Original Article

Modulation of gene expression by traditional Asian antidiabetic nutraceuticals: A review of potential effects

Katrin Roosita PhD¹, Karina Rahmadia Ekawidyani MD, PhD¹, Rosyanne Kushargina PhD², Fathimah MPH^{3,4}, Mohamad Rafi PhD⁵, Uus Saepuloh PhD⁶

Background and Objectives: Type 2 diabetes mellitus (T2DM) has emerged as a significant global public health concern. Multiple studies have shown that traditional nutraceuticals are potential to treat T2DM and its complications. This review will explore traditional nutraceuticals with antidiabetic properties with a focus on traditional Asian nutraceuticals and their antioxidant effects on gene expression associated to T2DM. Methods and Study Design: Literature searching was conducted in Pubmed, Scopus, and Science Direct using the keywords "nutraceutical", "antidiabetic", "insulin resistance", "Diabetes Mellitus", "herbal medicine", "mechanism", "pathway", "traditional food", "functional food", "antioxidant", "clinical", "preclinical", "animal studies", and "Asian" combined with Boolean operators "OR". Results: Nutraceuticals sourced from traditional Indonesian herbal beverages, including Galohgor, Bir Pletok, and Wedang Uwuh, have shown potential efficacy in reducing hyperglycemia, oxidative stress, and obesity in T2DM. Furthermore, multiple Asian plants and their bioactive compounds, such as curcumin, kaempferol, cinnamon, saponin, quercetin, myricetin, anthocyanin, terpenoid, alkaloid, and gallic acid, have been shown to beneficially influence glucose homeostasis, insulin sensitivity, and problems associated with diabetes. Moreover, bioactive compounds of these traditional nutraceuticals have been proven in modulating gene expression associated with β-cell function, insulin signaling pathway, and antioxidant activity, which may offer a new therapeutic target. Conclusions: This review highlights the increasing scientific evidence on the role of traditional nutraceuticals for the prevention and management of diabetes mellitus, presenting promising alternatives to standard pharmacological therapy. Nonetheless, double-blind randomized clinical trials are required to validate these antidiabetic effects.

Key Words: antioxidants, Asian traditional nutraceutical, gene expression, oxidative stress, type 2 diabetes mellitus

INTRODUCTION

Diabetes mellitus has become a major global public health issue, with its prevalence quadrupling over the past three decades. The global prevalence of diabetes among individuals aged 20–79 was estimated at 10.5%, affecting approximately 536.6 million people. This figure is projected to rise to 12.2% (783.2 million) by 2045. Diabetes rates were comparable between men and women, with the highest prevalence observed in the 75–79 age group. The prevalence in urban areas (12.1%) was higher than in rural areas (8.3%), and diabetes was more common in high-income countries (11.1%) compared to low-income nations (5.5%). The most significant relative increase in diabetes prevalence between 2021 and 2045 is projected in middle-income countries (21.1%), surpassing that of high-income (12.2%) and low-income (11.9%) nations. ^{2,3}

Diabetes mellitus is a burdensome disease due to its

long-term complications and potential to cause disability. It is also the ninth primary cause of death. In 2021, an estimated 6.7 million deaths worldwide were attributed to diabetes among individuals aged 20–79 years. Furthermore, global direct healthcare expenditure on diabetes for individuals aged 20–79 was estimated at USD 966 billion in 2021 and is projected to rise to USD 1,054 billion by

Corresponding Author: Prof. Katrin Roosita and Dr Karina Rahmadia Ekawidyani, Department of Community Nutrition, Faculty of Human Ecology, IPB University, Jl. Lingkar Akademik, Kampus IPB Darmaga, Bogor 16680, Indonesia Tel: +622518625066

Email: kroosita2@apps.ipb.ac.id; karinare@apps.ipb.ac.id Manuscript received 26 December 2024. Initial review completed 20 January 2025. Revision accepted 12 May 2025. doi: 10.6133/apjcn.202510_34(5).0006

¹Department of Community Nutrition, Faculty of Human Ecology, IPB University, Bogor, Indonesia

²Nutrition Study Program, Faculty of Medicine and Health, Universitas Muhammadiyah Jakarta, Jakarta, Indonesia

³Doctoral Study Program in Nutrition Sciences, Department of Community Nutrition, Faculty of Human Ecology, IPB University, Bogor, Indonesia

⁴Nutritional Science Study Program, Faculty of Health Sciences, Universitas Darussalam Gontor, Ponorogo, East Java, Indonesia

⁵Department of Chemistry, Faculty of Mathematics and Natural Science, IPB University, Bogor, Indonesia ⁶Primate Research Center IPB University, Bogor, Indonesia

2045.² The long-term complication of diabetes mellitus involved micro- and macrovascular complications, including cardiovascular complications, kidney failure, and cerebral complications.⁵

Current treatment for type 2 diabetes mellitus (T2DM) involves pharmacological therapy and lifestyle modification. Antidiabetic drugs are classified into several groups, including insulin sensitizers (including biguanides, thiazolidinediones), insulin secretagogues (including sulfonylureas, meglitinides), alpha-glucosidase inhibitors, as well as newer therapies such as incretin-based agents and sodium-glucose co-transporter 2 (SGLT2) inhibitors.⁶

Due to the potential adverse effects of long-term medication use, natural therapeutic approaches, such as nutraceuticals, have gained increasing attention as complementary or alternative treatments for diabetes.^{6,7} Nutraceuticals are products containing food extracts or bioactive components that have demonstrated significant health advantages for customers in recent years.8 According to Mali et al. (2022), nutraceutical is a food or a part of food that has a long historical background in the treatment of various diseases. Nutraceuticals play a significant role in maintaining normal physiological functions and overall human health. They encompass a wide range of products, including dietary supplements, herbal products, genetically engineered "designer" foods, and isolated nutrients. One category of nutraceuticals, known as traditional nutraceuticals, is derived directly from natural sources and includes nutrients, herbs, and phytochemicals.⁹ These are habitually used within specific groups, communities, or cultures.

The efficacy of traditional nutraceuticals and functional food in the treatment of T2DM and its complications has been demonstrated in multiple previous studies.^{7,10,11} In addition, Blahova et al.⁶ and Venkatakrishnan et al.⁷ concluded that combining conventional hypoglycemic drugs with lifestyle modifications and functional foods or traditional nutraceuticals can significantly improve glucose control and reduce diabetes-related complications.

Asia possesses diverse ancient medicinal system originating from herbs and functional foods, including traditional Chinese medicine, Ayurvedic practices from India, and Jamu from Indonesia. In the past decade, there has been a surge in research on traditional nutraceuticals components and functional foods exhibiting antidiabetic properties. These herbal remedies and functional foods contain bioactive compounds, including flavonoid and non-flavonoid polyphenols, which have been shown to lower blood sugar levels. These bioactive compounds exhibit multiple antidiabetic mechanisms, including antioxidant activity and the regulation of various processes: enhancement of glucose reabsorption, glucagon-like peptide-1 homeostasis, beta cell function, and insulin resistance.

Several comprehensive reviews have examined traditional nutraceuticals and functional foods with antidiabetic properties, highlighting the molecular pathways involved in the prevention and management of diabetes. An Nonetheless, these studies exclusively address the antidiabetic properties of individual plants or food. This review aims to explore traditional nutraceuticals with antidiabetic properties derived from various plants and foods,

with a particular emphasis on Asian traditional medicine. This review will focus on their mechanisms for improving hyperglycemia, highlighting their antioxidant effects on gene expression involved in the pathophysiology of T2DM.

METHODS

Literature searching was conducted in Pubmed, Scopus, and Science Direct using the keywords "nutraceutical", "antidiabetic", "insulin resistance", "Diabetes Mellitus", "herbal medicine", "mechanism", "pathway", "traditional food", "functional food", "antioxidant", "clinical", "preclinical", "animal studies", and "Asian" combined with Boolean operators "OR". Articles from 1973–2024 were included in this review. The inclusion criteria for articles in this review are in vitro studies, animal studies, and clinical trials that investigated the effect of nutraceuticals, herbal, and traditional medicine from Asian countries as antidiabetic and its mechanistic pathway. The exclusion criteria are systematic reviews and meta-analyses articles. We also discovered some relevant literature by manually searching the references to the included articles and using Google Scholar. Ethics approval is not required for this type of study.

RESULTS

Development of type 2 diabetes mellitus

T2DM is a metabolic disorder primarily characterized by hyperglycemia. However, it also disrupts lipid and protein metabolism due to reduced insulin secretion, insulin resistance, or both. T2DM is a complex condition influenced by hereditary and environmental factors. Risk factors for developing diabetes mellitus include advanced age, non-white ancestry, family history of diabetes, genetic factors, overweight or obesity, polycystic ovarian syndrome, history of atherosclerotic heart disease, unhealthy eating habits (such as red meat, sugary drinks, low intake of whole grains and fiber), smoking, sedentary lifestyle, history of macrosomia or gestational diabetes, skin hyperpigmentation (acanthosis nigricans), short or excessively long sleep duration, shift work, and economic and psychosocial factors. 15

Unhealthy foods are an important risk factor for T2DM. Several pathways could be influenced by the consumption of particular foods and nutrients. Refined grains and sugar sweetened beverages contain simple carbohydrates that directly induce postprandial plasma glucose and insulin secretion. Simple carbohydrates are rapidly absorbed, thus rapidly increase postprandial plasma glucose. Prolonged intake of high simple carbohydrate could increase insulin resistance. 16,17

Insulin resistance

Insulin resistance is characterized by a reduced biological response to insulin in target tissues, primarily the liver, muscle, and adipose tissue. This impairment disrupts glucose metabolism, prompting a compensatory increase in insulin secretion by pancreatic β -cells, resulting in hyperinsulinemia. In obesity, insulin resistance in the adipose tissue increases lipolysis and free fatty acids (FFA) level in plasma. The rise of plasma FFA stimulates gluconeogenesis, worsens insulin resistance in muscle and the liver, and plays a role in β -cell failure, impairing insulin

secretion. These abnormalities caused by increased plasma FFA are called lipotoxicity. 17,18

In obese T2DM patients, increased levels of pro-inflammatory cytokines, such as interleukin-6 (IL-6) and Tumor Necrosis Factor alpha (TNF-α), and increased numbers of macrophages and other inflammatory cells are observed in adipose tissue, liver, and serum. Inflammation appears mainly in adipose tissue and the liver. Infiltration of macrophages in adipose tissue promotes lipolysis and an increase level of IL-6 can trigger hepatic gluconeogenesis and generate hepatic insulin resistance.¹⁵

Initially, insulin resistance occurs but can be compensated by elevated insulin production. As the condition advances, the pancreatic beta cells lose their capacity to sustain elevated insulin production, resulting in reduced glucose tolerance and manifest diabetes. Factors contributing to beta-cell failure comprise glucotoxicity, lipotoxicity, inflammation, and abnormalities in genetics.

The core defects in T2DM are insulin resistance in the muscle and liver, along with reduced insulin secretion by the pancreatic β -cells. Insulin resistance in muscle results in reduced glucose uptake, whereas in the liver increased hepatic glucose production (gluconeogenesis).

β-cell dysfunction

In normal condition, insulin secretion by β -cell is governed tightly by coupling between glucose metabolites from the tricarboxylic acid (TCA) cycle and nucleotides. Glucose enters the β -cell via glucose transporter 2 (Glut2) by exploiting a concentration gradient. During glycolysis, glucose undergoes phosphorylation that creates pyruvate, which is then transported into the mitochondria, resulting in Adenosine Triphosphate (ATP) generation. This process also produces reactive oxygen species (ROS) as by-products. Increased ATP elevates the ATP/Adenosine Diphosphate (ADP) ratio causing the cellular KATP-channels to close, subsequently depolarizes cell membrane and opens the Ca²+ channels. The increase of Ca²+ levels in the cytosol triggers insulin exocytosis. Increase of Ca²+ levels in the cytosol triggers insulin exocytosis.

During hyperglycemic states, pyruvate excess enters the TCA cycle elevates nicotinamide adenine dinucleotide/flavin adenine dinucleotide (NADH/FADH2) entry into the mitochondrial electron transport chain, hence augmenting ROS formation.²² In hyperlipidemia, increased FFA levels result in the oxidation of both FFA and acetyl coenzyme A (CoA) within the TCA cycle. This enhances donation of NADH/FADH2 to the electron transport chain, leading to excessive ROS generation and oxidative stress.²³ Elevated amounts of ROS can induce mitochondrial injury by triggering the mitochondrial permeability transition pore to open and subsequent depolarization. Consequently, endogenous antioxidants escape from mitochondria, resulting in mitochondrial depletion and death.²⁴ Mitochondrial injury diminishes the ATP/ADP ratio, obstructing membrane depolarization and the activation of Ca²⁺ channels. This results in hindered and inadequate insulin secretion.²⁵

The involvement of other systems or organs in hyperglycemia

According to De Fronzo et al., ¹⁵ there were eight organs known as the 'ominous octet' involved in the pathogenesis of hyperglycemia in T2DM. They are pancreas β -cells and α -cells, muscle, liver, adipose tissue, gastrointestinal (stomach, intestine, and colon), kidney, and brain. In addition, three pathophysiological abnormalities contributing to muscle insulin resistance, i.e., activation of inflammatory pathways and immune system, as well as impaired insulin-mediated vasodilation, also have a role in hyperglycemia, making the 'decadent decuplet' (Figure 1).

Normally, incretin effects performed by glucagon-like polypeptide-1 (GLP-1) and glucose-dependent insulinotropic polypeptide (GIP) increase insulin secretion when glucose is administered orally compared to intravenously. Meanwhile, GLP-1 deficiency and GIP resistance is found in T2DM, which reduce incretin effects in the gastrointestinal tract, consequently, increase hyperglycemia.⁵

Accelerated gastric emptying is more common in early diabetes with few complications. Gastric emptying is a major factor that influences postprandial blood glucose and vice versa. Accelerated gastric emptying will increase the rate of nutrient delivery to small intestine which will increase the glucose absorption in the intestine and stimulates the release of GLP-1 and GIP. Increased glucose absorption will increase the level of postprandial blood glucose. Both the acute hyperglycemia and GLP-1 will slow gastric emptying. Meanwhile, GLP-1 and GIP will stimulate the secretion of insulin from pancreas, that will lower the blood glucose. However, in the chronic and complicated diabetes, gastroparesis (delayed gastric emptying) is more common.²⁶ In addition, dysbiosis of the gut microbiota also contributes to the development of T2DM through production of abnormal metabolites, including short chain fatty acids, lipopolysaccharides, trimethylamine, and metabolites of aromatic amino acids.²⁷

The function of pancreas α -cells is to produce glucagon. During the fasting state, its level in plasma will increase. This increase, together with augmented hepatic sensitivity to glucagon, will enhance hepatic glucose production. 5,15

Kidney also has a role in T2DM pathogenesis. The maintenance of hyperglycemia is in part due to increased renal glucose reabsorption by the sodium/glucose co-transporter 2 (SGLT2) and the increased threshold for glucose spillage in the urine. ¹⁵ The kidney filters 163 g of glucose in a day, of which 90% are reabsorbed by SGLT2 in the convoluted proximal tubule. The rest 10% will be reabsorbed by SGLT1 in the ascending and descending tubules; thus, no glucose will be found in urine. In T2DM patients, SGLT2 gene expression is increased. ⁵

The brain is involved in controlling the appetite that can contribute to weight gain and loss. Unfortunately, insulin resistance also happens in the brain impairing the appetite-suppressive effects of insulin. In addition, leptin resistance, GLP1, amylin, and peptide YY, together with low brain dopamine and increased brain serotonin levels, contribute to weight gain. All of this exacerbates the underlying resistance.¹⁵

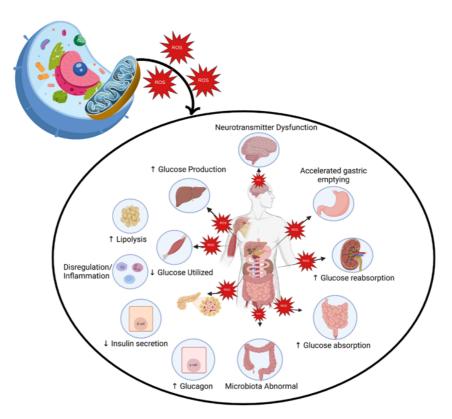


Figure 1. Organs involved in the pathogenesis of hyperglycemia in type 2 diabetes mellitus (T2DM). The pathogenesis of hyperglycemia in TD2M begins with oxidative stress in cells caused by increased Reactive Oxygen Species (ROS). Eleven organs and cells were involved in the pathogenesis of hyperglycemia in TD2M, including neurotransmitter dysfunction in the brain, dysfunction in pancreas β -cells and α -cells, accelerated gastric emptying, increased glucose reabsorption in the kidney, glucose absorption in the intestine and abnormal microbiota in the colon, lipolysis in adiposity, and glucose production in the liver. Pathogenesis also involves insulin resistance, decreased glucose utilization in muscle cells and dysregulation of Antibody Cell. (This figure was generated using BioRender.com)

Vascular insulin resistance reduces microvascular recruitment, in which only one of three capillaries are open at rest.¹⁵ In normal conditions, vasodilation results from insulin metabolic signaling via increased endothelial cell nitric oxide (NO) production, thus increasing bioavailable NO. Conversely, insulin resistance promotes vasoconstriction and vascular proliferation by activating the mitogen-activated protein kinase (MAPK) cascade. This cascade coordinates vasoconstriction induced by insulin resistance and growth-promoting effects.²⁸

