

Original Article

Dietary, metabolic and gut microbiota influences on primary ovarian failure: a two-sample Mendelian randomization study

Xueying Liu MD^{1,2†}, Zhongliang Lin MD^{1,2†}, Kejing Zhu MD^{1,2}, Renke He MD^{1,2}, Zhaoying Jiang MD^{1,2}, Haiyan Wu MD², Jiaen Yu MD², Qinyu Luo MD², Jianzhong Sheng MD^{1,2,3}, Jiexue Pan MD^{3,4,5}, Hefeng Huang MD^{1,2,3,4,5}

¹Department of Obstetrics and Gynecology, Center for Reproductive Medicine, the Fourth Affiliated Hospital of School of Medicine, and International School of Medicine, International Institutes of Medicine, Zhejiang University, Yiwu, China

²Key Laboratory of Reproductive Genetics, Department of Reproductive Endocrinology, Women's Hospital, Zhejiang University School of Medicine, Hangzhou, China

³Research Units of Embryo Original Diseases, Chinese Academy of Medical Sciences, Shanghai, China

⁴Shanghai Key Laboratory of Reproduction and Development, Shanghai, China

⁵Obstetrics and Gynecology Hospital, Institute of Reproduction and Development, Fudan University, Shanghai, China

†Both authors contributed equally to this manuscript

Background and Objectives: Previous studies have reported there were associations between ovarian function and dietary factors, metabolic factors and gut microbiota. However, it is unclear whether causal associations exist. We aimed to explore the causal relationship of these factors with risk of primary ovarian failure (POF). **Methods and Study Design:** Two-sample Mendelian randomization (MR) analysis was performed to genetically predict the causal effects of dietary and metabolic factors and gut microbiota on POF. The inverse variance weighted (IVW) method was used as the primary statistical method. A series of sensitivity analyses, including weighted median, MR-Egger, simple mode, weighted mode methods, and leave-one-out analysis, were conducted to assess the robustness of the MR analysis results. **Results:** IVW analysis revealed that cigarettes smoked per day, coffee intake and cooked vegetable intake were not causally correlated with POF at the genetic level. However, POF were associated with fresh fruit intake, BMI, *Eubacterium (hallii group)*, *Eubacterium (ventriosum group)*, *Adlercreutzia*, *Intestinibacter*, *Lachnospiraceae (UCG008)*, and *Terrisporobacter*. These findings were robust according to extensive sensitivity analyses. **Conclusions:** This study identified several dietary factors, metabolic factors and gut microbiota taxa that may be causally implicated in POF, potentially offering new therapeutic targets.

Key Words: primary ovarian failure, dietary factors, metabolic factors, gut microbiota, Mendelian randomization

INTRODUCTION

The ovary is essential for establishing and maintaining secondary sexual characteristics and fertility in females. However, primary ovarian failure (POF) negatively influences reproductive health and induces disorders of ovarian function. POF is defined as the presence of postmenopausal levels of follicle-stimulating hormone (FSH) (> 40 IU/L) in woman under 40 years of age, accompanied by four or more months of secondary amenorrhea, indicating the exhaustion of the ovarian reserve before age 40. In addition, women with POF experience menopausal symptoms and are adversely affected by long-term estrogen deprivation, which significantly affects their physical and mental health.¹ Given that the chance of spontaneous conception is 5%-10%,² adoption or *in vitro* fertilization and embryo transfer using donor oocytes are considered

effective fertility treatments for women with POF. The etiology of POF is heterogeneous, including genetic defects, autoimmune diseases, iatrogenic factors (radiotherapy, chemotherapy, and ovarian surgery), and environmental factors.³ However, most patients are idiopathic an-

Corresponding Author: Prof Hefeng Huang, Department of Obstetrics and Gynecology, Center for Reproductive Medicine, the Fourth Affiliated Hospital of School of Medicine, and International School of Medicine, International Institutes of Medicine, Zhejiang University, Yiwu, China. 1575 Chouzhou North Rd., Yiwu, Zhejiang, China, 322000

Tel: +86-13906526300

Email: hhf57@zju.edu.cn

Manuscript received 16 June 2024. Initial review completed 18 June 2024. Revision accepted 13 August 2024.

doi: 10.6133/apjcn.202502_34(1).0005

the cause is unclear. Compared with immobile etiologies such as genetic and iatrogenic factors, learning and understanding the influence of modifiable factors such as diet, metabolic traits and gut microbiota in POF seem more valuable for the prevention and treatment of this disease. The most established and well-learned dietary factor associated with POF is smoking, while caffeine intake has been suggested as a potential factor.⁴ Several observational studies have suggested that smoking duration,⁵⁻⁸ caffeine consumption,⁹⁻¹¹ and fruit intake^{12,13} were associated with the age of menopause. In addition, the gastrointestinal tract, which hosts ten trillion diverse symbionts (50 bacterial phyla and approximately 100–1000 bacterial species), has been extensively studied owing to its basic functions in the immunological, metabolic, structural and neurological landscapes in humans.¹⁴ The interactions of the gut microbiota with estrogen, androgens, insulin, and other hormones appear to be crucial for the reproductive endocrine system.¹⁵ Imbalance of the gut microbiota composition can lead to polycystic ovary syndrome (PCOS),¹⁶⁻¹⁸ endometriosis,^{19,20} ovarian dysfunction,²¹ and ovarian cancer.²² However, less is known about the exact role of diet and gut microbiota in ovarian physiology, and few studies have explored the causal relationship between the gut microbiota and POF.²³

Although randomized controlled trials (RCTs) are the gold standard for establishing causal relationships, they can be costly, time-consuming and even impractical.²⁴ On

the other hand, observational studies may not robustly reflect causal relationships owing to many potential biases, confounders and reverse causation.²⁵ Mendelian randomization (MR) is an approach that uses genetic variants associated with an exposure as instrumental variables (IVs) to examine the causality of exposure–outcome associations. MR can minimize potential confounders and reverse causality because genetic variants segregate randomly and independently, preceding the outcome of interest.²⁴ Furthermore, during the last decade, the publication of a large volume of genome-wide association studies (GWASs) has enabled MR studies to be conducted without recruiting new patients. Therefore, MR provides a suitable method for inferring the causal effect between risk factors and POF. Here, we conducted an MR study to investigate the associations between dietary and metabolic factors and gut microbiota with the risk of POF.

METHODS

We assessed the causal links between lifestyle-related exposure factors and POF using two-sample MR. An overview of the analytical approach is shown in Figure 1A.

