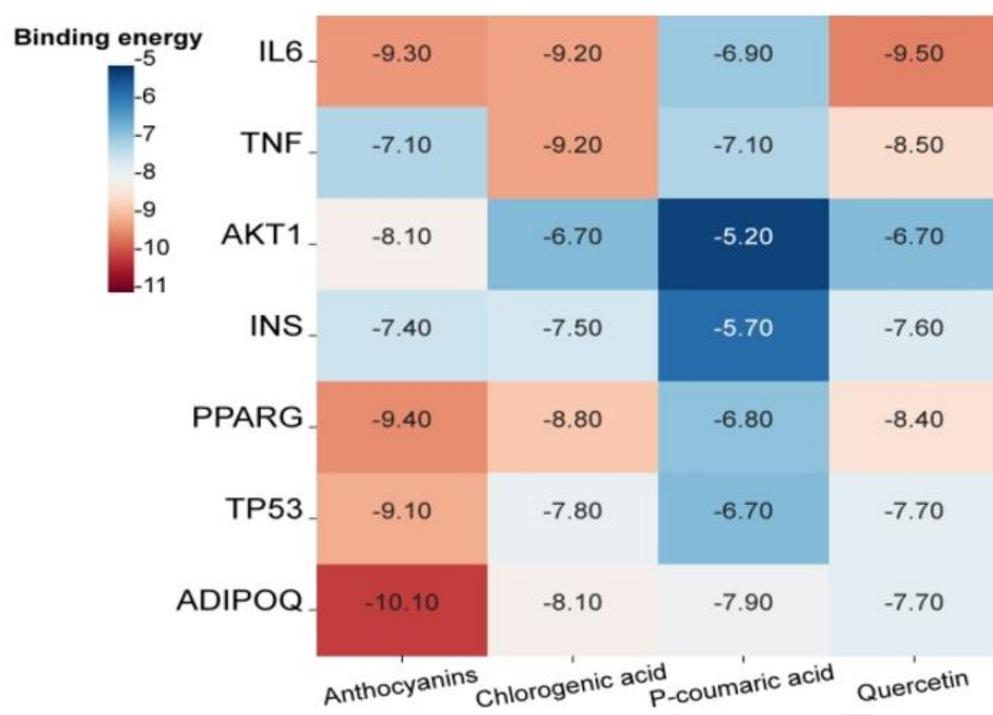


Figure 7. Binding-energy heatmap of tart cherry constituents and target proteins