The modulation of gene expression involved in the gluconeogenesis pathway plays a pivotal role in the regulation of glucose homeostasis within the body. The mechanism of gluconeogenesis involving the phosphoenolpycarboxy kinase (PEPCK) and glucose-6phosphatase (G6Pase) genes is crucial in the regulation of glucose production in the liver. PEPCK is a key enzyme in the gluconeogenesis pathway that catalyzes the conversion of oxaloacetate to phosphoenolpyruvate (PEP). PEPCK gene expression is regulated by various transcription factors influenced by hormones, such as glucagon and cortisol, which increase PEPCK expression in the fasting state to increase glucose production. G6Pase is an enzyme that catalyzes the hydrolysis of glucose-6phosphate (G6P) to glucose, which is the final step in gluconeogenesis. G6Pase gene expression is also regulated by hormones and metabolic conditions. When glucose levels are low, hormones such as glucagon will increase the expression of G6Pase to favor the release of glucose into the circulation. The expression of these two genes

(PEPCK and G6Pase) is regulated by hormonal signals that reflect the nutritional status of the body. In the fasting state, hormones such as glucagon and cortisol will increase the expression of these genes, while insulin will decrease their expression to reduce glucose production. Although the liver is the main source of glucose production, studies show that when gluconeogenesis in the liver is impaired, extra-hepatic tissues such as the kidneys and intestines can compensate by increasing their gluconeogenic enzyme activity.²⁹

Role of oxidative stress in T2DM development and complications

ROS are primary contributors to oxidative stress, and their generation is an inevitable result of metabolic processes. The main source of ROS production is the leakage of electrons from the mitochondrial respiration process, which then converts to molecular oxygen, leading to the synthesis of superoxide anion (O_2^-) .³⁰ The activation of the NADPH oxidase enzyme could also generate O_2^- .³¹ Elevated auto-oxidation and nonenzymatic glycosylation are additional potential pathways that may initiate the generation of free radicals and radical-induced lipid peroxidation. The elevated generation of ROS disrupts the equilibrium between oxidant and antioxidant levels, leading to a pro-oxidative condition.³²

Oxidative stress is also involved in the pathogenesis and development of T2DM. Genetic predispositions and other environmental factors including overnutrition, high energy intake, high fat and carbohydrate consumption, and physical inactivity may contribute to obesity and metabolic syndrome. This condition increases oxidative stress due to high levels of ROS. Oxidative stress together with increase in FFA and cytokines produced by adipose tissue will induce insulin resistance.³³

Hyperglycemia induces ROS overproduction through the activation of the polyol pathway, augmented production of advanced glycation end products (AGEs), stimulation of the diacylglycerol-protein kinase C (DAG-PKC) pathway, and excessive activation of the hexosamine pathway. These mechanisms interact to facilitate mitochondrial failure, inflammation, and disruption of insulin signaling, leading to insulin resistance and pancreatic βcell malfunction, which are characteristic of T2DM. 32,34 Pancreatic beta cells generate elevated levels of ROS and have decreased synthesis of antioxidative enzymes. Consequently, these cells possess a diminished endogenous antioxidant capability, rendering them particularly vulnerable to oxidative stress.35 They only have approximately 50% of the superoxide dismutase (SOD) and 5% of the hydrogen peroxide-scavenging enzymes, i.e. glutathione peroxidase (GPx) and catalase (CAT), relative to the quantities present in the liver.^{35–37}

Oxidative stress is a major factor of glucose toxicity in DM as it leads to decreased levels of two insulin gene transcription factors, Pancreatic and Duodenal Homeobox 1 (PDX-1) and MafA. PDX-1 and MafA normally bind to the insulin promoter and stimulate insulin gene transcription. Decreased levels of these two proteins lead to decreased insulin promoter activity, insulin gene expression and insulin secretion.³⁸ Clinical observations of the relationship between glucose toxicity and oxidative stress are associated with decreased levels of two insulin promoter transcription factors namely, PDX-1 and Regulatory Insulin Promoter Element (RIPE)-3b1 activator / Musculoaponeurotic fibrosarcoma oncogene homolog A (MafA).³⁹ Hyperglycemic risk in mice is increased due to disruption of PDX-1, MafA binding to Deoxyribonucleic Acid (DNA), decreased insulin messenger Ribonucleic Acid (mRNA), decreased glucose-induced insulin secretion in islets of Langerhans in the pancreas.⁴⁰

Among various transcription factors, PDX-1 plays an important role in pancreatic development and pancreatic beta cell differentiation and serves as a transcriptional activator of the insulin gene. Neurogenin-3 (Ngn3) and NeuroD are also important transcription factors for pancreatic endocrine cell differentiation. MafA expression is at a late stage of pancreatic beta cell differentiation and functions as a potential insulin gene activator of transcription. Pancreatic transcription factors have the potential to induce various beta cell-related genes including insulin. Insulin.

MafA is β -cell specific and also functions as an insulin gene activator. Expression of pancreatic transcription factors in non-beta cells of the pancreas, liver, intestine, and bone marrow cells induces gene expression of various beta cells including insulin. Baumel-Alterzon S. et al⁴² reported that nuclear factor erythroid 2-related factor (Nrf2) controls redox balance and affects PDX-1 levels, where pharmacological activation of the Nrf2 pathway can alleviate diabetes by maintaining PDX-1 levels.

Oxidative stress also contributes to the development of T2DM complications. Increased ROS could immediately impair DNA, proteins, and lipids, modifying their function as well as their structure, leading to cellular malfunction and impairment of normal biological functions. Oxidative damage also stimulates inflammation which contributes to the onset of microvascular problems such as diabetes nephropathy, retinopathy, and neuropathy, and it also causes macrovascular complications such as cardiovascular disease. ^{34,43}

Given the significant impact of oxidative stress in the development and complications of T2DM, efforts for limiting oxidative stress in diabetes should include antioxidant therapy, in addition to lifestyle changes and efficient hyperglycemic management. Yet, more study is required to completely figure out the fundamental mechanisms of oxidative stress in diabetes. Additionally, studies to assess the effectiveness of antioxidant therapies, including natural antioxidants derived from traditional nutraceuticals, in managing diabetes and preventing its complications are also essential. By focusing on oxidative stress, it is expected to reduce diabetes problems and enhance the health of patients.⁴³

Traditional nutraceuticals from Asia with potential antidiabetic effects

Traditional nutraceuticals from Indonesia

Traditional nutraceuticals from Indonesia have gained attention for their potential health effects, especially those passed down through generations, such as galohgor, bir pletok, wedang uwuh, and loloh Bali. These herbal concoctions, rich in natural bioactive compounds, are deeply rooted in Indonesian culture and have been used for centuries to promote health and wellness. The diverse blend of spices, herbs, and medicinal plants in these traditional drinks contributes to their ability to help regulate blood glucose levels, improve insulin sensitivity, and combat oxidative stress. This makes them promising alternatives or complementary options for managing diabetes in a natural way.

Galohgor

Galohgor is one of the Indonesian traditional herbal medicines (Jamu) originating from the Sundanese ethnicity of West Java. The composition of Galohgor found in Sukajadi Village, Tamansari District, Bogor Regency, consists of 56 types of medicinal plants in the form of leaves, nuts, herbs, and spices. Based on literature studies, kaempferol was identified in 75% of the plant species used in the formulation of Galohgor, contributing to 95.4% of the total weight of the 56 plants analyzed. Notably, several of these plants have demonstrated potential antidiabetic properties, including Psidium guajava, 44 Zingiber aromaticum Veleton, 45 Kaempferia galanga L., 46 Melastoma malabathricum, 47 Sonchus arvensis L., 48 Myristica fragrans, 49 Zingiber officinale, 50 Persea americana,⁵¹ Piper betle,⁵² Pluchea indica, Zea mays L.,⁵³ Phaseolus radiatus L.,53 and Amomum cardamomum L.45 (Table 1). Galohgor is consumed by the Sundanese people in powder form as a snack. Galohgor has a content of fat, protein, carbohydrates, zinc, magnesium, vitamin C, carotenoids (β-carotene), vitamin E, and phenolic compounds. Based on pre-clinical and clinical studies,

Table 1. Types and nutraceutical content of Asian plants with potential to improve diabetes mellitus outcome

Plant name	Phytochemicals	Potential effect on DM
Ficus religiosa	Tanin, saponin, Polyphenol compounds, flavonoid, dan sterol	-Antidiabetic ⁹⁶
(Bodhi in Indonesia and Malaysia, Peepal in India,	Leucocyanidin 3-O-β-D-galactosyl cello bioside, leucopelargonidin-3-O-alpha-	-Antihyperglicemic ⁹⁷
puti shu in China, Pho in Thailand)	L rhamnoside ^{130,131}	-↓ blood glucose levels
		-increased insulin levels
		-modulates antioxidant enzymes to counteract oxidative stress
Eugenia jambolana	Seeds: alkaloid, jambosine and the glycoside jambolin,	-antioxidant defense
(Jamblang and Duwet in Indonesia, Jambulana in		-hypoglycemic
Malaysia, Jamun in India, Hei pu tao in China, Wa	Fruits: Glucose, fructose, raffinose, malic acid, anthocyianin ¹³²	-↓ glucose
in Thailand)		-↑ insulin secretion inhibition of insulin degradation ⁹⁸
	Leaves: Acylated flavonol glycosides, quercetin, myricetin, and tanin ¹³³	-↓ blood cholesterol, triglycerides, and free fatty acids ¹³⁴
		-↓ 3-HMG Co-A reductase enzyme activity ¹³⁵
		-↓ blood pressure
		-hipoglycemic ¹³³
Momordica charantia	Seeds: Vicine, charantin, and triterpenoids along with several antioxidants,	-antidiabetic and hypoglycemic ¹³⁶
(Pare or Paria in Indonesia, Peria or Peria Katak	saponin ⁹⁹	-reduction in waist circumference, improvement in diabetes, and
in Malaysia, Karela in India, Kugua in China,		symptoms of metabolic syndrome ¹⁰⁰
Mara in Thailand)		-repair β-cells stimulate insulin levels ¹³⁷
		-↑ insulin sensitivity/signaling ¹³⁸
		-inhibits glucose uptake ¹³⁹
		-inhibits β-glucosidase activity ¹⁴⁰
		-inhibition of β-amylase and β-glucosidase
		-stimulates insulin secretion ¹⁴¹
Ocimum sanctum L	Leaves: Eugenol, flavonoids, saponins, tannins, triterpenoids, rosmarinic acid,	-↓ blood glucose ¹⁴³
(Ruku-ruku/kemangi hutan/ holy Basil in Indone-	apigenin, isothymusin, isothymonin, cirsimaritin, orientin, vicenin, and antho-	-↑ insulin secretion ¹⁴⁴
sia, Selasih Hutan or Selasih India in Malaysia,	cyanin ¹⁴²	-↓ glucose serum ¹⁴⁵
Tulsi in India, Luole in China, Kaphrao in Thai-		-↑ antioxidant ¹⁴⁶
land),	Seeds: fatty acid and sitosterol	-↓ glucose serum, cholesterol, triglyserid, dan LDL ¹⁴⁷
Pterocarpus marsupium	Terpenoids and phenolic compounds: β-sitosterol, lupeol, aurone glycosides,	-Antidiabetic activity ¹⁴⁸
(Vijaysar in India)	epicatechin and iso-flavonoid ⁹²	-hypoglycemic and β cell regeneration ¹⁴⁹
		-improves HbA1C ¹⁵⁰
		-regulates glucose production through modulating AKT and
		AMPK in HepG2 cells ¹⁵¹
Trigonella foenum-graecum	Saponins, 4-hydroxyisoleucine, trigonelline, alkaloid, and steroid ⁹²	-↓ glucose and lipid ¹⁵²
(Kelabat in Indonesia, Halba in Malaysia, Methi in		-† insulin sensitivity and glucose uptake in peripheral tissues ¹⁵³
India, Hu' lu' ba' in China, Luk Sanai in Thailand)		

3-HMG Co-A: 3-Hydroxy-3-Methylglutaryl-Coenzyme A, LDL: Low-Density Lipoprotein, HbA1C: Hemoglobin A1c, AKT: serine/threonine kinase, AMPK: AMP-activated protein kinase, HepG2: Human Hepatocellular Carcinoma cell line 2, GLUT-4: Glucose Trasporter 4, 3T3-L1: mouse embryo fibroblasts, MDA: Malondialdehyde, AR: Androgen Receptor, TNF-α: Tumor Necrosis Factor-alpha, IL-6: Interleukin 6, SOD: Superoxide Dismutase, PTP1B: Protein Tyrosine Phosphatase 1B, HOMAIR: Homeostasis Model Assessment of Insulin Resistance, ROS: Reactive Oxygen Species.

Table 1. Types and nutraceutical content of Asian plants with potential to improve diabetes mellitus outcome (cont.)

Plant name	Phytochemicals	Potential effect on DM
Gymnema sylvestre	Triterpenoid saponins, gymnemic acids, and gymnema saponins	-Regeneration of β cells in the pancreas
(Gurmart in Indonesia, Pokok Mas Cotek Gurmar		reduce blood sugar levels ¹⁵⁴
in Malaysia, Gurmar in India, Jian ming cao in		
China, Khwao Khurea in Thailand)		
Allium sativum	allicin, allixin, ajoene, and other organosulfur compounds.	-↓ fasting blood sugar level ¹⁵⁵
(Bawang Putih in Indonesia and Malaysia, Lahsun		- \uparrow insulin secretion from β cells
in India, Da suan in China, Krathiam in Thailand)		-↑ endogenous antioxidant defense ¹⁵⁶
Tisanes	alkaloids, carotenoids,	-↑ insulin secretion ⁹³
(Wedang in Indonesia, Teh herbal in Malaysia,	coumarins, flavonoids, phenolic acids, polyacetylenes, saponins, terpenoids ⁹³	
Kadha in India, Huacao cha in China, Nam Chea		
in Thailand)		
Rhus chinensis Mill	Leaves, stem and branches: Phenolics (protocatechuic acid, p-coumaric acid,	-↓ Postprandial glucose levels ⁹⁵
(Kakkarsingi in India, Yan fu mu in China)	gallic acid, catechin, quercetin, methyl gallate), antioxidants (hydroxydamma-	
	renone, semialactone, moronic acid, betulonic acid)	
	Fruit: Antioxidants, phenolics (gallic acid, tannic acid), flavonoids	
	Gall: Hydrolysable tannins (gallotannins), phenolic acid (gallic acid), antioxi-	
	dant	
Murraya koenigii l. (meethi nimba)	α-pinene, sabinene, dan β-caryophyllene	-↓ hyperglycemia
(Daun Kari in Indonesia and Malaysia, Kadi Patta	r, r ., r y . r y	-↑ insulin sensitivity ⁹⁴
in India, <i>Yue ju</i> in China, <i>Bai karip</i> in Thailand)		
Andrographis paniculata (Burm.f.) Nees/ Fa thalai	Aerial parts :	-↓ HbA1c
<i>chon</i> ¹⁵⁷	Glycosides, terpenoid, alkaloid, flavonoid, saponin, tannins	
(Sambiloto in Indonesia, Hempedu bumi in Malay-		
sia, Kalmegh in India, Chuan xin lian in China,		
Fah Talai Jone in Thailand)		
Pluchea indica (L.) Less/ Khluu ⁶⁴	Leaves: chlorogenic acid (CGA), 3,4-O-dicaffeoylquinic acid (3,4 diCQA),	-↓ Blood glucose levels
(Beluntas in Indonesia and Malaysia, rasna in	3,5-O-dicaffeoylquinic acid (3,5 diC-	
India, Po bu mu in China, Phak Krathin Ban in	QA), quercetin, kaempferol, myricetin, monoterpenes, and sesquiterpenoids	
Thailand)		
Apium graveolens L. /Khuen chaai ⁶⁵ (Celery)	flavonoids, alkaloids, terpenoids, and phenolic acids	-↓ Blood glucose levels
(Seledri in Indonesia, Seleri in Malaysia, Ajmod in		
India, <i>Qincai</i> in China, <i>Khuen Chai</i> in Thailand)	Al.,	1 1 14
Aloe vera (L.) Burm.f./Wan Hang Chora Khe Nees ^{67–69,158}	Aloe-emodin, Aloetic-acid, Anthranol, Barbaloin, Mannan, 8-C-glusoly-(2'-O-	-\ weight
	cinnamoly), –7-O-methlyaloediol A, Alkaline phosphatese, amylase, bradyki-	-\ fat mass
(Lidah buaya in Indonesia and Malaysia, Ghritku-	nase, carboxypeptidase, catalase, cyclooxidase, cyclooxygenase, lipase, oxi-	-\ insulin response
mari in India, Lu hui in China, Wan Hang Chora	dase, phosphoenolpyruvate, carboxylase	-↓ Blood glucose levels
Khe Nees in Thailand)		

3-HMG Co-A: 3-Hydroxy-3-Methylglutaryl-Coenzyme A, LDL: Low-Density Lipoprotein, HbA1C: Hemoglobin A1c, AKT: serine/threonine kinase, AMPK: AMP-activated protein kinase, HepG2: Human Hepatocellular Carcinoma cell line 2, GLUT-4: Glucose Trasporter 4, 3T3-L1: mouse embryo fibroblasts, MDA: Malondialdehyde, AR: Androgen Receptor, TNF-α: Tumor Necrosis Factor-alpha, IL-6: Interleukin 6, SOD: Superoxide Dismutase, PTP1B: Protein Tyrosine Phosphatase 1B, HOMAIR: Homeostasis Model Assessment of Insulin Resistance, ROS: Reactive Oxygen Species.