Exposure data

Diet-related exposure factors used in this study included cigarettes smoked per day, coffee intake, fresh fruit intake, and cooked vegetable intake. Metabolism-related

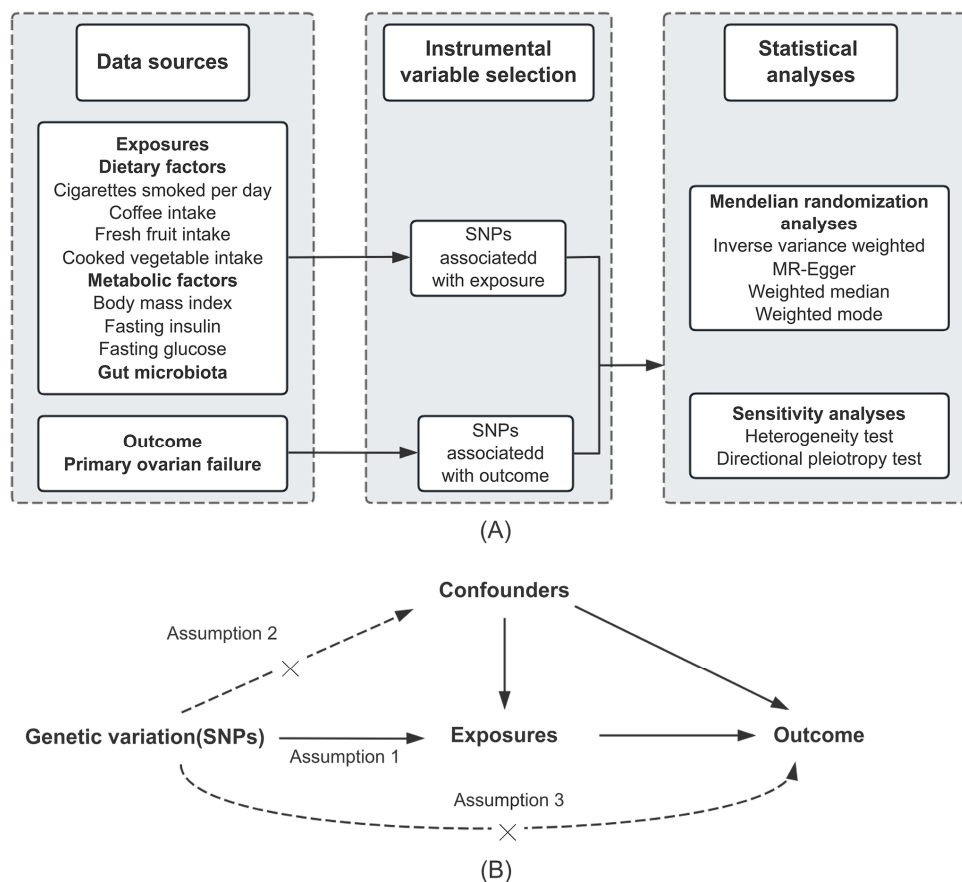


Figure 1. Study design (A) Flowchart showing the process for the MR analyses, including data collection, IVs selection, and statistical analysis. (B) Directed acyclic graph showing the assumptions of the MR methodology. MR relies on three assumptions: the genetic variants selected as instruments must (1) be associated with the exposures, (2) not be associated with confounders, (3) not directly affect the outcome, except through their effect on the exposures. SNP, single-nucleotide polymorphisms.

exposure factors used in this study included body mass index (BMI), fasting insulin, and fasting glucose. These GWAS summary-level data were extracted from IEU open GWAS project. We obtained genetic variant information related to the human gut microbiome composition from the latest large-scale genome-wide meta-analysis conducted by the MiBioGen consortium (<https://mibiogen.gcc.rug.nl/>) based on European-dominated participants.²⁶ This study analyzed genome-wide genotypes and 16S fecal microbiome data from 18,340 individuals from 24 cohorts. Accordingly, the genus level was the lowest. A total of 131 genera with a mean abundance greater than 1% were identified, 12 of which were unknown genera.²⁶ As a result, we included 119 genus-level taxa in this study. More information about the exposure datasets is presented in Table 1.

Outcome data

GWAS summary statistics related to POF were obtained from the FinnGen Consortium release data (<https://www.r8.finnngen.fi/>), which is one of the largest nationwide genetic studies with access to comprehensive electronic health register data of participants. Detailed information on used exposure datasets is presented in Table 1.

Instrumental variable selection

Single-nucleotide polymorphisms (SNPs) are used as IVs in MR analysis to provide evidence of causality between the exposure and outcome. To ensure the accuracy and robustness of the causal links, SNPs must satisfy three core assumptions to be used as IVs (Figure 1B).²⁷ Therefore, we selected independent SNPs (linkage disequilibrium $R^2 < 0.001$ and clumping distance=10,000 kb, based on the European-based 1000 Genome Projects reference panel) associated with each exposure factor at a genome-

wide threshold of significance ($p < 5 \times 10^{-8}$, diet-related and metabolism-related exposure factors) or at a locus-wide threshold of significance ($p < 1 \times 10^{-5}$, gut microbiome-related exposure factors).

Statistical analysis

The inverse variance weighted (IVW) method was used as the primary statistical method and can provide the most accurate causal estimates provided that the pleiotropic effect is balanced and that all IVs meet the MR assumptions.²⁸ Given the difficulty in verifying that IVs influence the outcome solely through the exposure of interest, we performed a series of sensitivity analyses under various assumptions. These were designed to assess the robustness of the associations and to examine horizontal pleiotropy for exposures, using methods such as weighted median, MR-Egger, simple mode, and weighted mode. The weighted median of SNP-specific estimates provides valid results when more than 50% of the information is contributed by IVs.²⁹ MR-Egger regression provides a valid estimate of causal estimates under the instrument strength independent of direct effect (InSIDE) assumption.³⁰ However, this approach was used to detect and adjust for unbalanced horizontal pleiotropy rather than to produce causal estimates due to the low statistical power of MR-Egger. A MR-Egger intercept significantly different from 0 ($p < 0.05$) indicates the occurrence of directional pleiotropy and a potentially biased IVW estimate. To further test the robustness of our results, Cochran's Q test was used to evaluate heterogeneity among the SNPs included in each analysis. Q statistics significant at $p < 0.05$ provide evidence for heterogeneity between individual genetic variants and the existence of invalid instruments.³¹ In addition, leave-one-out analysis was performed to assess whether an outcome was driven by a single outlying SNP,³² indicating the presence of hetero-