Table 1. Types and nutraceutical content of Asian plants with potential to improve diabetes mellitus outcome (cont.)

Plant name	Phytochemicals	Potential effect on DM
Ipomoea aquatica Forssk./ Pak bung ⁷⁰	euphornin, lucidenic acid, and myricitin glycosides,	-↓ Blood glucose levels
Lagerstroemia speciosa (L.) Pers./ Inthanin nam ⁷¹	flavonoids, saponins, tannins, steroids, and triterpenoids	-↓ Blood glucose levels
(Kangkung in Indonesia and Malaysia, Kalmi Saag		
in India, Kong xin cai in China, pak bung in Thai-		
land)		
Terminalia bellirica (Gaertn.) Roxb. /Samo phi	Fruit: gallic acid, galloyl glucose, chebulagic acid, ellagic acid, β-sitosterol,	-↓ Blood glucose levels
phek ⁷⁵	ethylgallate, sugar, bellericanin, lignans, and flavan.	-↓ Lipids levels
(Balirik in Indonesia, Bahera in Malaysia and		-↑ Antioxidants
India, He zi in China, Samo Phi Phe in Thailand)		
Terminalia chebula Retz./ Samo thai ⁷⁵	chebulanin, chebulagic acid, and chebulinic acid	- Blood glucose levels
(Kayu kuning / haritaki in Indonesia, Ketepeng		-↓ Lipids levels
and Buah Keras in Malaysia, Haritaki in India, He		-↑ Antioxidants
zi in China, Samo Thailand)		- DI 1 1 1 1
Morinda citrifolia L. /Yo Baan ⁷⁹ (noni)	amino acids, anthraquinones, fatty acids, flavonoids, iridoids,	-↓ Blood glucose levels
(Mengkudu in Indonesia and Malaysia, Indian	lignans, polysaccharides, sterols, terpenoid	
Mulberry/noni in India, Ba ji Tian in China, Yo	D. 1. 1. C. 1. (c. 1. (c. 1. c. 1. c	
Ban in Thailand)	Bark, leaf: α-ketoglutaric and malic acids	D11-111-
Zea mays L./Khao Phot ⁷⁸	volatile oils, steroids, saponins, polysaccharides, alkaloids,	- Blood glucose levels
(Jagung in Indonesia and Malaysia, Makai in India, Yumi in China, Khao Phot in Thailand)	flavonoids, organic acids and other phenolic compounds	-↓ oxidative damage in the brain of diabetic mice ⁶⁵
Eclipta prostrata (L.) / Ka meng ⁸⁰	Whole plant: wedelolactone, eclalba saponins, ursolic acid, oleanolic acid,	-↓ Blood glucose levels
	luteolin, and apigenin	
(Urang-aring in Indonesia and Malaysia, Bhringraj in India, Mo Han Lian in China, Ka	ideom, and apigenm	-↓ Postprandial glucose levels
meng in Thailand)		
Phyllanthus amarus / Ma kham pom ^{81,82}	Alkaloids: securinine, epibubbialine, isobubbialine	-↓ Blood glucose levels
(Meniran in Indonesia and Malaysia, Bhui Amla in	Aikaroids. securinine, epiodooraniie, isoodooraniie	- Postprandial glucose levels
India, Yi ke yin chen in China, Ma kham pom in		- to stprandial glucose levels
Thailand)		
Curcuma longa L. /Khamin ⁸³	Rhizomes: Curcumin, curcumol, and bisdemethoxycurcumin	-↓ Blood glucose levels
(Kunyit in Indonesia and Malaysia, Haldi in India,	ranzones. Carcanni, carcanoi, and obsernation year canin	- Postprandial glucose levels
Jiang Huang in China, Khamin in Thailand)		-\ Arterial stiffness,
views 22views in Children, Historiem in Thailand)		-\ Endothelial dysfunction
		-↑ β-cells function
Gymnema inodorum (Lour.) Decne./Chiang Daa ⁷²	Leaves: phenolic acids, flavonoids, triterpenoid compounds, and pregnane gly-	-\ Postprandial glucose levels
(Chiang Daa in Thailand)	cosides	v 1 0

³⁻HMG Co-A: 3-Hydroxy-3-Methylglutaryl-Coenzyme A, LDL: Low-Density Lipoprotein, HbA1C: Hemoglobin A1c, AKT: serine/threonine kinase, AMPK: AMP-activated protein kinase, HepG2: Human Hepatocellular Carcinoma cell line 2, GLUT-4: Glucose Trasporter 4, 3T3-L1: mouse embryo fibroblasts, MDA: Malondialdehyde, AR: Androgen Receptor, TNF-α: Tumor Necrosis Factor-alpha, IL-6: Interleukin 6, SOD: Superoxide Dismutase, PTP1B: Protein Tyrosine Phosphatase 1B, HOMAIR: Homeostasis Model Assessment of Insulin Resistance, ROS: Reactive Oxygen Species.

Table 1. Types and nutraceutical content of Asian plants with potential to improve diabetes mellitus outcome (cont.)

Plant name	Phytochemicals	Potential effect on DM
Aegle marmelos (L.) / Ma tuum ⁷³ (Maja in Indonesia, Pokok Maja in Malaysia, Bael in India, Mu Ju in China, Matum in Thailand)	marmenol, marmin, marmelosin, marmelide, psoralen, alloimperatorin, rutaretin, scopoletin, aegelin, marmelin, fagarine, anhydromarmelin, limonene, âphellandrene, betulinic acid, marmesin, imperatorin, marmelosin, luvangentin and auroptene	-↓ Postprandial glucose levels
Glycine max (L.) Merr ¹⁵⁹ /black soybean (Kedelai in Indonesia, kacang soya in Malaysia, Soya Bean in India, Dadou in China, Tua Luang in Thailand)	Leaf: isoflavones and anthocyanins	-↓ Postprandial glucose levels
Salacia chinensis L./ Kam paeng jed chan ¹⁶⁰ (Srigading/akar kuning in Indonesia, Akar kuning/seruntun in Malaysia, Ponkoranti in India, Hei Mian Shen in China, Ching-cha-cha in Thailand)	Steam: salacinol, kotalanol, ponkoranol, and salaprinol and their corresponding de-0-sulfonated compounds. In addition, triterpenes, sesquiterpenes, lignans, xanthones, flavanols, flavonoids	-↓ Postprandial glucose levels -Hypoglycemic effect
Artocarpus heterophyllus Lam. / Khanun ⁶⁶ (Jackfruit) (Nangka in Indonesia and Malaysia, Kethal in India, Boluomi in China, Khanun in Thailand)	Leaves: flavonoid	-↑ insulin response
Phyllanthus emblica L./ Ma Kham Pom ⁷⁷ (Malaka/Ciremai India in Indonesia, Melaka/Pokok malaka in Malaysia, Amla in India, Yougan in China, Ma-Kham-Pom in Thailand)	Fruits: phenolic compounds (such as tannins, phenolic acids, and flavonoids), alkaloids, phytosterols, terpenoids, organic acids, amino acids, and vitamins	-↓ oxidative stress ⁷⁷
Guava leaf / Psidium guajava Linn (Jambu biji/Jambu klutuk in Indonesia, Jambu batu in Malaysia, Amrud in India, Fan shiliu in China, Farang in Thailand)	Potassium, phosporus, nitrogen, ascorbic acid, ficose, rhamnose, arabinose, galactose, glucose, mannose, xylose, phenol, sulfat, carbohydtrat, proteins, vitamin, minerals, α -Pinene Benzaldehyde, p-cymene 0.52% Limonene, 1,8-Cineole, β -cis-Ocimene, γ -Terpinene, α -Terpineol, β -Caryophyllene, α -Humulene, Quercetin, avicularin, apigenin, guaijaverin, kaempferol, hyperin, myricetin, Gallic acid, catechin, epicatechin, chlorogenic acid, epigallocatechin gallate, caffeic acid, Proanthocyanidins (PAs) ¹⁶¹	-↑ the function of β-cells of pancreatic islets and hepatocyte morphology ¹⁰⁸ -↓ activity of the blood glucose homeostasis enzyme dipeptidyl-peptidase IV ¹⁶² -inhibited intracellular lipid aggregation by impeding glucose uptake through GLUT-4 in vitro and revealed no distinct toxicity for 3T3-L1 adipose cells ¹⁶³ -reduction in total cholesterol, triglycerides, glycated serum protein, creatinine, fasting blood glucose, and malonaldehyde content, and increased total superoxide dismutase and total antioxidant capacity enzyme activity in vivo ¹⁶⁴ -inhibitors of α-amylase and α-glucosidase enzyme can decline postprandial glucose absorption ¹⁶⁵
Kaempferia galangal L (Kencur in Indonesia, Cekur in Malaysia, Chan- dramula in India, Shajiang in China, Proh Horm in Thailand)	Terpenoids (kaempulchraol I, E, L, kaemgalangol A) phenolics (p-metho-xybenzoicacid, p-hydroxybenzoic acid, vanillic acid, ferulic acid), and flavonoids (kaempferol, luteolin, kaempferide) ¹⁶⁶	-↓ blood lipid levels, along with reducing MDA, AR, TNF-α, and IL-6 levels and increasing SOD levels, ↓ blood glucose, insulin resistance, reducing the AR pathway as well as anti-oxidation and anti-inflammation ¹⁶⁷

3-HMG Co-A: 3-Hydroxy-3-Methylglutaryl-Coenzyme A, LDL: Low-Density Lipoprotein, HbA1C: Hemoglobin A1c, AKT: serine/threonine kinase, AMPK: AMP-activated protein kinase, HepG2: Human Hepatocellular Carcinoma cell line 2, GLUT-4: Glucose Trasporter 4, 3T3-L1: mouse embryo fibroblasts, MDA: Malondialdehyde, AR: Androgen Receptor, TNF-α: Tumor Necrosis Factor-alpha, IL-6: Interleukin 6, SOD: Superoxide Dismutase, PTP1B: Protein Tyrosine Phosphatase 1B, HOMAIR: Homeostasis Model Assessment of Insulin Resistance, ROS: Reactive Oxygen Species.

Table 1. Types and nutraceutical content of Asian plants with potential to improve diabetes mellitus outcome (cont.)

Plant name	Phytochemicals	Potential effect on DM
Cinnamomum verum	cinnamic acid, E)-cinnamaldehyde, cinnamaldehyde dimethyl aceta, Ascorbic	- Modulating insulin secretion and insulin receptor signaling,
(Kayu manis in Indonesia and Malaysia, Dalchini in India, ZhenGui in China, Ob Chuey in Thailand)	acid, Fumaric acid, Caffeic acid, Luteolin-7-rutinoside, Luteolin 7-glycoside, Rutin, Apigenin 7-glycoside, Quercitrin, Quercetin, kaempferol, Naringenin, Luteolin, Apigenin, Hispidulin, Chrysin ^{168,169}	enhancing GLUT4 expression, and exhibiting anti- α -glucosidase and anti- α -amylase activities
Zingiber aromaticum Veleton (Lempuyang in Indonesia, Lempoyang in Malaysia, Fangxiang Jiang in China, Plai Farang in Thai- land)	humulatrien-5-ol-8-one, kaempferol-3,4'-di-O-methyl ether, and (S)-6-gingerol. ⁴⁵	-Inhibits the activity of the enzyme protein tyrosine phosphatase $1B\ (PTP1B)^{45}$
Melastoma malabathricum (Senduduk in Indonesia and Malaysia, Malabar Malastome in India, Ye mudan in China, Koi / Ngaa Khao in Thailand)	quercetin, quercitrin, rutin, kaempferol, kaempferol-3-O-($2''$, $6''$ -di-O-p-transcoumaroyl)- β -D-glucoside, naringin, malabathrins A, B, C, and D, nobotanins B, D, G, and H, casuarictin, strictinin, pterocarinin C, pedunculagin, epicatechin, epicatechin gallate, and patriscabatrin. 170	-↓ Blood glucose levels ⁴⁷
Soncus arvensis L. (Tempuyung in Indonesia, Tempuyung/Daun dewa liar in Malaysia, ku cai in China, Phak Kat Khwai in Thailand)	Terpenoid, flavonoid (catechin, mirecetin, kaempferol, quercetin), phenolic, alkaloids. 171	-↓ blood glucose levels and cell regeneration in the tubular ⁴⁸
Myristica fragrans (Pala in Indonesia and Malaysia, Jaiphal in India, Rou Dou Kou in China, Chan Thet in Thailand)	ellagic acid (35.42 mg/g), rutin (91.07 mg/g), quercitrin (35.83 mg/g), quercetin (41.16 mg/g), and kaempferol (36.81 mg/g) 172	-↓ fasting blood glucose and HbA1C ¹⁷³
Zingiber officinale (Jahe in Indonesia, Halia in Malaysia, Adrak in India, Sheng Jiang in China, Khing in Thailand)	ginger, such as gingerol, shogaols, paradols, and zingerone, phenolic compounds (rutin, kaempferol, 6-gingerol, zingerone, naringenin, and quercetin). 174	-↓ fasting blood glucose, HbA1C, HOMAIR ⁵⁰
Persea americana (Alpukat in Indonesia, Avocado in Malaysia, Makhanphal in India, Niu You Guo in China, Abo-Ka-Do in Thailand)	Kaempferol 3-O- β -d-fucopyranoside, juglanin and astragaline, afzelin and quercitrin, catechin and epicatechin 175	-↓ fasting blood glucose and regeneration of islets of Langerha ⁵¹
Piper betle (Sirih in Indonesia, Sireh in Malaysia, Paan in India, Luo Ye in China, Phlu in Thailand)	alkaloids, terpenoids/steroids, flavonoids, polyphenols, tannins, and saponins ¹⁷⁶	-↓ Blood glucose levels ⁵²
Pluchea indica (Beluntas in Indonesia and Malaysia, Rasan in India, Ci Hao in China, Khlu in Thailand)	kaempferol 3-[2"',3"',5"'-triacetyl]-alpha-L-arabinofuranosyl-(1->6)-glucoside, myricetin 3-glucoside-7-galactoside, quercetin 3-(3"-sulfatoglucoside), and kaempferol 7,4'-dimethyl ether 3-O-sulfate ¹⁷⁷	-↓ Blood glucose levels ⁵³
Phaseolus radiatus L (Kacang Hijau in Indonesia and Malaysia, Moong Dal in India, Lu Dou in China, Tua Kiew in Thai- land)	flavonol (quercetin and kaempferol) aglycones while few others contain either of the two aglycone. 178	-↓ROS formation, inhibits alpha glucosidase and alpha amylase activity ¹⁷⁹

3-HMG Co-A: 3-Hydroxy-3-Methylglutaryl-Coenzyme A, LDL: Low-Density Lipoprotein, HbA1C: Hemoglobin A1c, AKT: serine/threonine kinase, AMPK: AMP-activated protein kinase, HepG2: Human Hepatocellular Carcinoma cell line 2, GLUT-4: Glucose Trasporter 4, 3T3-L1: mouse embryo fibroblasts, MDA: Malondialdehyde, AR: Androgen Receptor, TNF- α : Tumor Necrosis Factor-alpha, IL-6: Interleukin 6, SOD: Superoxide Dismutase, PTP1B: Protein Tyrosine Phosphatase 1B, HOMAIR: Homeostasis Model Assessment of Insulin Resistance, ROS: Reactive Oxygen Species.

Table 2. Modulation of gene expression in diabetic conditions by traditional nutraceuticals compound

Nutraceutical compound	Actions	References
and target genes Curcumin		
SOD, GSH	Upregulate SOD expression in the liver Significantly increased reduced glutathione (GSH) levels in diabetic rats, highlighting its ability to enhance antioxidant defenses. Activate the Nrf2 pathway, which protects β -cells from oxidative damage.	Xia et al. 2020 ¹⁰³ Belhan et al. 2020 ¹⁰¹ Serafini et al. 2020 ¹⁰⁵ Shahcheraghi et al. 2021 ¹⁰⁶
Kaempferol (flavonoid) SOD1, SOD2. GSH- Px IRS-1, PI3K, PIP2, PIP3, AKT, GLUT4	Upregulates the expression of SOD1 and SOD2 genes Increasing glutathione peroxidase (GSH-Px) activity, enhancing intracellular GSH levels, and promoting the nuclear translocation of nuclear factor erythroid 2-related factor 2 (Nrf2), which regulates the expression of antioxidant genes. Enhances insulin signaling in diabetic conditions by modulating IRS-1 gene expression, upregulates IRS-1 in skeletal muscle, activating the insulin receptor pathway. This leads to PI3K activation, PIP3 production, and subsequent activation of AKT, which phosphorylates downstream targets, including GLUT4. The process facilitates GLUT4 translocation to the plasma membrane, improving	Kluska et al. 2022 ¹²³ Al-Abbasi and Kazmi, 2022 ¹²² Alshehri et al. 2021 ¹²⁴ Jayachandran et al, 2020 ¹²⁰ Moore at al. 2023 ¹⁰²
SIRT1	glucose uptake into muscle cells and enhancing cellular glucose utilization. Modulates the SIRT1 pathway	Chong et al., 2024 ¹¹¹
Cinnamon	115 datation and 21111 patients	enong et un, 202 :
PI3K, AKT, PPARy	Regulating the expression of genes related to the insulin signaling pathway (PI3K and AKT), and enhancing the expression of PPARγ genes, which can reduce insulin resistance.	Cortez-Navarrete et al. 2023 ¹⁰⁴
Saponin		
SOD1	Activate SOD1 gene expression.	Kim et al. 1996 ¹²⁵ Cui et al. 2021 ¹²⁶
TCF7F2, β-catenin	Increasing activities of SOD, GSH-Px and CAT in the 200 µg/mL alfalfa saponin protecting β cell survival and regeneration by mechanisms involving the activation of Wingless-related integration site (Wnt3a)/β-catenin/ transcription factor 7-like 2 (TCF7L2) signaling	Cui et al. 2020 ¹²⁷
Quercetin (flavonoids)		
SOD1, CAT, GPX1	Quercetin normalized the expression mRNA levels of CAT, SOD1, GPX1 to near the normal level. Moreover, quercetin treatment normalized TAC levels.	Bagheri et al. 2021 ¹⁸⁰
Sirt3	Protected islet β-cells from oxidation-induced apoptosis via Sirt3 in T2DM	Wang et al. 2021 ¹⁸¹

glutathione (GSH), glutathione peroxidase (GSH-Px), nuclear factor erythroid 2-related factor 2 (Nrf2), insulin receptor substrate (IRS), phosphatidylinositol 3-kinase (PI3K), protein kinase B (Akt), Akt substrate of 160 kDa (AS160), glucose transporter 4 (GLUT4), phosphatidylinositol-3,4-bisphosphate (PIP2), phosphatidylinositol-3,4,5-trisphosphate (PIP3), sirtuin 1 (SIRT1), superoxide dismutase (SOD), peroxisome proliferator-activated receptor gamma (PPAR γ), Wingless-related integration site (Wnt3a)/ β -catenin/ transcription factor 7-like 2 (TCF7L2), catalase (CAT), protein kinase B (PKB), glucose transporter-2 (GLUT-2), high glucose (HG), endoplasmic reticulum (ER), cyclin-dependent kinase 5 (CDK5), pancreatic duodenal homeobox 1 (PDX1), sarcoendoplasmic reticulum calcium ATPase 2b (SERCA2b), nitric oxide (NO), nuclear factor kappa-light-chain-enhancer of activated B (NF- κ B), tumor necrosis factor alpha (TNF- α), interleukin (IL), intercellular adhesion molecule 1 (ICAM-1), vascular cell adhesion protein 1 (VCAM-1), transforming growth factor beta (TGF- β), AMP-activated protein kinase (AMPK), phosphorylated transducer of regulated CREB 2 (p-TORC2), uncoupling protein 2 (UCP2), insulinoma cell line (INS-1E).

Table 2. Modulation of gene expression in diabetic conditions by traditional nutraceuticals compound (cont.)

Nutraceutical compound	Actions	References
and target genes		
Myricetin		
(flavonoid)		
PKB, IRS1, IRS2, GLUT2, GLUT4	Normalized the insulin signaling molecule expression like PKB (protein kinase B), IRS-1 (insulin receptor-1), IRS-2 (insulin receptor-2), GLUT-2 (glucose transporter-2) and GLUT-4 (glucose transporter-4)	Kandasamy et al. 2014 ¹⁸²
CDK5, PDX1, SER-	Protects β-cells from high glucose (HG)-induced apoptosis by mitigating endoplasmic reticulum (ER) stress, potentially via cyclin-	Karunakaran et al. 2019 ¹⁸³
CA2b	dependent kinase 5 (CDK5) inhibition, leading to the upregulation of pancreatic duodenal homeobox 1 (PDX1) and sarcoendoplasmic reticulum calcium ATPase 2b (SERCA2b).	
Anthocyanin		
·	increases nitric oxide (NO) bioactivity and upregulation of nuclear factor erythroid 2-related factor 2 (Nrf2), which induces the production of endogenous antioxidants and limits oxidative stress, proposed to downregulate nuclear factor kappa-light-chain-enhancer of activated B (NF-κB) and lead to reduced expression and production of cytokines involved (tumor necrosis factor alpha (TNF-α), interleukin (IL)-1β, IL-6, intercellular adhesion molecule 1 (ICAM-1), vascular cell adhesion protein 1 (VCAM-1)) and further reduces the inflammatory response. NF-κB downregulates transforming growth factor beta (TGF-β) expression	Sapian et al. 2022 ¹¹²
Terpenoid		
IRS-1, Akt, GLUT4	Up-regulated the expression of the insulin receptor, insulin receptor substrate 1, glycogen synthase kinase 3β, Akt serine/ threonine kinase, and the transcript levels of GLUT4 and AMP-activated protein kinase.	Singh et al. 2022 ¹²¹
Alkaloids	· · · · · · · · · · · · · · · · · · ·	
p-AMPK, p-TORC2,	Up-regulated protein expression of liver kinase, AMPK, p-AMPK and p-TORC2. Down-regulated protein expression of gluconeo-	Dange et al. 2016 ¹¹³
SOD1, UCP2	genic Enzymes Inhibited high glucose-elevated nitrotyrosine level, reduced SOD-1 and UCP2 expression and AMPK phosphorylation in INS-1E cells	Dong et al. 2016 ¹⁸⁴ Jiang et al. 2015 ¹²⁸
Gallic acid	tion in 185-1E cens	Jiang et al. 2013
PPARy, GLUT4, Akt	Concurrent activation of PPAR- γ and C/EBP promotes GLUT4 translocation in adipose cells; improves insulin sensitivity through regulation of Akt and AMPK signaling pathways; regulation of TNF- α and adipocytokine expression; and improves β -cell function by inhibiting caspase-9-related to cell apoptosis.	Xu et al. 2021 ¹¹⁴

glutathione (GSH), glutathione peroxidase (GSH-Px), nuclear factor erythroid 2-related factor 2 (Nrf2), insulin receptor substrate (IRS), phosphatidylinositol 3-kinase (PI3K), protein kinase B (Akt), Akt substrate of 160 kDa (AS160), glucose transporter 4 (GLUT4), phosphatidylinositol-3,4-bisphosphate (PIP2), phosphatidylinositol-3,4,5-trisphosphate (PIP3), sirtuin 1 (SIRT1), superoxide dismutase (SOD), peroxisome proliferator-activated receptor gamma (PPARγ), Wingless-related integration site (Wnt3a)/β-catenin/ transcription factor 7-like 2 (TCF7L2), catalase (CAT), protein kinase B (PKB), glucose transporter—2 (GLUT–2), high glucose (HG), endoplasmic reticulum (ER), cyclin-dependent kinase 5 (CDK5), pancreatic duodenal homeobox 1 (PDX1), sarcoendoplasmic reticulum calcium ATPase 2b (SERCA2b), nitric oxide (NO), nuclear factor kappa-light-chain-enhancer of activated B (NF-κB), tumor necrosis factor alpha (TNF-α), intercellular adhesion molecule 1 (ICAM-1), vascular cell adhesion protein 1 (VCAM-1), transforming growth factor beta (TGF-β), AMP-activated protein kinase (AMPK), phosphorylated transducer of regulated CREB 2 (p-TORC2), uncoupling protein 2 (UCP2), insulinoma cell line (INS-1E).

Galohgor as an antidiabetic has been proven to reduce hyperglycemia, oxidative stress, adipose tissue, and body weight in T2DM.^{54,55}

Bir pletok

The traditional Indonesian herbal drink known as Bir Pletok, originating from the Betawi tribe, is renowned for its therapeutic properties. Historically used for its warming and health-promoting properties, Bir Pletok is made from a blend of spices including ginger (Zingiber officinale), lemongrass (Cymbopogon citratus), pandan leaves (Pandanus amaryllifolius) and cinnamon (Cinnamomum verum). Gingerols, flavonoids and polyphenols are among the bioactive compounds in Bir Pletok, known for their ability to regulate glucose metabolism and provide antioxidant protection. Due to its abundance of bioactive chemicals with antidiabetic properties, there has recently been increased interest in its potential use as a nutraceutical for the management of diabetes. With its blend of conventional herbal components, Bir Pletok may have encouraging antidiabetic benefits. Numerous studies have confirmed the ability of Bir Pletok's main constituents to control blood sugar levels, improve insulin sensitivity and reduce inflammation.⁵⁶

Wedang uwuh

Indonesia's traditional herbal drink from Yogyakarta, Wedang Uwuh, is well known for its abundance of natural spices and herbs and for its health advantages. Literally translating to "herbal trash drink," "Wedang Uwuh" refers to the vibrant concoction of ingredients that include nutmeg (Myristica fragrans), ginger (Zingiber officinale), cinnamon (Cinnamomum verum), cloves (Syzygium aromaticum), Sappan wood (Caesalpinia sappan), and lemongrass (*Cymbopogon citratus*). For centuries, people have drunk this beverage because of its warming properties and potential health benefits. Wedang Uwuh has garnered interest lately due to its potential as a conventional nutraceutical, especially in the treatment of long-term conditions like diabetes. Consequently, there is a growing interest in supplementary approaches to diabetes management, such as Wedang Uwuh, which are natural remedies. Bioactive substances like polyphenols, flavonoids, and essential oils found in the herbs and spices of Wedang Uwuh have been demonstrated to boost insulin sensitivity, lessen oxidative stress, and improve glucose metabolism.57

Loloh Bali

The herbal liquids known as *Loloh* are made and drunk only in Bali, Indonesia, and are used to treat and prevent a variety of illnesses. There are 51 plant species in all, from 32 families, that have been identified in the different Loloh preparations. Loloh is made from a variety of plants and plant components that are used to treat various ailments. The most common ways to prepare these plants are as decoctions, juices, or just as ingredients. *Alstonia scholaris (L.) R. Br., Blumea balsamifera (L.) DC., Cinnamomum burmanni Nees ex Bl.*, and *Piper betle L.* are the plants that are most frequently mentioned (>30 informants). The pharmacological effects of these well-

researched plants, such as their antibacterial, anticancer, and antidiabetic properties, have been reported.⁵⁸

Traditional Chinese Medicine

For thousands of years, traditional Chinese medicine (TCM) has been used to treat a wide range of illnesses, including metabolic diseases like diabetes. Traditional nutraceuticals, which are natural products made from medicinal plants and herbs, have drawn a lot of attention due to their potential therapeutic benefits among their various therapies. In addition to offering vital nutrients, these nutraceuticals also include bioactive substances that have positive effects on health, especially when it comes to managing chronic conditions like diabetes. The utilization of medicinal characteristics of herbs that have been demonstrated to control blood glucose levels, enhance insulin sensitivity, and lowering oxidative stress is how traditional Chinese nutraceuticals provide an alternative method. Preclinical and clinical research has shown encouraging antidiabetic effects for some TCM-based nutraceuticals. Herbs such as Momordica charantia, Berberis aristata, and Panax ginseng contain compounds that have been demonstrated to control glucose metabolism, boost pancreatic function, and reduce inflammation. Research on the discovery and characterization of these bioactive components from conventional nutraceuticals is still ongoing, intending to create safer and more efficient diabetes treatment therapies.⁵⁹

Traditional nutraceuticals from Malaysia

Malaysia, a country renowned for its varied natural vegetation and rich cultural legacy, has long used traditional treatments for a range of illnesses, including diabetes. Traditional nutraceuticals, products derived from natural sources such as herbs and plants that provide both nutritional and therapeutic benefits, have gained popularity due to the nation's rich diversity of medicinal plants. These age-old cures, which are frequently handed down through the generations, have drawn interest because of their possible application in the treatment of diabetes, an increasingly widespread health issue. Bioactive compounds found in traditional Malaysian nutraceuticals Gynura procumbens (Sabung Nyawa), and Curcuma longa (Turmeric) have been shown to potentially improve insulin sensitivity, lower blood sugar levels, and reducing the complications associated with diabetes. These therapeutic plants are currently being researched for their unique antidiabetic properties. Historically, they have been utilized to treat a wide range of illnesses. 60,61

Traditional Thai medicine

Herbal medicine in Thailand has become popular in recent years through the Thai Traditional Medicine (TTM) revitalization program. The Thai government has made various efforts to accelerate research on plants in Thailand that have nutraceutical content for various diseases, including diabetes mellitus. This is based on the habits of local rural communities in Thailand who use many herbal plants for diabetes mellitus. The nutraceutical contents of these herbal plants have been proven to exhibit a positive impact on the management of diabetes mellitus. Research conducted from the *in vitro* stage to clinical trials

on healthy subjects and patients with diabetes mellitus conditions has been proven to reduce glycated hemoglobin A1c (HbA1C), blood glucose levels, postprandial glucose levels, body weight, fat mass, insulin, lipids levels, oxidative stress, arterial stiffness, and endothelial dysfunction. Besides, TTM also improves insulin response, antioxidants, and β -cells function. Various TTMs are used ranging from aerial parts, 63 leaves, $^{64-74}$ fruits, $^{75-77}$ styles, 78 bark, leaf, 79 whole plant, $^{80-82}$ stems, 98 and rhizomes. $^{83-85}$

Different parts of the same plant often have similar phytochemical profiles, thus exhibiting identical pharmacological properties. Research continues to expand on the use of nutraceuticals in TTM for drug therapy, but the use in the form of food therapy also remains an option. For example, Thai papaya cultivar leaves is a vegetable commonly consumed by Thai people. Research has shown that three types of Thai papaya cultivar leaves have antidiabetic activity. Clinical trials on various commonly consumed food plants can increase consumption alternatives for the community to maintain health and management of non-communicable diseases including diabetes mellitus.

Various studies have proven that TTM can have a positive impact on DM conditions, including decreasing HbA1c, reducing blood glucose levels, increasing insulin response, and reducing oxidative damage in the brains of diabetic mice. These plants include Andrographis paniculata (Burm.f.) Nees, Pluchea indica (L.) Less, Apium graveolens L., Aloe vera (L.) Burm.f., Ipomoea aquatica Forssk., Lagerstroemia speciosa (L.) Pers. (bungur leaf), Terminalia bellirica (Gaertn.) Roxb., Terminalia chebula Retz., Momordica charantia L., Zea mays L., Morinda citrifolia L., Eclipta prostrata (L.), Phyllanthus amarus, Curcuma longa L., Gymnema inodorum (Lour.) Decne., Aegle marmelos (L.), Glycine max (L.) Merr, Salacia chinensis L., Artocarpus heterophyllus Lam., and Phyllanthus emblica L., Salacia chinensis L. (Table 1).

India herbal plants

The prevalence of diabetes mellitus in India is reported to be high, ⁸⁸ which is attributed to excess fat, low muscle mass and genetic factors of racial predisposition that increase the risk of T2DM in India. ⁸⁹ Treatment of T2DM using herbs is growing in developing countries including India. World Health Organization (WHO) data states that 90% of people in developing countries use plants as traditional medicine for health. ⁹⁰ India has 2500 species of herbal plants out of 21,000 plants worldwide registered by WHO. ⁹¹ The nutraceutical contents in various plants were studied and showed a positive impact on diabetes mellitus conditions, such as *Ficus religiosa*, *Pterocarpus marsupium*, *Gymnema sylvestre*, *Allium sativum*, *Eugenia jambolana*, *Momordica charantia*, and *Trigonella foenum-graecum* (Table 1). ⁹²

Various plants in India contain polyphenols, flavonoids, fatty acids, and fiber. 92-95 Ficus religiosa, which can be used for its flowers and stem bark, is known to have antidiabetic 96 and antihyperglycemic 97 effects which can reduce blood glucose levels and modulate antioxidant enzymes to fight oxidative stress. Consumption of Eugenia jambolana can affect antioxidant defenses, decrease glucose, increase insulin secretion, inhibit insulin degradation, and have hypoglycemic effects. 98 The parts of this plant that are commonly utilized are seeds, skin, fruit, and leaves. Momordica charantia is known to contain vicine, charantin, and triterpenoids along with several antioxidants, saponins 99 that have a positive impact on diabetic conditions. 100 Research at the clinical stage can continue to be developed to prove the potential of various nutraceuticals in Indian plants. Isolation, purification, and characterization of bioactive compounds contained therein can be used as a basis for the development of functional food products that are potential for the prevention and management of diabetes.

Nutraceutical compounds in traditional plants from Asia and their potential effect on diabetes mellitus are shown in Table 1. Some results from studies on the antidiabetic effects of Asian traditional nutraceutical metabolites include: 1) Curcuma longa⁶⁵ contains curcumin, curcumol, and bisdemethoxycurcumin, which help lower blood glucose and postprandial glucose levels while improving β -cell function; 2) Zea mays⁶¹ is rich in volatile oils, steroids, saponins, polysaccharides, alkaloids, flavonoids, organic acids, and phenolic compounds, has been shown to reduce blood glucose levels; 3) Glycine max¹⁰¹ contains isoflavones and anthocyanins, which are effective in lowering postprandial glucose levels; 4) Andrographis contains terpenoid glycosides, alkaloids, flavonoids, saponins, and tannins, which help reduce HbA1c levels: 4) Allium^{102,103} contains allicin, allixin, ajoene, and other organosulfur compounds that reduce fasting blood sugar levels, enhance insulin secretion by β -cells, and boost antioxidant defenses of the body; 5) Phyllanthus⁶⁰ family contains phenolic compounds (e.g., tannins, phenolic acids, flavonoids), alkaloids, phytosterols, terpenoids, organic acids, amino acids, and vitamins, which reduce oxidative stress.