Table 1. Information of the exposures and outcome datasets

Exposure or outcome	IEU GWAS id	Consortium	Cases	Controls	Sample size	Population
Cigarettes smoked per day	ieu-b-142	GSCAN	NA	NA	249,752	European
Coffee intake	ukb-b-5237	MRC-IEU	NA	NA	428,860	European
Fresh fruit intake	ukb-b-3881	MRC-IEU	NA	NA	446,462	European
Cooked vegetable intake	ukb-b-8089	MRC-IEU	NA	NA	448,651	European
BMI	ukb-b-19953	MRC-IEU	NA	NA	461,460	European
Fasting insulin	ebi-a-GCST90002238	NA	NA	NA	151,013	European
Fasting glucose	ebi-a-GCST90002232	NA	NA	NA	200,622	European
Gut microbiome	NA	MiBioGen consortium	NA	NA	18,340	European (N=13,266), Middle-Eastern (N=481), East Asian (N=811), American Hispanic/Latin (N=1097), African American (N=114) multi-ancestry (N=2571)
POF	NA	FinnGen consortium	25,117	148,629	173,746	European

BMI, body mass index; POF, premature ovarian failure

geneous SNPs. Furthermore, if the genetic variants do not explain enough of the variance, there will be significant weak instrumental bias toward the confounded estimate.³³ To address this concern, SNP-specific F-statistics, approximated by the square of the beta divided by the variance for the SNP-exposure association, were calculated to evaluate the strength of the instruments used, and values exceeding the standard threshold of 10 are indicative of strong genetic instruments.³³

All tests were two-sided and performed using R Version 4.2.1 with the R packages “TwoSampleMR” and “MendelianRandomization”. A p value < 0.05 indicated statistical significance of the MR effect estimate. No ethical approval was required since we used publicly available summary data.

RESULTS

SNP selection

There were 22 SNPs associated with cigarettes smoked per day, 38 SNPs associated with coffee intake, 53 SNPs associated with fresh fruit intake, 17 SNPs associated with cooked vegetable intake, 414 SNPs associated with BMI, 37 SNPs associated with fasting insulin, 60 SNPs associated with fasting glucose, and 1508 SNPs associated with gut microbiota selected for the MR analyses according to the IV selection criteria. The detailed information and F-statistic for the selected instruments are shown in Supplementary Table 1. The overall instrument had a high F-statistic (>10), indicating the good strength of the genetic instruments used.

MR analysis

Figure 2 shows causal effect estimates of the dietary and metabolic factors and gut microbiota on POF from the IVW MR analyses. Associations for exposures using the different MR methods are presented in Supplementary Tables 2-4. Scatter and forest plots of the SNP-outcome associations against the SNP-exposure associations are shown in Supplementary Figures 1-6, allowing visualization of the causal effect estimate for each individual SNP on POF. Leave-one-out plots are shown in Supplementary

Figures 7-9 to evaluate the influential outliers.

MR analysis via the IVW method showed that cigarettes smoked per day (OR = 1.00, 95% CI: 0.77–1.30, p = 0.982), coffee intake (OR = 2.05, 95% CI: 0.87–4.84, p = 0.103), cooked vegetable intake (OR = 3.13, 95% CI: 0.37–26.09, p = 0.292), fasting insulin (OR = 1.61, 95% CI: 0.66–3.96, p = 0.298), and fasting glucose (OR = 1.04, 95% CI: 0.67–1.60, p = 0.864) had no genetic causal relationship with POF (Figure 2). However, fresh fruit intake (OR = 7.33, 95% CI: 2.36–22.71, p = 0.001) and BMI (OR = 1.99, 95% CI: 1.60–2.48, p < 0.001) were related to an increased risk of POF. In addition, six gut microbiome taxa were significantly associated with POF risk (Figure 2). IVW method revealed that *Eubacterium (hallii group)* and *Eubacterium (ventriosum group)* were negatively associated with the risk of POF (OR = 0.49, 95% CI: 0.26–0.90, p = 0.022; OR = 0.51, 95% CI: 0.27–0.97, p = 0.040), while *Adlercreutzia*, *Intestinibacter*, *Lachnospiraceae (UCG008)*, and *Terrisporobacter* were positively associated with the risk of POF (OR = 3.01, 95% CI: 1.38–6.60, p = 0.006; OR = 1.82, 95% CI: 1.04–3.20, p = 0.037; OR = 1.73, 95% CI: 1.08–2.76, p = 0.023; OR = 2.47, 95% CI: 1.14–5.36, p = 0.022) (Figure 2).

Sensitivity analyses

The observed causal associations were consistent in sensitivity analyses. MR-Egger regression showed no evidence of directional pleiotropic effect across the genetic variants (intercept, p > 0.05) (Table 2 and Supplementary Tables 5-7). There was no evidence of heterogeneity in the IVW analysis using Cochran’s Q test (p > 0.05) (Table 2 and Supplementary Tables 5-7). Although there were outliers present on visual inspection in both scatter (Supplementary Figures 1-3) and forest plots (Supplementary Figures 4-6), the results of the leave-one-out sensitivity analysis indicated that the associations between dietary and metabolic factors and gut microbiota with POF were not substantially driven by any individual SNP (Supplementary Figures 7-9), suggesting the robustness of the results.

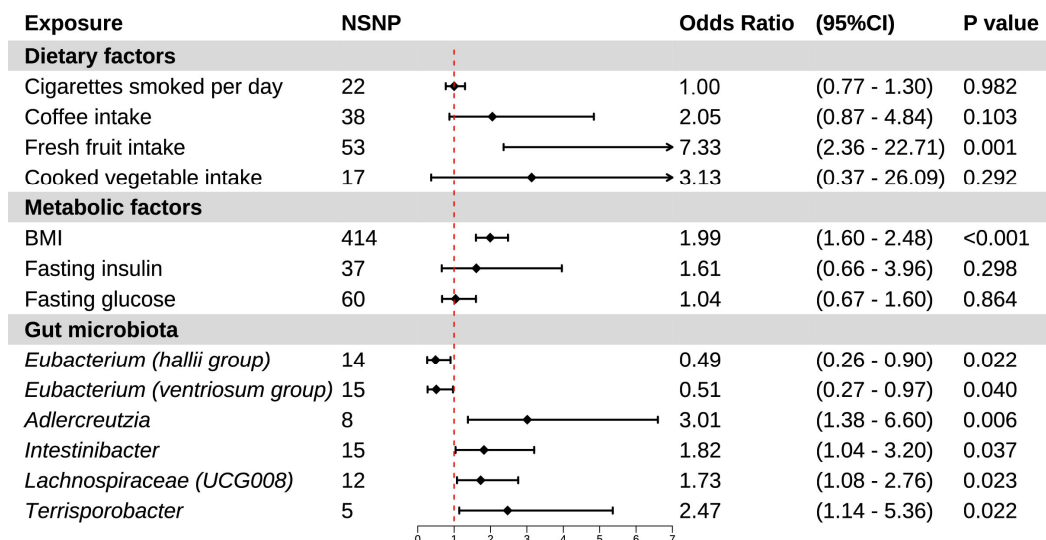


Figure 2. Associations of genetically predicted dietary and metabolic factors and gut microbiota with risk of POF. BMI, body mass index; POF, premature ovarian failure; SNP, single-nucleotide polymorphisms.

Table 2. Heterogeneity and directional pleiotropy tests from MR analysis of the dietary and metabolic factors and gut microbiota with risk of POF

Exposure	Heterogeneity		MR–Egger		
	Cochrane's Q	<i>p</i>	Egger Intercept	SE	<i>p</i> _{intercept}
Dietary factors					
Cigarettes smoked per day	12.09	0.937	0.000	0.02	0.990
Coffee intake	43.06	0.228	0.021	0.01	0.142
Fresh fruit intake	45.16	0.738	0.009	0.02	0.609
Cooked vegetable intake	24.40	0.081	0.000	0.13	0.999
Metabolic factors					
BMI	440.57	0.168	-0.006	0.01	0.233
Fasting insulin	43.27	0.189	0.002	0.02	0.944
Fasting glucose	50.14	0.788	0.003	0.01	0.758
Gut microbiota					
<i>Eubacterium (hallii group)</i>	9.01	0.773	-0.006	0.05	0.916
<i>Eubacterium (ventriosum group)</i>	10.02	0.761	-0.020	0.11	0.858
<i>Adlercreutzia</i>	9.55	0.215	0.279	0.14	0.085
<i>Intestinibacter</i>	12.90	0.535	-0.080	0.08	0.307
<i>Lachnospiraceae (UCG008)</i>	10.44	0.491	0.096	0.12	0.446
<i>Terrisporobacter</i>	2.93	0.570	0.050	0.12	0.703

BMI, body mass index.

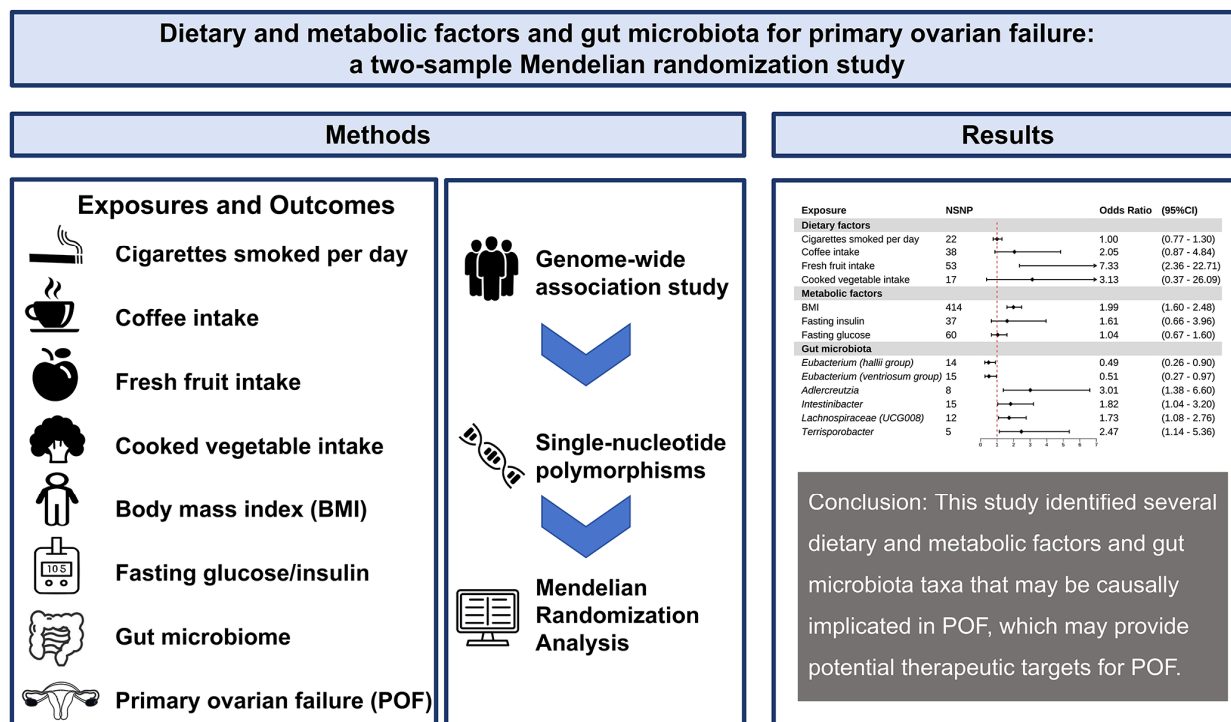
DISCUSSION

We conducted MR analyses by using the largest GWAS datasets to systematically investigate the causal relationship between the dietary and metabolic factors and gut microbiota with risk of POF. Our results showed that fresh fruit intake and BMI was associated with an increased risk of POF. Six gut microbiome taxa were associated with the risk of POF. *Eubacterium (hallii group)* and *Eubacterium (ventriosum group)* appeared to confer a protective effect against POF, while *Adlercreutzia*, *Intestinibacter*, *Lachnospiraceae (UCG008)*, and *Terrisporobacter* increased the risk of POF. This study could provide important insight into the genetic relationship between dietary and metabolic factors and gut microbiota with POF and shed new light on the potential causes and therapeutic strategies for POF.