Other beneficial effects of traditional nutraceuticals in diabetes include reducing lipid levels and enhancing antioxidant activity, as seen in plants from the Terminalia family.⁵⁷ Additionally, some plants improve insulin sensitivity and stimulate glucose uptake in peripheral tissues,¹⁰⁴ such as *Trigonella foenum-graecum*, which contains saponins, 4-hydroxyisoleucine, trigonelline, alkaloids, and steroids.⁶⁶ These studies demonstrate that traditional nutraceutical metabolites from Asia hold promise as alternative non-pharmacological therapies for managing diabetes.

Modulation of gene expression by traditional nutraceuticals

There is significant potential for nutraceuticals to modulate gene expression in the treatment of diabetes. However, this review will focus on three main mechanisms of gene expression modulation based on the primary pathophysiology of T2DM, i.e., impaired insulin secretion from beta cells (β -cell dysfunction), disrupted insulin signaling mechanisms, and abnormalities induced by oxidative stress exacerbating the diabetic condition.

Genes involved in \(\beta\)-cell dysfunction

Traditional nutraceuticals, derived from natural plants, herbs, and bioactive compounds, have emerged as promising agents in modulating gene expressions involved in β-cell dysfunction (Table 2). Compounds such as curcumin, resveratrol, kaempferol, and quercetin have shown the ability to regulate pathways linked to oxidative stress, inflammation, and apoptosis. For instance, curcumin has been reported to upregulate the Nrf2 pathway, which protect β -cells from oxidative damage. Similarly, resveratrol upregulate the sirtuin 1 (SIRT1) pathway, promoting β-cell survival and improving insulin secretion under stress conditions. 107,108 Resveratrol's inhibition of nuclear factor kappa-light-chain-enhancer of activated B cells (NF- κ B) operates to downregulate the pancreatic β cell apoptotic pathway resulting in improved β-cell survival. 109

In pancreatic β -cells, kaempferol has been found to enhance glucose-stimulated insulin secretion (GSIS) by activating the mitochondrial calcium uniporter (MCU) and increasing mitochondrial calcium uptake. This effect was validated in both insulinoma cell line (INS-1E) cells and human islets, suggesting that kaempferol can improve β -cell function under stress conditions. Interestingly, while kaempferol upregulate the SIRT1 pathway, other studies have identified alternative mechanisms for improving β -cell function and survival.

Anthocyanins, a subclass of flavonoids with potent antioxidant and anti-inflammatory properties, have been shown to upregulate NO bioactivity and Nrf2. This activation strengthens endogenous antioxidant defense mechanisms and mitigates oxidative stress, which is a key contributor to pancreatic β-cell dysfunction in diabetes. The increased Nrf2 activity leads to enhanced production of endogenous antioxidants, providing protection against oxidative damage. Concurrently, anthocyanins are proposed to downregulate NF-kB, a key regulator of inflammation. This suppression results in the downregulation of pro-inflammatory cytokines, TNF-α, IL-1β, IL-6, intercellular adhesion molecule 1 (ICAM-1), and vascular cell adhesion protein 1 (VCAM-1), ultimately reducing chronic low-grade inflammation—a hallmark of insulin resistance and diabetes progression. Additionally, since NFκB negatively regulates transforming growth factor-beta (TGF- β), its inhibition by anthocyanins indirectly upregulates TGF-β expression, contributing to improved cellular homeostasis and immune regulation. Through these mechanisms, anthocyanins play a crucial role in preserving pancreatic β-cell function, enhancing insulin sensitivity, and mitigating diabetes-related complications by reducing oxidative stress and inflammation.¹¹²

Alkaloids, a diverse group of naturally occurring bioactive compounds, have been shown to modulate key signaling pathways involved in glucose metabolism and oxidative stress regulation. These compounds upregulate the expression of liver kinase B1 (LKB1), AMPK, phosphorylated AMPK (p-AMPK), and phosphorylated transducer of regulated cyclic adenosine monophosphate responsive element-binding protein-2/CREB 2 (p-TORC2), which play critical roles in maintaining cellular energy balance and metabolic homeostasis.¹¹³ Gallic acid, a polyphenolic compound with strong antioxidant and anti-inflammatory

properties, has been found to enhance insulin sensitivity and β -cell function through multiple molecular mechanisms. It regulates the expression of TNF- α and adipocytokines, reducing inflammation associated with insulin resistance. Gallic acid also exerts a β -cell protective effect by inhibiting caspase-9-mediated apoptosis, which is essential for maintaining pancreatic function and preventing β -cell loss in diabetes.¹¹⁴

The therapeutic potential of these nutraceuticals lies in their ability to target multiple signaling pathways that influence β -cell health. By modulating genes involved in stress response, apoptosis, and insulin biosynthesis, traditional nutraceuticals offer a complementary approach to conventional diabetes treatments. Future research should focus on clinical studies to validate their efficacy and investigate the optimal doses and formulations for protecting and restoring β -cell function in diabetes. 115,116

Genes involved in the insulin signaling pathway

Insulin gene expression is influenced by nutraceuticals including flavonoids (Table 2). Flavonoids are now considered as indispensable components in various nutraceutical, pharmaceutical, medicinal and cosmetic applications. This is due to their anti-oxidative, anti-inflammatory, anti-mutagenic and anti-carcinogenic properties and their capacity to modulate enzyme function.¹¹⁷

Nutraceutical plants have shown significant potential in modulating gene expression in insulin signaling pathways, providing an alternative approach to diabetes management. Some of the herbs that have been extensively studied include turmeric (*Curcuma longa*), cinnamon (*Cinnamomum verum*), bitter melon (*Momordica charantia*), and guava leaf.¹¹⁸

Turmeric, with its main active compound curcumin, has been shown to improve insulin sensitivity by modulating the expression of genes involved in glucose and lipid metabolism. Studies show that cinnamon extract can upregulate the expression of genes related to insulin signaling pathways, including phosphoinositide 3-kinase (PI3K) and protein kinase B (AKT), which play an important role in glucose uptake by cells. In addition, cinnamon has also been shown to reduce insulin resistance by upregulating the expression of the Peroxisome Proliferator-Activated Receptor Gamma (PPAR)-γ gene, which is involved in lipid metabolism and insulin sensitivity. 104

Guava leaf extract (GLE), which contains the bioactive compound kaempferol, has the potential to improve insulin signaling under diabetic conditions by modulating insulin receptor substrate 1 (IRS-1) gene expression. In diabetic rats, GLE enhances glucose uptake in cells by upregulating IRS-1 gene expression in skeletal muscle, thereby activating insulin receptors. Activated IRS-1 stimulates the enzyme PI3K, which subsequently converts phosphatidylinositol 4,5-bisphosphate (PIP2) in the cell membrane into phosphatidylinositol 3,4,5-trisphosphate (PIP3). PIP3 acts as a signaling molecule that binds to and activates AKT, also known as PKB. Activated AKT phosphorylates several downstream targets, including proteins that regulate vesicles containing glucose transporter-4 (GLUT4). Phosphorylation by AKT induces upregulation of gene expression of GLUT4 (Solute carrier family 2, member 4 (SLC2A4)), causing the vesicles containing GLUT4 to translocate to the plasma cell membrane. GLUT4 is a glucose transporter found in intracellular vesicles, and once translocated to the cell membrane, it facilitates glucose entry into the cell muscle. ^{102,120} Figure 2 shows modulation of IRS and GLUT4 genes by bioactive compounds of traditional nutraceuticals, i.e. kaempferol, myricetin and terpenoids.

Terpenoids, a diverse class of naturally occurring compounds derived from isoprene units, are widely found in medicinal plants. These bioactive compounds have been shown to upregulate the expression of key proteins involved in insulin signaling and glucose metabolism, including the insulin receptor (IR) and IRS-1, which play crucial roles in enhancing insulin sensitivity. Additionally, terpenoids upregulate the expression of glycogen synthase kinase 3β (GSK-3β) and Akt serine/threonine kinase (Akt), both of which regulate glycogen synthesis and glucose uptake. Furthermore, terpenoids promote the transcription of GLUT4 and AMPK, facilitating glucose uptake into cells and improving energy homeostasis. Through these molecular mechanisms, terpenoids contribute to enhanced insulin sensitivity, improved glucose utilization, and potential therapeutic benefits in metabolic disorders such as diabetes. 121 Additionally, alkaloids have been reported to downregulate the expression of key gluconeogenic enzymes, thereby reducing hepatic glucose production.113

Gallic acid concurrently activates PPAR- γ and CCAAT/enhancer-binding protein (C/EBP), which facilitate GLUT4 translocation in adipose cells, thereby promoting glucose uptake. Additionally, gallic acid improves insulin sensitivity by modulating the Akt and AMPK signaling pathways, which are crucial for glucose homeostasis. 114

Modulation of antioxidant gene expression

SOD is a key antioxidant enzyme that catalyzes the conversion of superoxide radicals into hydrogen peroxide and

oxygen, thereby reducing oxidative damage. Traditional nutraceuticals, such as turmeric (Curcuma longa), which contains curcumin—a natural polyphenol—have demonstrated significant effects on upregulated SOD gene expression (Table 2). For example, in a study investigating the effects of curcumin on high-fat diet and streptozotocin-induced hyperglycemia and hyperlipidemia in rats, curcumin treatment was found to upregulate SOD expression in the liver. Furthermore, curcumin significantly increased reduced glutathione (GSH) levels in diabetic rats, highlighting its ability to enhance antioxidant defenses. Furthermore, curcumin significantly increased reduced glutathione (GSH) levels in diabetic rats, highlighting its ability to enhance antioxidant defenses.

Other traditional nutraceuticals, GLE which contain kaempferol as a primary secondary metabolite, have also shown promise in modulating antioxidant gene expression. In studies related to diabetes, kaempferol demonstrated the ability to increase the activity of glutathione peroxidase (GSH-Px) *in vivo*, indicating its potential to enhance the antioxidant defense system. 122 Additionally, research has shown that kaempferol at a concentration of 50 µg/mL modestly upregulates the expression of superoxide dismutase 1 (SOD1) and SOD2 genes. 123

As a flavonoid, kaempferol exerts its antioxidant effects by enhancing cellular GSH levels, which is achieved by increasing the nuclear translocation of Nrf2, a key regulator of antioxidant response genes. ¹²⁴ Collectively, these findings indicate that nutraceuticals such as curcumin and kaempferol hold significant potential in mitigating oxidative stress and improving antioxidant defenses through the modulation of key genes, including those encoding SOD and GSH-related enzymes.

Myricetin, a flavonoid compound commonly found in Asia traditional nutraceutical, has been shown to protect pancreatic β -cells from apoptosis induced by high glucose (HG) conditions. This antioxidant protective effect is primarily mediated through the attenuation of endoplasmic reticulum (ER) stress, potentially via the inhibition of cyclin-dependent kinase 5 (CDK5). Consequently, this pathway enhances the regulation of PDX1, a critical

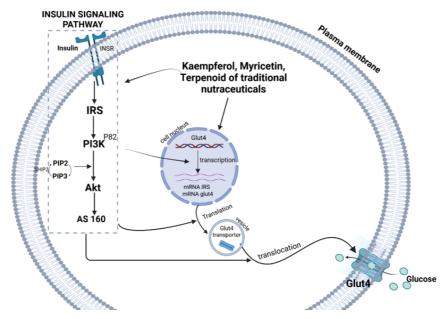


Figure 2. Modulation of IRS and GLUT4 genes by bioactive compounds of traditional nutraceuticals, i.e. kaempferol, myricetin and terpenoids. (This figure was generated using BioRender.com)

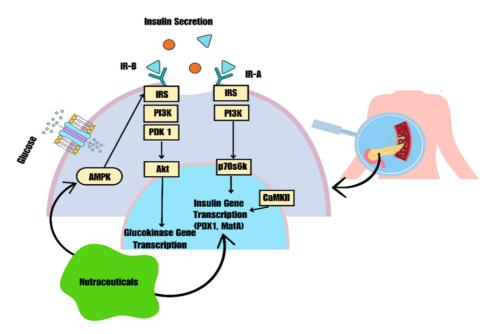


Figure 3. Nutraceuticals Modulation of PDX-1 and MafA Gene Expression Involved in Alleviating β -Cell Dysfunction. (This figure was generated using BioRender.com)

transcription factor for β -cell function and survival (Figure 3).

Saponins, a class of bioactive glycosides with strong antioxidant and cytoprotective properties, have been shown to upregulate SOD1 gene expression, thereby enhancing the endogenous antioxidant defense system. 125 Treatment with 200 µg/mL alfalfa-derived saponins significantly increases the enzymatic activities of SOD, GSH-Px, and CAT, which collectively mitigate oxidative stress and prevent cellular damage. 126 Furthermore, saponins play a critical role in preserving pancreatic β -cell viability and promoting β -cell regeneration through the activation of the Wingless-related integration site (Wnt3a)/ β -catenin/transcription factor 7-like 2 (TCF7L2) signaling pathway. This pathway is essential for maintaining β -cell function, enhancing proliferation, and regulating insulin secretion. 127

Alkaloids inhibit high glucose-induced nitro tyrosine accumulation, a marker of oxidative stress, while simultaneously downregulate SOD-1 and uncoupling protein 2 (UCP2) expression, which are linked to mitochondrial dysfunction. In INS-1E pancreatic β -cells, alkaloids prevent excessive AMPK phosphorylation, thereby protecting against cellular stress and dysfunction, highlighting their potential role in diabetes management. 128

Conclusion

Various traditional nutraceuticals from Asia have potential antidiabetic effects. These nutraceuticals contain antioxidants and exert their effect on modulating gene expressions involved in β -cell dysfunction, insulin signaling pathway, and antioxidant activity. This review highlights the growing scientific interest in exploring traditional nutraceuticals for the prevention and management of diabetes mellitus, offering potential alternatives to conventional treatments. However, further clinical trials are needed to confirm these antidiabetic effects.

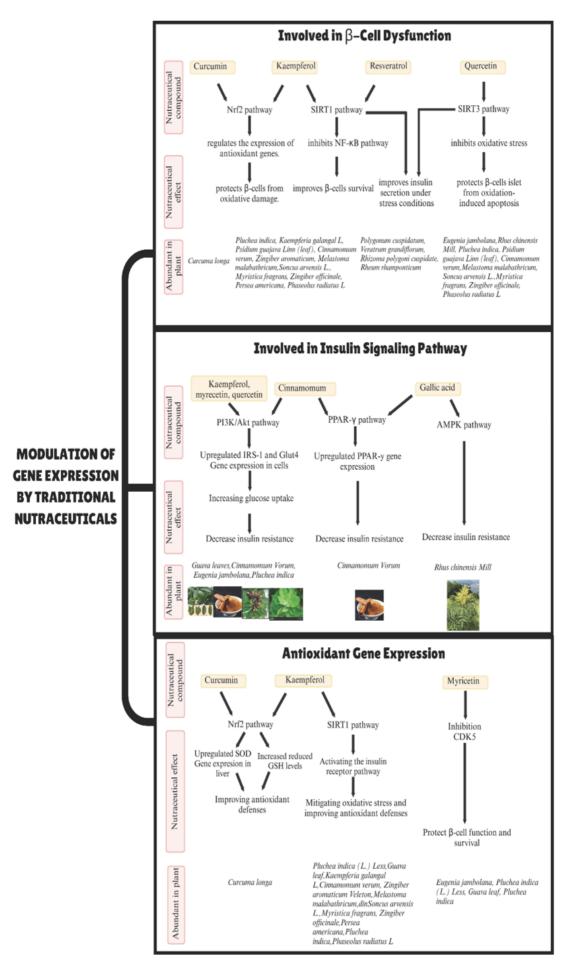
CONFLICT OF INTEREST AND FUNDING DISCLOSURES

The authors declare no conflict of interest.

This work was supported by Ministry of Education, Culture, Research, and Technology, Republic of Indonesia under the Regular Fundamental Research Grant [Grant No. 22049/IT3.D10/PT.01.03/P/B/2024].

REFERENCES

- Zheng Y, Ley SH, Hu FB. Global aetiology and epidemiology of type 2 diabetes mellitus and its complications. Nat Rev Endocrinol. 2018; 14:88–98. doi: 10.1038/nrendo.2017.151.
- Sun H, Saeedi P, Karuranga S, Pinkepank M, Ogurtsova K, Duncan BB, et al. IDF diabetes atlas: global, regional and country-level diabetes prevalence estimates for 2021 and projections for 2045. Diabetes Res Clin Pract. 2022; 183:109119. doi: 10.1016/j.diabres.2021.109119.
- 3. Ogurtsova K, da Rocha Fernandes JD, Huang Y, Linnenkamp U, Guariguata L, Cho NH, et al. IDF Diabetes Atlas: Global estimates for the prevalence of diabetes for 2015 and 2040. Diabetes Res Clin Pract. 2017; 128:40–50. doi: 10.1016/j.diabres.2017.03.024.
- 4. International Diabetes Federation. IDF diabetes atlas 10th edn. Brussels, Belgium; 2021.
- Soelistijo SA, Novida H, Rudijanto A, Soewondo P, Suastika K, Manaf A, et al. Konsensus Pengelolaan dan pencegahan diabetes melitus tipe 2 di Indonesia 2015 (Consensus on the Management and Prevention of Type 2 Diabetes Mellitus in Indonesia 2015). Pengurus Besar Perkumpulan Endokrinologi Indonesia (The Indonesian Society of Endocrinology 2015). (in Indonesian)
- Blahova J, Martiniakova M, Babikova M, Kovacova V, Mondockova V, Omelka R. Pharmaceutical drugs and natural therapeutic products for the treatment of type 2 diabetes mellitus. Pharmaceuticals. 2021; 14:806 doi: 10.3390/ph14080806.
- Venkatakrishnan K, Chiu H-F, Wang C-K. Popular functional foods and herbs for the management of type-2-diabetes mellitus: A comprehensive review with special reference to clinical trials and its proposed mechanism. J Funct Foods. 2019; 57:425–38. doi: 10.1016/j.jff.2019.04.039.