Altered gut microbial profiles have been observed in women with POF.²¹ Additionally, Elgart et al.³⁴ reported that the gut bacteria of *Drosophila* can affect oogenesis and maternal-to-zygotic transition during embryo development. In this study, we found that *Eubacterium (hallii group)* and *Eubacterium (ventriosum group)* had protective effects on POF. *Eubacterium* produces short-chain fatty acids (SCFAs). SCFAs, including propionate, acetate and butyrate, are the main products of the fermentation of dietary fiber by the intestinal microbiota.³⁵ Butyrate can enhance the expression of tight-junction proteins and mucin to maintain the intestinal epithelial barrier,³⁶ which is the first line of defense in the intestine. The abundance of *Eubacterium* in the gut is strongly correlated with SCFA levels and the beneficial effects of SCFAs under a range of clinical conditions.³⁷ Several studies have shown that SCFAs play a major role in the modulation of inflammation through the inhibition of proinflammatory cytokines, such as interferon (IFN)- γ , interleukin (IL)-1 β , IL-6, IL-8, and tumor necrosis factor receptor- α (TNF- α), while upregulating the expression of anti-inflammatory cytokines, such as IL-10 and transforming growth factor- β (TGF- β).^{38,39} The human ovary is a ubiquitous target for autoimmune attack, leading to the consequent occurrence of POF.⁴⁰ Autoimmunity is responsible

for approximately 4–30% of POF cases.^{41,42} *E. hallii* and *E. ventriosum* may act as anti-inflammatory agents to protect the ovary from inflammation. On the other hand, we found that *Adlercreutzia*, *Intestinibacter*, *Lachnospiraceae (UCG008)*, and *Terrisporobacter* increased the risk of POF. Other studies have shown that these 4 gut microbiome taxa are correlated with the risk of diabetic retinopathy, male infertility, periodontitis, and sepsis.⁴³⁻⁴⁶ However, there is a lack of corresponding research evidence to clarify the underlying mechanism by which these gut microbiome taxa contribute to POF, thus providing new directions for future studies.

Smoking is a worldwide issue. Cigarette smoke contains several toxicants, including polycyclic aromatic hydrocarbons (PAHs), such as benzo(a)pyrene (BaP), nitrosamines, heavy metals (cadmium), alkaloids and aromatic amines, which have different properties and targets. Therefore, these chemical compounds may exert hazardous effects on the entire reproductive system in women.⁴⁷ It has been documented that active smoking was associated with earlier menopause.⁵⁻⁸ The tobacco-mediated ovarian injury characterizes by a significant decline in steroidogenesis,^{47,48} and folliculogenesis.⁴⁹⁻⁵⁴ Evidence from experimental models have shown that a single high dose of PAHs led to the loss of primordial and primary follicles.⁵⁵ In addition, *in-vitro* studies have demonstrated that BaP could induce follicular demise and alter the growth of rat and mouse follicles^{50,52}, and exposure to nicotine could cause decrease estradiol production in cultured granulosa bovine cells.⁵⁶ The pathophysiological mechanism behind tobacco-mediated ovarian injury involves a range of factors such as oxidative stress,⁵⁷⁻⁶⁰ DNA damages,⁶¹ and follicle loss through autophagy/apoptosis.⁶²⁻⁶⁵ However, a meta-analysis comprising 15 studies found a relationship in earlier age of natural menopausal in current smokers but the association disappeared in former smokers.⁶⁶ The results of this study show that there was no causal relationship between cigarettes smoked per day and POF at the genetic level. The association between cigarettes smoked per day and premature ovarian failure (POF) might be attenuated due to the way



Graphical abstract.

the smoking phenotypes are defined, as they include both current and former smokers. Although several studies were devoted to investigating the relationship between drinking coffee and the age of menopause, data are lacking on POF. Current studies have suggested that there is no association between coffee intake and early menopause or ovarian age indicators such as anti-Müllerian hormone (AMH) and FSH.⁹⁻¹¹ Combined with the results of our study, we considered that there is no causal relationship between coffee intake and POF at the genetic level. A large prospective study involving 33,054 Shanghai women has found that a high level of fruit intake (>383.2 g/day) was associated with delayed menopause.¹² Another study also supported this finding.¹³ The association of fruit intake with POF could, in part, be related to the antioxidant content in fruit. However, according to our study, fresh fruit intake was associated with an increased risk of POF. Potential mechanisms underlying this association needs to be explored in mechanistic studies. The association between BMI and POF remains much less understood and even controversial. Both overweight and underweight had been reported to be associated with earlier menopause. Our study can provide evidence of causal association. Various mechanisms could explain how overweight might influence the development of ovarian aging. It is known that being overweight can increase oxidative stress in the body through a number of potential mechanisms.^{67,68} In addition, obesity is related to chronic low-grade inflammation in the body.⁶⁹ Adipose tissue is an important endocrine organ that produces adipokines contributing to a state of inflammation.

This study has several strengths. The major merit is MR design which can exclude the interference of confounding factors and reverse causality to a large extent. Furthermore, non-overlapping exposure and outcome

summary-level data were used to avoid unnecessary bias.⁷⁰ In order to ensure the accuracy of MR analysis, horizontal pleiotropy was detected and excluded by the MR-Egger regression intercept test. Limitations should be considered when interpreting our results. First, our study analyzed only European populations, so the generalizability of the results should be approached with caution when extending them to other populations. Secondly, our study was only conducted at the genetic level, and did not explore the exact mechanisms behind the association. Finally, since genus was the lowest taxonomic level in the exposure datasets, we were unable to further explore the causal association between gut microbiota and POF at the species level.

Conclusion

To conclude, we provided evidence to show that fresh fruit intake and BMI was associated with an increased risk of POF. Six gut microbiome taxa were associated with the risk of POF. *Eubacterium (hallii group)* and *Eubacterium (ventriosum group)* appeared to confer a protective effect against POF, while *Adlercreutzia*, *Intestinibacter*, *Lachnospiraceae (UCG008)*, and *Terrisporobacter* increased the risk of POF. Our results provided potential therapeutic targets for POF. At the same time, it is necessary to validate these findings and explore the underlying mechanisms in clinical trials and animal models.

SUPPLEMENTARY MATERIALS

All supplementary tables and figures are available upon request.

ACKNOWLEDGEMENTS

We express our gratitude to the participants of the FinnGen study and the MiBioGen consortium for releasing the gut microbiota GWAS summary statistics.

CONFLICT OF INTEREST AND FUNDING DISCLOSURES

The authors report there are no competing interests to declare.

This research was funded by National Key Research and Development Program of China (No. 2022YFC2703803, No.2022YFC2703001, No.2021YFC2700603), National Natural Science Foundation of China (No.82088102, No.82171613, No.82171688), CAMS Innovation Fund for Medical Sciences (2019-I2M-5-064), Collaborative Innovation Program of Shanghai Municipal Health Commission (2020CXJQ01), Key Discipline Construction Project (2023-2025) of Three-Year Initiative Plan for Strengthening Public Health System Construction in Shanghai (GWVI-11.1-35), Shanghai Clinical Research Center for Gynecological Diseases (22MC1940200), Shanghai Urogenital System Diseases Research Center (2022ZZ01012), Shanghai Frontiers Science Research Center of Reproduction and Development, Zhejiang Province College Student Science and Technology Innovation Program (Xinmiao Plan) (2023R401210).

REFERENCES

- Woad KJ, Watkins WJ, Prendergast D, Shelling AN. The genetic basis of premature ovarian failure. *Aust N Z J Obstet Gynaecol.* 2006; 46: 242-4. doi:10.1111/j.1479-828X.2006.00585.x.
- Goswami D, Conway GS. Premature ovarian failure. *Hum Reprod Update.* 2005; 11: 391-410. doi:10.1093/humupd/dmi012.
- Qin Y, Jiao X, Simpson JL, Chen ZJ. Genetics of primary ovarian insufficiency: new developments and opportunities. *Hum Reprod Update.* 2015; 21: 787-808. doi: 10.1093/humupd/dmv036.
- Yang Y, Huang W, Yuan L. Effects of environment and lifestyle factors on premature ovarian failure. *Adv Exp Med Biol.* 2021;1300:63-111. doi: 10.1007/978-981-33-4187-6_4.
- Mattison DR. The effects of smoking on fertility from gametogenesis to implantation. *Environ Res.* 1982; 28: 410-33. doi: 10.1016/0013-9351(82)90139-6.
- Adena MA, Gallagher HG. Cigarette smoking and the age at menopause. *Ann Hum Biol.* 1982; 9:121-30. doi: 10.1080/03014468200005591.
- Baron JA, La Vecchia C, Levi F. The antiestrogenic effect of cigarette smoking in women. *Am J Obstet Gynecol.* 1990; 162: 502-14. doi: 10.1016/0002-9378(90)90420-c.
- Sun L, Tan L, Yang F, Luo Y, Li X, Deng HW, Dvornyk V. Meta-analysis suggests that smoking is associated with an increased risk of early natural menopause. *Menopause.* 2012; 19: 126-32. doi: 10.1097/gme.0b013e318224f9ac.
- Mikkelsen TF, Graff-Iversen S, Sundby J, Bjertness E. Early menopause, association with tobacco smoking, coffee consumption and other lifestyle factors: a cross-sectional study. *BMC Public Health.* 2007; 7: 149. doi: 10.1186/1471-2458-7-149.
- Kline J, Tang A, Levin B. Smoking, alcohol and caffeine in relation to two hormonal indicators of ovarian age during the reproductive years. *Maturitas.* 2016; 92: 115-22. doi: 10.1016/j.maturitas.2016.07.010.
- Kinney A, Kline J, Kelly A, Reuss ML, Levin B. Smoking, alcohol and caffeine in relation to ovarian age during the reproductive years. *Hum Reprod.* 2007; 22: 1175-85. doi: 10.1093/humrep/del496.
- Dorjgochoo T, Kallianpur A, Gao YT, Cai H, Yang G, Li H, Zheng W, Shu XO. Dietary and lifestyle predictors of age at natural menopause and reproductive span in the Shanghai Women's Health Study. *Menopause.* 2008; 15: 924-33. doi: 10.1097/gme.0b013e3181786adc.
- Wang M, Gong WW, Hu RY, Wang H, Guo Y, Bian Z, Lv J, Chen ZM, Li LM, Yu M. Age at natural menopause and associated factors in adult women: Findings from the China Kadoorie Biobank study in Zhejiang rural area. *PLoS One.* 2018;13: e0195658. doi: 10.1371/journal.pone.0195658.
- Adak A, Khan MR. An insight into gut microbiota and its functionalities. *Cell Mol Life Sci.* 2019; 76: 473-93. doi: 10.1007/s00018-018-2943-4.
- Qi X, Yun C, Pang Y, Qiao J. The impact of the gut microbiota on the reproductive and metabolic endocrine system. *Gut microbes.* 2021; 13: 1-21. doi: 10.1080/19490976.2021.1894070.
- Zhou L, Ni Z, Cheng W, Yu J, Sun S, Zhai D, Yu C, Cai Z. Characteristic gut microbiota and predicted metabolic functions in women with PCOS. *Endocr Connect.* 2020; 9: 63-73. doi: 10.1530/EC-19-0522.
- Lindheim L, Bashir M, Münzker J, Trummer C, Zachhuber V, Leber B, Horvath A, Pieber TR, Gorkiewicz G, Stadlbauer V, Obermayer-Pietsch B. Alterations in gut microbiome composition and barrier function are associated with reproductive and metabolic defects in women with polycystic ovary syndrome (PCOS): a pilot study. *PloS one.* 2017; 12: e0168390. doi: 10.1371/journal.pone.0168390.
- Liu R, Zhang C, Shi Y, Zhang F, Li L, Wang X, et al. Dysbiosis of gut microbiota associated with clinical parameters in polycystic ovary syndrome. *Front Microbiol.* 2017; 8: 324. doi: 10.3389/fmicb.2017.00324.
- Ata B, Yildiz S, Turkgeldi E, Brocal VP, Dinleyici EC, Moya A, Urman B. The endobiota study comparison of vaginal, cervical and gut microbiota between women with stage 3/4 endometriosis and healthy controls. *Sci Rep.* 2019; 9: 2204. doi: 10.1038/s41598-019-39700-6.
- Shan J, Ni Z, Cheng W, Zhou L, Zhai D, Sun S, Yu C. Gut microbiota imbalance and its correlations with hormone and inflammatory factors in patients with stage 3/4 endometriosis. *Arch Gynecol Obstet.* 2021; 304: 1363-73. doi: 10.1007/s00404-021-06057-z.
- Wu J, Zhuo Y, Liu Y, Chen Y, Ning Y, Yao J. Association between premature ovarian insufficiency and gut microbiota. *BMC Pregnancy Childbirth.* 2021; 21: 418. doi: 10.1186/s12884-021-03855-w.
- Hu X, Xu X, Zeng X, Jin R, Wang S, Jiang H, et al. Gut microbiota dysbiosis promotes the development of epithelial ovarian cancer via regulating Hedgehog signaling pathway. *Gut Microbes.* 2023; 15: 2221093. doi: 10.1080/19490976.2023.2221093.
- Lynch SV, Pedersen O. The human intestinal microbiome in health and disease. *NEJM.* 2016; 375: 2369-79. doi: 10.1056/NEJMra1600266.
- Evans DM, Davey Smith G. Mendelian randomization new applications in the coming age of hypothesis-free causality. *Annu Rev Genomics Hum Genet.* 2015; 16: 327-50. doi: 10.1146/annurev-genom-090314-050016.
- Lawlor DA, Harbord RM, Sterne JAC, Timpson N, Davey Smith G. Mendelian randomization using genes as instruments for making causal inferences in epidemiology. *Stat Med.* 2008; 27: 1133-63. doi: 10.1002/sim.3034.
- Kurilshikov A, Medina-Gomez C, Bacigalupe R, Radjabzadeh D, Wang J, Demirkan A, et al. Large-scale association analyses identify host factors influencing human gut microbiome composition. *Nat Genet.* 2021; 53: 156-65. doi: 10.1038/s41588-020-00763-1.
- Sekula P, Del Greco MF, Pattaro C, Köttgen A. Mendelian randomization as an approach to assess causality using