- Sharma A, Rani J, Kaur P, Dwivedi SK, Sharma M. Potential food nutraceutical ingredients bt handbook of nutraceuticals: science, technology and engineering. In: Rajakumari R, Thomas S, editors. Cham: Springer International Publishing; 2024. p. 1–44. doi://doi.org/10.1007/978-3-030-69677-1_2-1.
- Mali S, Rathod S, Kale N, Shinde N. Overview of nutraceuticals. Asian J Pharm Res. 2022; 61-70. doi: 10.52711/2231-5691.2022.00010.
- Willcox ML, Elugbaju C, Al-Anbaki M, Lown M, Graz B. Effectiveness of medicinal plants for glycaemic control in type 2 diabetes: an overview of meta-analyses of clinical trials. Front Pharmacol. 2021; 12:1-13. doi: 10.3389/fphar.2021.777561.
- 11. Pang G-M, Li F-X, Yan Y, Zhang Y, Kong L-L, Zhu P, et al. Herbal medicine in the treatment of patients with type 2 diabetes mellitus. Chin Med J (Engl). 2019; 132:78–85. doi: 10.1155/2013/343594.
- Gaonkar VP, Hullatti K. Indian Traditional medicinal plants as a source of potent anti-diabetic agents: A Review. Journal of Diabetes and Metabolic Disorders. 2020; 19:1895–908. doi: 10.1007/s40200-020-00628-8.
- 13. Handayani VA, Hutabarat EH. Elucidation of the Mechanism of Indonesian traditional medicine (jamu) based on case studies of type 2 diabetes networks. Int J Eng Manag Res. 2020; 10:87–91. doi: 10.31033/ijemr.10.1.16.
- 14. Unuofin JO, Lebelo SL. Antioxidant effects and mechanisms of medicinal plants and their bioactive compounds for the prevention and treatment of type 2 diabetes: an updated review. Oxid Med Cell Longev. 2020; 2020:1-36. doi: 10.1155/2020/1356893.
- 15. DeFronzo RA, Ferrannini E, Groop L, Henry RR, Herman WH, Holst JJ, et al. Type 2 diabetes mellitus. Nat Rev Dis Prim. 2015; 1:1–22. doi: 10.1038/nrdp.2015.19.
- Macdonald IA. A review of recent evidence relating to sugars, insulin resistance and diabetes. Eur J Nutr. 2016; 55:17–23. doi: 10.1007/s00394-016-1340-8.
- Bessesen DH. The role of carbohydrates in insulin resistance. J Nutr. 2001; 131:2782–6. doi: 10.1093/jn/131.10.2782s.
- Freeman AM, Pennings N. Insulin Resistance StatPearls -NCBI Bookshelf. StatPearls Publishing, Treasure Island (FL). 2021.
- Brun T, Maechler P. Beta-cell mitochondrial carriers and the diabetogenic stress response. Biochimica et Biophysica Acta
 Molecular Cell Research. 2016; 1863:2540-9. doi: 10.1016/j.bbamcr.2016.03.012.
- 20. Ashcroft FM, Rorsman P. KATP channels and islet hormone secretion: New insights and controversies. Nature Reviews Endocrinology. 2013; 9:660-9. doi: 10.1038/nrendo.2013.1 66.
- Eliasson L, Abdulkader F, Braun M, Galvanovskis J, Hoppa MB, Rorsman P. Novel aspects of the molecular mechanisms controlling insulin secretion. Journal of Physiology. 2008; 586:3313-24. doi: 10.1113/jphysiol.2008.155317.
- Brownlee M. The pathobiology of diabetic complications. Diabetes. 2005; 54:1615-25. doi:10.2337/diabetes.54.6.16
 15.
- Panov A V., Mayorov VI, Dikalov SI. Role of fatty acids βoxidation in the metabolic interactions between organs. Int J Mol Sci. 2024; 25:12740. doi: 10.3390/ijms252312740.
- 24. Johnson R, Dludla P, Joubert E, February F, Mazibuko S, Ghoor S, et al. Aspalathin, a dihydrochalcone C-glucoside, protects H9c2 cardiomyocytes against high glucose induced shifts in substrate preference and apoptosis. Mol Nutr Food Res. 2016; 60: 922-34. doi: 10.1002/mnfr.201500656.

- 25. Kang GG, Francis N, Hill R, Waters D, Blanchard C, Santhakumar AB. Dietary polyphenols and gene expression in molecular pathways associated with type 2 diabetes mellitus: a review. Int J Mol Sci. 2019; 21:140. doi: 10.3390/ijms21010140.
- 26. Jalleh RJ, Jones KL, Rayner CK, Marathe CS, Wu T, Horowitz M. Normal and disordered gastric emptying in diabetes: recent insights into (patho)physiology, management and impact on glycaemic control. Diabetologia. 2022; 62: 1981-93. doi: 10.1007/s00125-022-05796-1.
- 27. Lu J, Ma KL, Ruan XZ. Dysbiosis of gut microbiota contributes to the development of diabetes mellitus. ('Infect. Microbes Dis. 2019; 1: 43-8. doi: 10.1097/IM9.000000000000011.
- Manrique C, Lastra G, Sowers JR. New insights into insulin action and resistance in the vasculature. Ann N Y Acad Sci. 2014; 1311:138–50. doi: 10.1111/nyas.12395.
- Sharabi K, Tavares CDJ, Rines AK, Puigserver P. Molecular pathophysiology of hepatic glucose production. Mol Aspects Med. 2015; 46:21–33. doi: 10.1016/j.mam.2015.09.003.
- Nowotny K, Jung T, Höhn A, Weber D, Grune T. Advanced glycation end products and oxidative stress in type 2 diabetes mellitus. Biomolecules. 2015; 5:194–222. doi: 10.3390/biom5010194.
- Valko M, Leibfritz D, Moncol J, Cronin MTD, Mazur M, Telser J. Free radicals and antioxidants in normal physiological functions and human disease. Int J Biochem Cell Biol. 2007; 39:44–84. doi: 10.1016/j.biocel.2006.07.001.
- Rehman K, Akash MSH. Mechanism of generation of oxidative stress and pathophysiology of type 2 diabetes mellitus: how are they interlinked?. J Cell Biochem. 2017; 118:3577–85. doi: 10.1002/jcb.26097.
- Tangvarasittichai S. Oxidative stress, insulin resistance, dyslipidemia and type 2 diabetes mellitus. World J Diabetes. 2015; 6:456. doi: 10.4239/wjd.v6.i3.456.
- 34. Bhatti JS, Sehrawat A, Mishra J, Sidhu IS, Navik U, Khullar N, et al. Oxidative stress in the pathophysiology of type 2 diabetes and related complications: Current therapeutics strategies and future perspectives. Free Radic Biol Med. 2022; 184:114–34. doi: 1016/j.freeradbiomed.2022.03.019.
- Wang J, Wang H. Oxidative stress in pancreatic beta cell regeneration. Oxid Med Cell Longev. 2017; 1:1-9. doi: 10.1155/2017/1930261.
- 36. Robertson RP, Harmon JS. Pancreatic islet β -cell and oxidative stress: The importance of glutathione peroxidase. FEBS Letters. 2007; 581: 3743-8. doi: 10.1016/j.febslet.2007.03.087.
- 37. Newsholme P, Keane KN, Carlessi R, Cruzat V. Oxidative stress pathways in pancreatic β-cells and insulin-sensitive cells and tissues: Importance to cell metabolism, function, and dysfunction. Am J Physiol Cell Physiol. 2019; 317: C420-33. doi: 10.1152/ajpcell.00141.2019.
- 38. Robertson RP, Harmon JS. Diabetes, glucose toxicity, and oxidative stress: A case of double jeopardy for the pancreatic islet β cell. Free Radic Biol Med. 2006; 41:177–84. doi: 10.1016/j.freeradbiomed.2005.04.030.
- 39. Sharma A. The reduction of insulin gene transcription in HIT-T15 beta cells chronically exposed to high glucose concentration is associated with the loss of RIPE3b1 and STF-1 transcription factor expression. Mol Endocrinol. 1995; 9: 1127-34. doi: 10.1210/me.9.9.1127.
- 40. Tanaka Y, Gleason CE, Tran POT, Harmon JS, Robertson RP. Prevention of glucose toxicity in HIT-T15 cells and Zucker diabetic fatty rats by antioxidants. Proc Natl Acad Sci USA. 1999; 96: 10857-62. doi: 10.1073/pnas.96.19.10857.

- 41. Kaneto H, Miyatsuka T, Fujitani Y, Noguchi H, Song KH, Yoon KH, et al. Role of PDX-1 and MafA as a potential therapeutic target for diabetes. Diabetes Res Clin Pract. 2007; 77:S127-37. doi: 10.1016/j.diabres.2007.01.046.
- Baumel-Alterzon S, Scott DK. Regulation of Pdx1 by oxidative stress and Nrf2 in pancreatic beta-cells. Front. Endocrinol. 2022; 13:1-9. doi: 10.3389/fendo.2022.1011187.
- 43. Caturano A, D'Angelo M, Mormone A, Russo V, Mollica MP, Salvatore T, et al. Oxidative Stress in Type 2 Diabetes: Impacts from Pathogenesis to Lifestyle Modifications. Curr. issues mol. biol. 2023; 45:6651-66. doi: 10.3390/cimb45080420.
- 44. Liu CW, Wang YC, Hsieh CC, Lu HC, Chiang WD. Guava (Psidium guajava Linn.) leaf extract promotes glucose uptake and glycogen accumulation by modulating the insulin signaling pathway in high-glucose-induced insulin-resistant mouse FL83B cells. Process Biochem. 2015; 50:1128-35. doi: 10.1016/j.procbio.2015.03.022.
- 45. Saifudin A, Kadota S, Tezuka Y. Protein tyrosine phosphatase 1B inhibitory activity of Indonesian herbal medicines and constituents of Cinnamomum burmannii and Zingiber aromaticum. J Nat Med. 2013; 67:264-70. doi: 10.1007/s11418-012-0674-7.
- 46. Wayan Sudatri N, Wirasiti N, Gusti Nyoman Gde Bidura I, Made Suartini N. Anti-diabetic and anti-cholesterol activity of Kaempferia galanga L. herbal medicine rhizome in albino rats. Int. J. Fauna Biol. Stud. 2019; 6: 13-7.
- Truong LC. Reducing the effects of blood sugar infusion of Melastoma Malabathricum L. in mus musculus. J Asian Multicult Res Med Heal Sci Study. 2020; 1:1-10. doi: 10.47616/jamrmhss.v1i1.17.
- 48. Tandi J, Nyoman Edi Sutrisna I, Pratiwi M, Handayani TW. Potential test of nephropathy sonchus arvensis L. leaves on male rats (Rattus norvegicus) diabetes mellitus. Pharmacogn J. 2020; 12:1115-20. doi: 10.5530/pj.2020.12.158.
- Chowdhury MAR, . M, Haq MM. Phytochemical and pharmacological activity of myristica fragrans Houtt (myristicaceae). Int J Toxicol Pharmacol Res. 2017; 9:17–9. doi: 10.25258/ijtpr.v9i01.9038.
- Shidfar F, Rajab A, Rahideh T, Khandouzi N, Hosseini S, Shidfar S. The effect of ginger (Zingiber officinale) on glycemic markers in patients with type 2 diabetes. J Complement Integr Med. 2015; 12: 165-70. doi: 10.1515/jcim-2014-0021.
- 51. Kouamé NM, Koffi C, N'zoué KS, Yao NAR, Doukouré B, Kamagaté M. Comparative antidiabetic activity of aqueous, ethanol, and methanol leaf extracts of persea Americana and their effectiveness in type 2 diabetic rats. Evidence-based Complement Altern Med. 2019; 1-4. doi: 10.1155/2019/5984570.
- 52. Khatun MM, Sapon MA, Hossain MS, Islam MR. Antidiabetic activity of Piper betle in alloxan induced type 1 diabetic model Rrats. Int J Pharm Sci Res. 2016; 7: 675-80. doi: 10.13040/IJPSR.0975-8232.7.675-80.
- 53. Nopparat J, Nualla-Ong A, Phongdara A. Ethanolic extracts of Pluchea indica (L.) leaf pretreatment attenuates cytokineinduced β-cell apoptosis in multiple low-dose streptozotocin-induced diabetic mice. PLoS One. 2019; 14: 1-19. doi: 10.1371/journal.pone.0212133.
- 54. Roosita K, Kusharto CM, Sekiyama M, Fachrurozi Y, Ohtsuka R. Medicinal plants used by the villagers of a Sundanese community in West Java, Indonesia. J Ethnopharmacol. 2008; 115:72-81. doi: 10.1016/j.jep.2007.09.010.
- 55. Roosita K, Ma'rifah B, Marliyati SA, Firdaus F, Damayanti RP, Setyaningsih S, et al. Eksplorasi dan pengujian produk antidiabet: nutrasetikal galohgor. IPB Press. 2020.

- Silalahi M, Wahyuningtyas RS, Kalima T. Ethnobotanical study of Bir Pletok as a traditional health drink for Betawi ethnic (Indonesia). GSC Biol Pharm Sci. 2023; 24:335–42. doi: 10.30574/gscbps.2023.24.2.0285.
- 57. Setyowati N, Masyhuri, Mulyo JH, Irham, Yudhistira B. The hidden treasure of wedang uwuh, an ethnic traditional drink from Java, Indonesia: Its benefits and innovations. Int J Gastron Food Sci. 2023; 31:1-11. doi: 10.1016/j.ijgfs.2023.100688.
- 58. Sujarwo W, Keim AP, Savo V, Guarrera PM, Caneva G. Ethnobotanical study of Loloh: traditional herbal drinks from Bali (Indonesia). J Ethnopharmacol. 2015; 169:34–48. doi: 10.1016/j.jep.2015.03.079.
- 59. Ji L, Tong X, Wang H, Tian H, Zhou H, Zhang L, et al. Efficacy and safety of traditional Chinese medicine for diabetes: A double-blind, randomised, controlled trial. PLoS One. 2013; 8:1-10. doi: 10.1371/journal.pone.0056703.
- 60. Kim YJ. The current studies of education for a traditional and complementary medicine in Malaysia. J Evid Based Complementary Altern Med. 2017; 22:531–7. doi: 10.1177/2156587217726882.
- 61. Khan TM, Hassali MA, Al-Haddad MSM. Nutraceuticals Use among the inhabitants of Penang, Malaysia. Public Health. 2011; 3. Available: http://iomcworld.com/ijcrimph/ijcrimph-v03-n05-08.htm.
- 62. Peltzer K, Pengpid S, Puckpinyo A, Yi S, Anh LV. The utilization of traditional, complementary and alternative medicine for non-communicable diseases and mental disorders in health care patients in Cambodia, Thailand and Vietnam. BMC Complement Altern Med. 2016; 16:92. doi: 10.1186/s12906-016-1078-0.
- 63. Andrade C, Gomes NGM, Duangsrisai S, Andrade PB, Pereira DM, Valentão P. Medicinal plants utilized in thai traditional medicine for diabetes treatment: ethnobotanical surveys, scientific evidence and phytochemicals. J Ethnopharmacol. 2020; 263:1-54. doi: 10.1016/j.jep.2020.113177.
- 64. Alara OR, Abdurahman NH, Ukaegbu CI, Azhari NH. Vernonia cinerea leaves as the source of phenolic compounds, antioxidants, and anti-diabetic activity using microwave-assisted extraction technique. Ind. Crop. Prod. 2018;122:533-44. doi: 10.1016/j.indcrop.2018.06.034.
- 65. Yusni Y, Zufry H, Meutia F, Sucipto KW. The effects of celery leaf (apium graveolens L.) treatment on blood glucose and insulin levels in elderly pre-diabetics. Saudi Med J. 2018;39:154-60. doi: 10.15537/smj.2018.2.21238.
- 66. Fernando MR, Wickramasinghe N, Thabrew MI, Ariyananda PL, Karunanayake EH. Effect of Artocarpus heterophyllus and Asteracanthus longifolia on glucose tolerance in normal human subjects and in maturity-onset diabetic patients. J Ethnopharmacol. 1991;31:277-82. doi: 10.1016/0378-8741(91)90012-3.
- 67. Alinejad-Mofrad S, Foadoddini M, Saadatjoo SA, Shayesteh M. Improvement of glucose and lipid profile status with Aloe vera in pre-diabetic subjects: a randomized controlledtrial. J Diabetes Metab Disord. 2015;14:22. doi: 10.1186/s40200-015-0137-2.
- 68. Devaraj S, Yimam M, Brownell LA, Jialal I, Singh S, Jia Q. Effects of Aloe vera supplementation in subjects with prediabetes/metabolic syndrome. Metab Syndr Relat Disord. 2013;11:35-40. doi: 10.1089/met.2012.0066.
- 69. Huseini HF, Kianbakht S, Hajiaghaee R, Dabaghian FH. Anti-hyperglycemic and anti-hypercholesterolemic effects of Aloe vera leaf gel in hyperlipidemic type 2 diabetic patients: a randomized double-blind placebo-controlled clinical trial. Planta Med. 2012;78:311-6. doi: 10.1055/s-0031-1280474.