- observational data. *J Am Soc Nephrol.* 2016; 27: 3253-65. doi: 10.1681/ASN.2016010098.
28. Burgess S, Scott RA, Timpson NJ, Davey Smith G, Thompson SG; EPIC-InterAct Consortium. Using published data in Mendelian randomization: a blueprint for efficient identification of causal risk factors. *Eur J Epidemiol.* 2015; 30: 543-52. doi:10.1007/s10654-015-0011-z.
 29. Bowden J, Davey Smith G, Haycock PC, Burgess S. Consistent estimation in Mendelian randomization with some invalid instruments using a weighted median estimator. *Genet Epidemiol.* 2016; 40: 304-14. doi: 10.1002/gepi.21965.
 30. Bowden J, Davey Smith G, Burgess S. Mendelian randomization with invalid instruments effect estimation and bias detection through Egger regression. *Int J Epidemiol.* 2015; 44: 512-25. doi: 10.1093/ije/dyv080.
 31. Greco M FD, Minelli C, Sheehan NA, Thompson JR. Detecting pleiotropy in Mendelian randomisation studies with summary data and a continuous outcome. *Stat Med.* 2015; 34: 2926-40. doi: 10.1002/sim.6522.
 32. Burgess S, Thompson SG. Interpreting findings from Mendelian randomization using the MR-Egger method. *Eur J Epidemiol.* 2017; 32: 377-89. doi: 10.1007/s10654-017-0255-x.
 33. Davies NM, Holmes MV, Davey Smith G. Reading Mendelian randomisation studies a guide, glossary, and checklist for clinicians. *BMJ.* 2018; 362. doi: 10.1136/bmj.k601.
 34. Elgart M, Stern S, Salton O, Gnainsky Y, Heifetz Y, Soen Y. Impact of gut microbiota on the fly's germ line. *Nat Commun.* 2016; 7: 11280. doi: 10.1038/ncomms11280.
 35. Miller TL, Wolin MJ. Pathways of acetate, propionate, and butyrate formation by the human fecal microbial flora. *Appl Environ Microbiol.* 1996; 62: 1589-92. doi: 10.1128/aem.62.5.1589-1592.1996.
 36. Morrison DJ, Preston T. Formation of short chain fatty acids by the gut microbiota and their impact on human metabolism. *Gut Microbes.* 2016; 7: 189-200. doi: 10.1128/aem.62.5.1589-1592.1996.
 37. Mukherjee A, Lordan C, Ross RP, Cotter PD. Gut microbes from the phylogenetically diverse genus *Eubacterium* and their various contributions to gut health. *Gut Microbes.* 2020; 12: 1802866. doi: 10.1080/19490976.2020.1802866.
 38. Mowat AM, Agace WW. Regional specialization within the intestinal immune system. *Nat Rev Immunol.* 2014; 14: 667-85. doi: 10.1038/nri3738.
 39. Corrêa-Oliveira R, Fachi JL, Vieira A, Sato FT, Vinolo MAR. Regulation of immune cell function by short-chain fatty acids. *Clin Transl Immunology.* 2016; 13: 461-72. doi: 10.1038/cti.2016.17.
 40. Petříková J, Lazúrová I. Ovarian failure and polycystic ovary syndrome. *Autoimmun Rev.* 2012; 11:A471-8. doi: 10.1016/j.autrev.2011.11.010.
 41. Ebrahimi M, Akbari Asbagh F. The role of autoimmunity in premature ovarian failure. *Iran J Reprod Med.* 2015; 13: 461-72.
 42. Jiao X, Zhang H, Ke H, Zhang J, Cheng L, Liu Y, Qin Y, Chen ZJ. Premature ovarian insufficiency phenotypic characterization within different etiologies. *J Clin Endocrinol Metab.* 2017; 102: 2281-90. doi: 10.1210/jc.2016-3960.
 43. Liu K, Zou J, Fan H, Hu H, You Z. Causal effects of gut microbiota on diabetic retinopathy: a Mendelian randomization study. *Front Immunol.* 2022; 13: 930318. doi: 10.3389/fimmu.2022.930318.
 44. Fu ZD, Wang Y, Yan HL. Male infertility risk and gut microbiota: a Mendelian randomization study. *Front Microbiol.* 2023; 14: 1228693. doi: 10.3389/fmicb.2023.1228693.
 45. Ye X, Liu B, Bai Y, Cao Y, Lin S, Lyu L, et al. Genetic evidence strengthens the bidirectional connection between gut microbiota and periodontitis: insights from a two-sample Mendelian randomization study. *J Transl Med.* 2023; 21: 674. doi: 10.1186/s12967-023-04559-9.
 46. Chen JH, Zeng LY, Zhao YF, Tang HX, Lei H, Wan YF, Deng YQ, Liu KX. Causal effects of gut microbiota on sepsis: a two-sample Mendelian randomization study. *Front Microbiol.* 2023; 14: 1167416. doi: 10.3389/fmicb.2023.1167416.
 47. Dechanet C, Anahory T, Mathieu Daude JC, Quantin X, Reyftmann L, Hamamah S, Hedon B, Dechaud H. Effects of cigarette smoking on reproduction. *Hum Reprod Update.* 2011; 17: 76-95. doi: 10.1093/humupd/dmq033.
 48. Soldin OP, Makambi KH, Soldin SJ, O'Mara DM. Steroid hormone levels associated with passive and active smoking. *Steroids.* 2011; 76: 653-9. doi: 10.1016/j.steroids.2011.02.042.
 49. Neal MS, Zhu J, Foster WG. Quantification of benzo[a]pyrene and other PAHs in the serum and follicular fluid of smokers versus non-smokers. *Reprod Toxicol.* 2008; 25: 100-6. doi: 10.1016/j.reprotox.2007.10.012.
 50. Igawa Y, Keating AF, Rajapaksa KS, Sipes IG, Hoyer PB. Evaluation of ovotoxicity induced by 7, 12-dimethylbenz[a]anthracene and its 3,4-diol metabolite utilizing a rat in vitro ovarian culture system. *Toxicol Appl Pharmacol.* 2009; 234: 361-9. doi: 10.1016/j.taap.2008.10.009.
 51. Ramesh A, Archibong AE, Niaz MS. Ovarian susceptibility to benzo[a]pyrene: tissue burden of metabolites and DNA adducts in F-344 rats. *J Toxicol Environ Health A.* 2010; 73: 1611-25. doi: 10.1080/15287394.2010.514225.
 52. Sadeu JC, Foster WG. Effect of in vitro exposure to benzo[a]pyrene, a component of cigarette smoke, on folliculogenesis, steroidogenesis and oocyte nuclear maturation. *Reprod Toxicol.* 2011; 31: 402-8. doi: 10.1016/j.reprotox.2010.12.006.
 53. Sadeu JC, Foster WG. Cigarette smoke condensate exposure delays follicular development and function in a stage-dependent manner. *Fertil Steril.* 2011; 95: 2410-7. doi: 10.1016/j.fertnstert.2011.03.072.
 54. Richardson MC, Guo M, Fauser BC, Macklon NS. Environmental and developmental origins of ovarian reserve. *Hum Reprod Update.* 2014; 20: 353-69. doi: 10.1093/humupd/dmt057.
 55. Mattison DR, Thorgeirsson SS. Ovarian aryl hydrocarbon hydroxylase activity and primordial oocyte toxicity of polycyclic aromatic hydrocarbons in mice. *Cancer Res.* 1979; 39: 3471-5.
 56. Sanders SR, Cuneo SP, Turzillo AM. Effects of nicotine and cotinine on bovine theca interna and granulosa cells. *Reprod Toxicol.* 2002; 16: 795-800. doi: 10.1016/s0890-6238(02)00049-7.
 57. Nampoothiri LP, Agarwal A, Gupta S. Effect of co-exposure to lead and cadmium on antioxidant status in rat ovarian granulosa cells. *Arch Toxicol.* 2007; 81: 145-50. doi: 10.1007/s00204-006-0133-x.
 58. Mai Z, Lei M, Yu B, Du H, Liu J. The effects of cigarette smoke extract on ovulation, oocyte morphology and ovarian gene expression in mice. *PLoS One.* 2014; 9: e95945. doi: 10.1371/journal.pone.0095945.
 59. Sobinoff AP, Pye V, Nixon B, Roman SD, McLaughlin EA. Jumping the gun: smoking constituent BaP causes premature