- Malalavidhane TS, Wickramasinghe SM, Perera MS, Jansz ER. Oral hypoglycaemic activity of Ipomoea aquatica in streptozotocin-induced, diabetic wistar rats and Type II diabetics. Phytother Res. 2003;17:1098-100. doi: 10.1002/ptr.1345.
- 71. Judy WV, Hari SP, Stogsdill WW, Judy JS, Naguib YM, Passwater R. Antidiabetic activity of a standardized extract (Glucosol) from Lagerstroemia speciosa leaves in Type II diabetics. A dose-dependence study. J Ethnopharmacol. 2003;87:115-7. doi: 10.1016/s0378-8741(03)00122-3.
- Chiabchalard A, Tencomnao T, Santiyanont R. Effect of Gymnema inodorum on postprandial peak plasma glucose levels in healthy human. Afr. J. Biotechnol. 2010; 9:1079-85; doi:10.5897/AJB09.1639
- MYM Ismail. Clinical Evaluation of Antidiabetic Activity of Bael Leaves. World Appl. Sci. J. 2009; 6: 1518-20.
- 74. Choi HC, Kim SJ, Son KY, Oh BJ, Cho BL. Metabolic effects of aloe vera gel complex in obese prediabetes and early non-treated diabetic patients: randomized controlled trial. Nutrition. 2013;29:1110-4. doi: 10.1016/j.nut.2013.02.015.
- 75. Singh N, Mahajan S, Subramani SK, Yadav D, Singh L, GBKS P. Triphala improves glucose homeostasis by alleviating atherogenic lipids and oxidative stress in human Type 2 diabetes mellitus. IJAM [Internet]. 2015. 25;6. https://www.ijam.co.in/index.php/ijam/article/view/0626201 5
- 76. John AJ, Cherian R, Subhash HS, Cherian AM. Evaluation of the efficacy of bitter gourd (momordica charantia) as an oral hypoglycemic agent--a randomized controlled clinical trial. Indian J Physiol Pharmacol. 2003;47:363-5.
- 77. Usharani P, Fatima N, Muralidhar N. Effects of Phyllanthus emblica extract on endothelial dysfunction and biomarkers of oxidative stress in patients with type 2 diabetes mellitus: A randomized, double-blind, controlled study. Diabetes, Metab Syndr Obes. 2013;6: 275–84. doi: 10.2147/DMSO.S46341.
- 78. Gomez EC. The Effects of Corn (Zea Mays) in The Dietary Management of Patients with Type 2 Diabetes Mellitus. In: Forum 2015 Abstracts [Internet]. British Medical Journal Publishing Group; 2015. Available from: https://bmjopen.bmj.com/lookup/doi/10.1136/bmjopen-2015-forum2015abstracts.113
- Algenstaedt P, Stumpenhagen A, Westendorf J. The Effect of Morinda citrifolia L. Fruit Juice on the Blood Sugar Level and Other Serum Parameters in Patients with Diabetes Type
 Evid Based Complement Alternat Med. 2018; 3565427. doi: 10.1155/2018/3565427.
- 80. Sazia S, Singh S, Shankar P, Nath R, Sachan AK, Dixit RK. Effects of Eclipta Alba and Diabetic Diet with Life Style Modifications on Blood Glucose Levels in Diabetic Patien. Int J Res Ayurveda Pharm [Internet]. 2015 Jun 30;6:375–8.
- 81. Moshi MJ, Lutale JJK, Rimoy GH, Abbas ZG, Josiah RM, Swai ABM. The effect of Phyllanthus amarus aqueous extract on blood glucose in non-insulin dependent diabetic patients. Phyther Res. 2001;15. doi: 10.1002/ptr.780.
- Srividya N, Periwal S. Diuretic, hypotensive and hypoglycaemic effect of Phyllanthus amarus. Indian J Exp Biol. 1995;33:861-4.
- 83. E. Y. Sukandar, H. Permana, I. K. Adnyana, J. I. Sigit, R. A. Ilyas, P. Hasimun, D. Mardiyah. Clinical Study of Turmeric (Curcuma longa L.) and Garlic (Allium sativum L.) Extracts as Antihyperglycemic and Antihyperlipidemic Agent in Type-2 Diabetes-Dyslipidemia Patients. Int. J. Pharmacol. 2010;6:456–463. doi: 10.3923/ijp.2010.456.463.
- 84. Srinivasan A, Selvarajan S, Kamalanathan S, Kadhiravan T, Prasanna Lakshmi NC, Adithan S. Effect of Curcuma longa on vascular function in native Tamilians with type 2 diabe-

- tes mellitus: A randomized, double-blind, parallel arm, placebo-controlled trial. Phytother Res. 2019;33:1898-911. doi: 10.1002/ptr.6381.
- Chuengsamarn S, Rattanamongkolgul S, Luechapudiporn R, Phisalaphong C, Jirawatnotai S. Curcumin extract for prevention of type 2 diabetes. Diabetes Care. 2012;35:2121-7. doi: 10.2337/dc12-0116.
- 86. Böttger A, Vothknecht U, Bolle C, Wolf A, Böttger A, Vothknecht U, Bolle C, Wolf A. Plant secondary metabolites and their general function in plants. in Lessons on Caffeine, Cannabis & Co, Learning Materials in Biosciences. Springer Nature. 2018. doi:10.1007/978-3-319-99546-5 1
- 87. Chaijan S, Chaijan M, Uawisetwathana U, Panya A, Phonsatta N, Shetty K, et al. Phenolic and Metabolic Profiles, Antioxidant Activities, Glycemic Control, and Anti-Inflammatory Activity of Three Thai Papaya Cultivar Leaves. Foods. 2024;13:1–19. doi: 10.3390/foods13111692.
- 88. Vaishya R, Misra A, Vaish A, Singh SK. Diabetes and tuberculosis syndemic in India: A narrative review of facts, gaps in care and challenges. J. Diabetes. 2024;16. doi: 10.1111/1753-0407.13427
- 89. Ali MK, Narayan KMV, Tandon N. Diabetes & coronary heart disease: Current perspectives. Indian J Med Res. 2010; 132:584-97. doi: 10.4103/IJMR.
- WHO. Traditional Medicine-Growing Needs and Potential.
 World Heal Organ Policy Perspect Med. 2002;2.
- Modak M, Dixit P, Londhe J, Ghaskadbi S, Devasagayam TPA. Indian herbs and herbal drugs used for the treatment of diabetes. J. Clin. Biochem. Nutr. 2007;40:163-73. doi: 10.3164/jcbn.40.163.
- 92. Rizvi SI, Mishra N. Traditional Indian medicines used for the management of diabetes mellitus. J Diabetes Res. 2013; 712092. doi: 10.1155/2013/712092.
- 93. Joshi DD, Deb L, Somkuwar BG, Rana VS. Relevance of Indian traditional tisanes in the management of type 2 diabetes mellitus: A review. Saudi Pharm J [Internet]. 2023;31:626–38. doi: 10.1016/j.jsps.2023.03.003.
- 94. Suthar P, Kumar S, Kumar V, Vaidya D, Sharma A, Sharma A. Murraya koenigii (L.) Spreng: Speculative ethnobotanical perspectives of ubiquitous herb with versatile nutra/functional properties. South African J Bot [Internet]. 2022;145:111–34. doi: 10.1016/j.sajb.2021.11.025.
- 95. Heirangkhongjam MD, Ngaseppam IS. Traditional medicinal uses and pharmacological properties of Rhus chinensis Mill.: A systematic review. Eur J Integr Med [Internet]. 2018;21:43–9. doi: 10.1016/j.eujim.2018.06.011.
- 96. Singh D, Singh B, Goel RK. Traditional uses, phytochemistry and pharmacology of Ficus religiosa: A review. J Ethnopharmacol. 2011;134:565-83. doi: 10.1016/j.jep.2011.01.046.
- 97. Deshmukh TA, Yadav B V., Badole SL, Bodhankar SL, Dhaneshwar SR. Antihyperglycaemic activity of petroleum ether extract of Ficus racemosa fruits in alloxan induced diabetic mice. Pharmacologyonline. 2007;2:504-15. Available from: https://pharmacologyonline.silae.it/files/archives/2007/vol2/47_Deshmukh.pdf
- Aybar MJ, Sánchez Riera AN, Grau A, Sánchez SS. Hypoglycemic effect of the water extract of Smallantus sonchifolius (yacon) leaves in normal and diabetic rats. J Ethnopharmacol. 2001;74:125-32. doi: 10.1016/S0378-8741(00)00351-2.
- Krawinkel MB, Keding GB. Bitter gourd (Momordica charantia): A dietary approach to hyperglycemia. Nutrition Reviews. 2006;64:331-7. doi: 10.1111/j.1753-4887.2006.tb00217.x.

- 100. Tsai CH, Chen ECF, Tsay HS, Huang CJ. Wild bitter gourd improves metabolic syndrome: A preliminary dietary supplementation trial. Nutr J. 2012;11:1-9. doi: 10.1186/1475-2891-11-4.
- 101. Belhan S, Yıldırım S, Huyut Z, Özdek U, Oto G, Algül S. Effects of curcumin on sperm quality, lipid profile, antioxidant activity and histopathological changes in streptozotocin-induced diabetes in rats. Andrologia. 2020;52:e13584. doi: 10.1111/and.13584.
- 102. Moore WT, Luo J, Liu D. Kaempferol improves glucose uptake in skeletal muscle via an AMPK-dependent mechanism. Food Sci Hum Wellness. 2023;12:2087-94.. doi: 10.1016/j.fshw.2023.03.028.
- 103. Xia ZH, Chen WB, Shi L, Jiang X, Li K, Wang YX, et al. The underlying mechanisms of curcumin inhibition of hyperglycemia and hyperlipidemia in rats fed a high-fat diet combined with STZ treatment. Molecules. 2020;25:271. doi: 10.3390/molecules25020271.
- 104. Cortez-Navarrete M, Pérez-Rubio KG, Escobedo-Gutiérrez M de J. Role of Fenugreek, Cinnamon, Curcuma longa, Berberine and Momordica charantia in Type 2 Diabetes Mellitus Treatment: A Review. Pharmaceuticals. 2023;16:515. doi: 10.3390/ph16040515.
- 105. Serafini MM, Catanzaro M, Fagiani F, Simoni E, Caporaso R, Dacrema M, et al. Modulation of Keap1/Nrf2/ARE signaling pathway by curcuma- And garlic-derived hybrids. Front Pharmacol. 2020;10:1597. doi: 10.3389/fphar.2019.01597.
- 106. Shahcheraghi SH, Salemi F, Peirovi N, Ayatollahi J, Alam W, Khan H, et al. Nrf2 regulation by curcumin: Molecular aspects for therapeutic prospects. Molecules. 2022;27:167. doi: 10.3390/molecules27010167.
- 107. Cai M, Wang J, Sun H, Guo Q, Zhang C, Yao H, et al. Resveratrol Attenuates Hydrogen Peroxide-induced Injury of Rat Ovarian Granulosa-lutein Cells by Resisting Oxidative Stress via the SIRT1/Nrf2/ARE Signaling Pathway. Curr Pharm Des. 2023;29:947-56. doi: 10.2174/1381612829666230403133322.
- 108. Zhu X, Ouyang W, Lan Y, Xiao H, Tang L, Liu G, et al. Anti-hyperglycemic and liver protective effects of flavonoids from Psidium guajava L. (guava) leaf in diabetic mice. Food Biosci. 2020;35:100574. doi: 10.1016/j.fbio.2020.100574.
- 109. Nanjan MJ, Betz J. Resveratrol for the management of diabetes and its downstream pathologies. Eur Endocrinol. 2014;10:31. doi: 10.17925/EE.2014.10.01.31.
- 110. Bermont F, Hermant A, Benninga R, Chabert C, Jacot G, Santo-Domingo J, et al. Targeting mitochondrial calcium uptake with the natural flavonol kaempferol, to promote metabolism/secretion coupling in pancreatic β-cells. Nutrients. 2020;12:538. doi: 10.3390/nu12020538.
- 111. Chong ACN, Vandana JJ, Jeng G, Li G, Meng Z, Duan X, et al. Checkpoint kinase 2 controls insulin secretion and glucose homeostasis. Nat Chem Biol. 2024;20:566-76. doi: 10.1038/s41589-023-01466-4.
- 112. Sapian S, Taib IS, Katas H, Latip J, Zainalabidin S, Hamid ZA, et al. The Role of Anthocyanin in Modulating Diabetic Cardiovascular Disease and Its Potential to Be Developed as a Nutraceutical. Pharmaceuticals. 2022;15:1344. doi: 10.3390/ph15111344.
- 113. Shende SS, Tilak A V. An Observational Study of the Anti-diabetic Activity of Berberine in Newly Diagnosed Type 2 Diabetes Mellitus Patients. 2016;6:230-3. doi: 10.20936/jpbms/160228.
- 114. Xu Y, Tang G, Zhang C, Wang N, Feng Y. Gallic acid and diabetes mellitus: Its association with oxidative stress. Molecules. 2021;26:7115. doi: 10.3390/molecules26237115.

- 115. Dama A, Shpati K, Daliu P, Dumur S, Gorica E, Santini A. Targeting Metabolic Diseases: The Role of Nutraceuticals in Modulating Oxidative Stress and Inflammation. Nutrients. 2024;16:507. doi: 10.3390/nu16040507.
- 116. Schaid MD, Zhu Y, Richardson NE, Patibandla C, Ong IM, Fenske RJ, et al. Systemic metabolic alterations correlate with islet-level prostaglandin e2 production and signaling mechanisms that predict β-cell dysfunction in a mouse model of type 2 diabetes. Metabolites. 2021;11:58. doi: 10.3390/metabo11010058.
- 117. Shukla R, Pandey V, Vadnere GP, Lodhi S. Role of Flavonoids in Management of Inflammatory Disorders. In: Bioactive Food as Dietary Interventions for Arthritis and Related Inflammatory Diseases [Internet]. 2nd ed. Elsevier; 2019. p. 293–322. doi: 10.1016/B978-0-12-813820-5.00018-0.
- 118. Mahdavi A, Bagherniya M, Mirenayat MS, Atkin SL, Sahebkar A. Medicinal Plants and Phytochemicals Regulating Insulin Resistance and Glucose Homeostasis in Type 2 Diabetic Patients: A Clinical Review. Adv Exp Med Biol. 2021; 1308:161-83. doi: 10.1007/978-3-030-64872-5_13.
- 119. Liu Y, Pang D, Xing D, Wang W, Li Q, Liao S, et al. Cinnamon free phenolic extract regulates glucose absorption in intestinal cells by inhibiting glucose transporters. Food Biosci. 2023;52:102405. doi: 10.1016/j.fbio.2023.102405.
- 120. Jayachandran M, Vinayagam R, Xu B. Guava leaves extract ameliorates STZ induced diabetes mellitus via activation of PI3K/AKT signaling in skeletal muscle of rats. Mol Biol Rep. 2020;47:2793-9. doi: 10.1007/s11033-020-05399-2.
- 121. Singh S, Bansal A, Singh V, Chopra T, Poddar J. Flavonoids, alkaloids and terpenoids: a new hope for the treatment of diabetes mellitus. J Diabetes Metab Disord. 2022;21:941-50. doi: 10.1007/s40200-021-00943-8.
- 122. Al-Abbasi FA, Kazmi I. Therapeutic role of kaempferol and myricetin in streptozotocin-induced diabetes synergistically via modulation in pancreatic amylase, glycogen storage and insulin secretion. Mol Cell Biochem. 2023;478:1927-37. doi: 10.1007/s11010-022-04629-4.
- 123. Kluska M, Juszczak M, Żuchowski J, Stochmal A, Woźniak K. Effect of Kaempferol and Its Glycoside Derivatives on Antioxidant Status of HL-60 Cells Treated with Etoposide. Molecules. 2022;27:333. doi: 10.3390/molecules27020333.
- 124. Alshehri AS, El-Kott AF, Eleawa SM, El-Gerbed MSA, Khalifa HS, El-Kenawy AE, et al. Kaempferol protects against streptozotocin-induced diabetic cardiomyopathy in rats by a hypoglycemic effect and upregulating sirt1. J Physiol Pharmacol. 2021;72. doi: 10.26402/jpp.2021.3.04.
- 125. Kim HS, Kim KS, Oh KW. Ginseng total saponin inhibits nicotine-induced hyperactivity and conditioned place preference in mice. J Ethnopharmacol. 1999;66:83-90. doi: 10.1016/s0378-8741(98)00192-5.
- 126. Cui Y, Li F, Zhu X, Xu J, Muhammad A, Chen Y, et al. Alfalfa saponins inhibit oxidative stress-induced cell apoptosis through the MAPK signaling pathway. Redox Rep. 2022;27:1-8. doi: 10.1080/13510002.2021.2017681..
- 127. Cui J, Duan J, Chu J, Guo C, Xi M, Li Y, et al. Chikusetsu saponin IVa protects pancreatic β cell against intermittent high glucose-induced injury by activating Wnt/β-catenin/TCF7L2 pathway. Aging (Albany NY). 2020 Jan 22;12:1591-609. doi: 10.18632/aging.102702.
- 128. Jiang SJ, Dong H, Li J Bin, Xu LJ, Zou X, Wang KF, et al. Berberine inhibits hepatic gluconeogenesis via the LKB1-AMPK-TORC2 signaling pathway in streptozotocin-induced diabetic rats. World J Gastroenterol. 2015;21:7777-85. doi: 10.3748/wjg.v21.i25.7777.
- 129. Leibiger B, Leibiger IB, Moede T, Kemper S, Kulkarni RN, Kahn CR, et al. Selective insulin signaling through A and B insulin receptors regulates transcription of insulin and glu-