- primordial follicle activation and impairs oocyte fusibility through oxidative stress. *Toxicol Appl Pharmacol.* 2012; 260: 70-80. doi: 10.1016/j.taap.2012.01.028.
60. Sobinoff AP, Beckett EL, Jarnicki AG, Sutherland JM, McCluskey A, Hansbro PM, McLaughlin EA. Scrambled and fried: cigarette smoke exposure causes antral follicle destruction and oocyte dysfunction through oxidative stress. *Toxicol Appl Pharmacol.* 2013; 271: 156-67. doi: 10.1016/j.taap.2013.05.009.
61. Jennings PC, Merriman JA, Beckett EL, Hansbro PM, Jones KT. Increased zona pellucida thickness and meiotic spindle disruption in oocytes from cigarette smoking mice. *Hum Reprod.* 2011; 26: 878-84. doi: 10.1093/humrep/deq393.
62. Tuttle AM, Stämpfli M, Foster WG. Cigarette smoke causes follicle loss in mice ovaries at concentrations representative of human exposure. *Hum Reprod.* 2009; 24: 1452-9. doi: 10.1093/humrep/dep023.
63. Gannon AM, Stämpfli MR, Foster WG. Cigarette smoke exposure leads to follicle loss via an alternative ovarian cell death pathway in a mouse model. *Toxicol Sci.* 2012; 125: 274-84. doi: 10.1093/toxsci/kfr279.
64. Gannon AM, Stämpfli MR, Foster WG. Cigarette smoke exposure elicits increased autophagy and dysregulation of mitochondrial dynamics in murine granulosa cells. *Biol Reprod.* 2013; 88: 63. doi: 10.1095/biolreprod.112.106617.
65. Furlong HC, Stämpfli MR, Gannon AM, Foster WG. Cigarette smoke exposure triggers the autophagic cascade via activation of the AMPK pathway in mice. *Biol Reprod.* 2015; 93: 93. doi: 10.1095/biolreprod.115.132183.
66. Schoenaker DA, Jackson CA, Rowlands JV, Mishra GD. Socioeconomic position, lifestyle factors and age at natural menopause: a systematic review and meta-analyses of studies across six continents. *Int J Epidemiol.* 2014;43:1542–62. doi: 10.1093/ije/dyu094.
67. Olusi S. Obesity is an independent risk factor for plasma lipid peroxidation and depletion of erythrocyte cytoprotective enzymes in humans. *Int J Obes.* 2002; 26, 1159–64. doi: 10.1038/sj.ijo.0802066.
68. Adnan MT, Amin MN, Uddin MG, Hussain MS, Sarwar MS, Hossain MK, Uddin SMN, Islam MS. Increased concentration of serum MDA, decreased antioxidants and altered trace elements and macro-minerals are linked to obesity among Bangladeshi population. *Diabetes Metab Syndr.* 2019; 13, 933–8. doi: 10.1016/j.dsx.2018.12.022.
69. Khanna D, Khanna S, Khanna P, Kahar P, Patel BM. Obesity: A chronic low-grade inflammation and its markers. *Cureus.* 2022, 14, e22711. doi: 10.7759/cureus.22711.
70. Burgess S, Davies NM, Thompson SG. Bias due to participant overlap in two-sample Mendelian randomization. *Genet Epidemiol.* 2016;40:597–608. doi: 10.1002/gepi.21998.