- cokinase genes in pancreatic β cells. Mol Cell. 2001;7:559–70. doi: 10.1016/s1097-2765(01)00203-9.
- 130. Bnouham M, Ziyyat A, Mekhfi H, Tahri A, Legssyer A. Medicinal plants with potential antidiabetic activity A review of ten years of herbal medicine research (1990-2000). Int. j. diabetes metab. 2006;14: 1–25. https://doi.org/10.1159/000497588.
- 131. Ayodhya S, Kusum S, Anjali S. Hypoglycaemic Activity of Different Extracts of Various Herbal Plants. Int J Res Ayurveda Pharm. 2010;1:212-24.
- 132. Ravi K, Rajasekaran S, Subramanian S. Antihyperlipidemic effect of Eugenia jambolana seed kernel on streptozotocininduced diabetes in rats. Food Chem Toxicol. 2005;43:1433-9. doi: 10.1016/j.fct.2005.04.004.
- 133. Grover JK, Vats V, Rathi SS. Anti-hyperglycemic effect of Eugenia jambolana and Tinospora cordifolia in experimental diabetes and their effects on key metabolic enzymes involved in carbohydrate metabolism. J Ethnopharmacol. 2000;73:461-70. doi: 10.1016/s0378-8741(00)00319-6.
- 134. Sagrawat H, Mann A, Kharya M. Pharmacological potential of Eugenia jambolana: A review. Pharmacogenesis Magazice, Vol. 2, 2006, pp. 96-104.
- 135. Ravi K, Ramachandran B, Subramanian S. Effect of Eugenia Jambolana seed kernel on antioxidant defense system in streptozotocin-induced diabetes in rats. Life Sci. 2004;75:2717-31. doi: 10.1016/j.lfs.2004.08.005.
- 136. Leung L, Birtwhistle R, Kotecha J, Hannah S, Cuthbertson S. Anti-diabetic and hypoglycaemic effects of Momordica charantia (bitter melon): A mini review. Br J Nutr. 2009;102:1703-8. doi: 10.1017/S0007114509992054.
- 137. Saxena A, Vikram NK. Role of Selected Indian Plants in Management of Type 2 Diabetes: A Review. J Altern Complement Med. 2004;10:369-78. doi: 10.1089/107555304323062365.
- 138. Wang ZQ, Zhang XH, Yu Y, Poulev A, Ribnicky D, Floyd ZE, et al. Bioactives from bitter melon enhance insulin signaling and modulate acyl carnitine content in skeletal muscle in high-fat diet-fed mice. J Nutr Biochem. 2011;22:1064-73. doi: 10.1016/j.jnutbio.2010.09.004.
- 139. Chaturvedi P. Antidiabetic potentials of momordica charantia: Multiple mechanisms behind the effects. J Med Food. 2012;15:101-7. doi: 10.1089/jmf.2010.0258.
- 140. Mahomoodally MF, Subratty AH, Gurib-Fakim A, Choudhary MI, Nahar Khan S. Traditional medicinal herbs and food plants have the potential to inhibit key carbohydrate hydrolyzing enzymes in vitro and reduce postprandial blood glucose peaks in vivo. Sci. World J. 2012;285284. doi: 10.1100/2012/285284.
- 141. Keller AC, Ma J, Kavalier A, He K, Brillantes AMB, Kennelly EJ. Saponins from the traditional medicinal plant Momordica charantia stimulate insulin secretion in vitro. Phytomedicine. 2011;19:32-7. doi: 10.1016/j.phymed.2011. 06.019.
- 142. Pattanayak P, Behera P, Das D, Panda S. Ocimum sanctum Linn. A reservoir plant for therapeutic applications: An overview. Pharmacogn Rev. 2010;4:95-105. doi: 10.4103/0973-7847.65323.
- 143. Narendhirakannan RT, Subramanian S, Kandaswamy M. Biochemical evaluation of antidiabetogenic properties of some commonly used Indian plants on streptozotocin-induced diabetes in experimental Clin Exp Pharmacol Physiol. 2006;33:1150-7. doi: 10.1111/j.1440-1681.2006.04507. x.
- 144. Hannan JMA, Marenah L, Ali L, Rokeya B, Flatt PR, Abdel-Wahab YHA. Ocimum sanctum leaf extracts stimulate insulin secretion from perfused pancreas, isolated islets and

- clonal pancreatic β -cells. J Endocrinol. 2006;189:127-36. doi: 10.1677/joe.1.06615.
- 145. Nair, Rathish; Kalariya, Tamanna; And Chanda, Sumitra. Antibacterial Activity of Some Selected Indian Medicinal Flora. Turk. J. Biol. 2005;29. Available at: https://journals.tubitak.gov.tr/biology/vol29/iss1/7
- 146. Singh PK, Baxi D, Banerjee S, Ramachandran A V. Therapy with methanolic extract of Pterocarpus marsupium Roxb and Ocimum sanctum Linn reverses dyslipidemia and oxidative stress in alloxan induced type I diabetic rat model. Exp Toxicol Pathol. 2012;64:441-8. doi: 10.1016/j.etp.2010.10.011.
- 147. Patil RN, Patil RY, Ahirwar B, Ahirwar D. Evaluation of antidiabetic and related actions of some Indian medicinal plants in diabetic rats. Asian Pac J Trop Med. 2011;4:20-3. doi: 10.1016/S1995-7645(11)60025-4.
- 148. Kameswara Rao B, Giri R, Kesavulu MM, Apparao C. Effect of oral administration of bark extracts of Pterocarpus santalinus L. on blood glucose level in experimental animals. J Ethnopharmacol. 2001;74:69-74. doi: 10.1016/s0378-8741(00)00344-5.
- 149. Grover JK, Vats V, Yadav SS. Pterocarpus marsupium extract (Vijayasar) prevented the alteration in metabolic patterns induced in the normal rat by feeding an adequate diet containing fructose as sole carbohydrate. Diabetes Obes Metab. 2005;7:414-20. doi: 10.1111/j.1463-1326.2005.00414.x.
- 150. Gupta R, Gupta RS. Effect of Pterocarpus marsupium in streptozotocin-induced hyperglycemic state in rats: comparison with glibenclamide. Diabetol. Croat. 2009;38:39-45.
- 151. Cordero-Herrera I, Martín MA, Bravo L, Goya L, Ramos S. Cocoa flavonoids improve insulin signalling and modulate glucose production via AKT and AMPK in HepG2 cells. Mol Nutr Food Res. 2013;57: 974-85. doi: 10.1002/mnfr.201200500
- 152. Vats V, Grover JK, Rathi SS. Evaluation of anti-hyperglycemic and hypoglycemic effect of Trigonella foe-num-graecum Linn, Ocimum sanctum Linn and Pterocarpus marsupium Linn in normal and alloxanized diabetic rats. J Ethnopharmacol. 2002;79:95-100. doi: 10.1016/s0378-8741(01)00374-9.
- 153. Singh AB, Tamarkar AK, Shweta, Narender T, Srivastava AK. Antihyperglycaemic effect of an unusual amino acid (4-hydroxyisoleucine) in C57BL/KsJ-db/db mice. Nat Prod Res. 2010;24:258-65. doi: 10.1080/14786410902836693.
- 154. Karthic R, Nagaraj S, Arulmurugan P, Seshadri S, Rengasamy R, Kathiravan K. Gymnema sylvestre R. Br. suspension cell extract show antidiabetic potential in Alloxan induced diabetic albino male rats. Asian Pac J Trop Biomed. 2012;2:S930-3. doi: 10.1016/S2221-1691(12)60339-6.
- 155. Drobiova H, Thomson M, Al-Qattan K, Peltonen-Shalaby R, Al-Amin Z, Ali M. Garlic increases antioxidant levels in diabetic and hypertensive rats determined by a modified peroxidase method. Evid Based Complement Alternat Med. 2011;2011:703049. doi: 10.1093/ecam/nep011
- 156. Diniz YS, Rocha KKHR, Souza GA, Galhardi CM, Ebaid GMX, Rodrigues HG, et al. Effects of N-acetylcysteine on sucrose-rich diet-induced hyperglycaemia, dyslipidemia and oxidative stress in rats. Eur J Pharmacol. 2006;543:151-7. doi: 10.1016/j.ejphar.2006.05.039.
- 157. Widjajakusuma EC, Jonosewojo A, Hendriati L, Wijaya S, Ferawati, Surjadhana A, et al. Phytochemical screening and preliminary clinical trials of the aqueous extract mixture of Andrographis paniculata (Burm. f.) Wall. ex Nees and Syzygium polyanthum (Wight.) Walp leaves in metformin treated patients with type 2 diabetes. Phytomedicine. 2019;55:137-47. doi: 10.1016/j.phymed.2018.07.002.
- 158. Devaraj S, Jialal R, Jialal I, Rockwood J. A pilot randomized placebo controlled trial of 2 Aloe vera supplements in

- patients with pre-diabetes/metabolic syndrome. Planta Medica. 2008: 74. doi 10.1055/s-0028-1083957
- 159. Urita Y, Noda T, Watanabe D, Iwashita S, Hamada K, Sugimoto M. Effects of a soybean nutrition bar on the postprandial blood glucose and lipid levels in patients with diabetes mellitus. Int J Food Sci Nutr. 2012;63:921-9. doi: 10.3109/09637486.2012.694847.
- 160. Koteshwar P, Raveendra KR, Allan JJ, Goudar K, Goudar KS, Agarwal A. Effect of NR-Salacia on post-prandial hyperglycemia: A randomized double blind, placebo-controlled, crossover study in healthy volunteers. Pharmacogn Mag. 13;9:344-9. doi: 10.4103/0973-1296.117831.
- 161. Kumar M, Tomar M, Amarowicz R, Saurabh V, Sneha Nair M, Maheshwari C, et al. Guava (Psidium guajava I.) leaves: Nutritional composition, phytochemical profile, and health-promoting bioactivities. Foods. 2021;10:752. doi: 10.3390/foods10040752.
- 162. Eidenberger T, Selg M, Krennhuber K. Inhibition of dipeptidyl peptidase activity by flavonol glycosides of guava (Psidium guajava L.): A key to the beneficial effects of guava in type II diabetes mellitus. Fitoterapia. 2013;89:74-9. doi: 10.1016/j.fitote.2013.05.015.
- 163. Fujimori K, Shibano M. Avicularin, a plant flavonoid, suppresses lipid accumulation through repression of C/EBPα-activated GLUT4-mediated glucose uptake in 3T3-L1 cells. J Agric Food Chem. 2013;61:5139-47. doi: 10.1021/jf401154c.
- 164. Shabbir H, Kausar T, Noreen S, Rehman HU, Hussain A, Huang Q, et al. In vivo screening and antidiabetic potential of polyphenol extracts from guava pulp, seeds and leaves. Animals. 2020:10:1714. doi: 10.3390/ani10091714.
- Nair SS, Kavrekar V, Mishra A. In vitro studies on alpha amylase and alpha glucosidase inhibitory activities of selected plant extracts. Eur. J. Exp. Biol. 2013;3:128-32.
- 166. Wang SY, Zhao H, Xu HT, Han XD, Wu YS, Xu FF, et al. Kaempferia galanga L.: Progresses in Phytochemistry, Pharmacology, Toxicology and Ethnomedicinal Uses. Front. Pharmacol. 2021;12:675350. doi: 10.3389/fphar.2021.6753 50.
- 167. Wu QM, Jin YM, Ni HX. Effect of kaempferol on correlation factors of chronic complications of type 2 diabetic rats. Chinese Tradit Herb Drugs. 2015;46. doi: 10.7501/j.issn.0253-2670.2015.12.018.
- 168. Mutlu M, Bingol Z, Uc EM, Köksal E, Goren AC, Alwasel SH, et al. Comprehensive Metabolite Profiling of Cinnamon (Cinnamomum zeylanicum) Leaf Oil Using LC-HR/MS, GC/MS, and GC-FID: Determination of Antiglaucoma, Antioxidant, Anticholinergic, and Antidiabetic Profiles. Life (Basel). 2023;13:136. doi: 10.3390/life13010136.
- 169. Mohsin SN, Saleem F, Humayun A, Tanweer A, Muddassir A. Prospective Nutraceutical Effects of Cinnamon Derivatives Against Insulin Resistance in Type II Diabetes Mellitus—Evidence From the Literature. Dose-Response. 2023;21. doi: 10.1177/15593258231200527.
- 170. Mayasari D, Murti YB, Pratiwi SUT, Sudarsono S, Hanna G, Hamann MT. TLC-Based Fingerprinting Analysis of the Geographical Variation of Melastoma malabathricum in Inland and Archipelago Regions: A Rapid and Easy-to-Use Tool for Field Metabolomics Studies. J Nat Prod. 2022;85:292-300. doi: 10.1021/acs.inatprod.1c00622
- 171. Rafi M, Suwartiny NL, Rohaeti E. Traditional Use, Phytochemical Composition, and Biological Activities of Sonchus arvensis. Indonesian J Pharm [Internet]. 2022;33:540-53. https://jurnal.ugm.ac.id/v3/IJP/article/view/3823
- 172. Odubanjo VO, Olasehinde TA, Oyeleye SI, Oboh G, Boligon AA. Seed extracts from Myristica fragrans (Nutmeg)

- and Moringa oleifera (Drumstick tree) inhibits enzymes relevant to erectile dysfunction and metal-induced oxidative damage in rats' penile tissues. J Food Biochem. 2018;42. doi: 10.1111/jfbc.12452
- 173. Ozaki Y, Soedigdo S, Wattimena YR, Suganda AG. Antiinflammatory effect of mace, aril of Myristica fragrans Houtt., and its active principles. Jpn J Pharmacol. 1989;49:155-63. doi: 10.1254/jjp.49.155.
- 174. Ivane NMA, Elysé FKR, Haruna SA, Pride N, Richard E, Foncha AC, Dandago MA. The anti-oxidative potential of ginger extract and its constituent on meat protein isolate under induced Fenton oxidation. J Proteomics. 2022;269:104723. doi: 10.1016/j.jprot.2022.104723.
- 175. Park SJ, Nam YH, Rodriguez I, Park JH, Kwak HJ, Oh Y, et al. Chemical constituents of leaves of Persea americana (avocado) and their protective effects against neomycininduced hair cell damage. Rev Bras Farmacogn. 2019;29: 739–43. doi: 10.1016/j.bjp.2019.08.004
- 176. Nabar BM. Antibacterial and wound healing activity of piper betle extracts against multidrug resistant strains. Med. Plants - Int. J. Phytomed. Rel. Ind. 2017; 5. doi:10.5958/0975-6892.2017.00045.4
- 177. Aishah Baharuddin S, Nadiah Abd Karim Shah N, Saiful Yazan L, Abd Rashed A, Kadota K, Al-Awaadh AM, et al. Optimization of Pluchea indica (L.) leaf extract using ultrasound-assisted extraction and its cytotoxicity on the HT-29 colorectal cancer cell line. Ultrason Sonochem. 2023;101. doi: 10.1016/j.ultsonch.2023.106702. https://www.sciencedirect.com/science/article/pii/S1350417723004145
- 178. Poe ML, Bates A, Onyilagha J. Distribution of Leaf Flavonoid Aglycones and Glucuronides in the Genus Phaseolus and Related Genera. Int. J. Biol. 2013;5:4. doi: 10.5539/ijb.v5n4p36.
- 179. Ansari P, Tahseen Khan J, Choudhary S, Depheny Reberio A, Kumar S, Seidel V, et al. Plant-based Diets and Phytochemicals in the Management of Diabetes Mellitus and Prevention of its Complications: A Review. Nutrients. 2024;16:3709. doi: 10.3390/nu16213709.
- 180. Bagheri A, Ebrahimpour S, Nourbakhsh N, Talebi S, Esmaeili A. Protective effect of quercetin on alteration of antioxidant genes expression and histological changes in the dental pulp of the streptozotocin-diabetic rats. Arch Oral Biol. 2021;125:105088. doi: 10.1016/j.archoralbio.2021.105088.
- 181. Wang JY, Nie YX, Dong BZ, Cai ZC, Zeng XK, Du L, et al. Quercetin protects islet β-cells from oxidation-induced apoptosis via Sirt3 in T2DM. Iran J Basic Med Sci. 2021;24:629-35. doi: 10.22038/ijbms.2021.52005.11792..
- 182. Kandasamy N, Ashokkumar N. Renoprotective effect of myricetin restrains dyslipidemia and renal mesangial cell proliferation by the suppression of sterol regulatory element binding proteins in an experimental model of diabetic nephropathy. Eur J Pharmacol. 2014;743:53-62. doi: 10.1016/j.ejphar.2014.09.014
- 183. Karunakaran U, Elumalai S, Moon JS, Jeon JH, Kim ND, Park KG, et al. Myricetin protects against high glucose-induced β-cell apoptosis by attenuating endoplasmic reticulum stress via inactivation of cyclin-dependent kinase 5. Diabetes Metab J. 2019;43:192-205. doi: 10.4093/dmj.2018.0052.
- 184. Dong Y, Chen YT, Yang YX, Zhou XJ, Dai SJ, Tong JF, et al. Metabolomics Study of Type 2 Diabetes Mellitus and the AntiDiabetic Effect of Berberine in Zucker Diabetic Fatty Rats Using Uplc-ESI-Hdms. Phyther Res. 2016;30:823-8. doi: 10.1002/ptr.5587.