

Original Article

Association between maternal iron status and the risk of pre-eclampsia: a case-control study

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Background and Objectives: This study aimed to assess the associations of maternal iron status and placental iron transport proteins expression with the risk of pre-eclampsia (PE) in Chinese pregnant women. **Methods and Study Design:** A total of 94 subjects with PE and 112 healthy pregnant women were enrolled. Fasting blood samples were collected to detect maternal iron status. The placenta samples were collected at delivery to detect the mRNA and protein expression of divalent metal transporter 1 (DMT1) and ferroportin-1 (FPN1). Logistic analysis was used to explore the associations of maternal iron status with PE risk. The associations of placental iron transport proteins with maternal iron status were explored. **Results:** After adjusting for covariates, dietary total iron, non-heme iron intake and serum hepcidin were negatively associated with PE, with adjusted ORs (95% CIs) were 0.40 (0.17, 0.91), 0.42 (0.18, 0.94) and 0.02 (0.002, 0.13) for the highest versus lowest tertile, respectively. For the highest tertile versus lowest tertile, serum iron (4.08 (1.58, 10.57)) and ferritin (5.61 (2.36, 13.31)) were positively associated with PE. The mRNA expressions and protein levels of DMT1 and FPN1 in placenta were up-regulated in the PE group ($p < 0.05$). The mRNA expressions of DMT1 and FPN1 in placenta showed a negative correlation with the serum hepcidin ($r = -0.71$, $p < 0.001$; $r = -0.49$, $p < 0.05$). **Conclusions:** In conclusion, the maternal iron status were closely associated with PE risk, placental DMT1 and FPN1 were upregulated in PE which may be a promising target for the prevention of PE.

Key Words: pre-eclampsia, iron, hepcidin, divalent metal transporter 1 (DMT-1), ferroportin-1 (FPN1)

INTRODUCTION

Pre-eclampsia (PE), a common pregnancy complication, is defined as the presence of new-onset hypertension and proteinuria or other end-organ damage occurring after 20 weeks gestation.¹ PE affects an estimated 2.2% of pregnancies in China and is the biggest single cause of maternal and child mortality.² Serious outcomes caused by PE include abruptio placentae, pulmonary edema, hepatic failure and acute renal failure.^{3,4} Currently, there is no effective therapeutic treatment for PE other than termination of the pregnancy.

The pathogenesis of PE has not been fully understood, and it is believed to be associated with endothelial dysfunction,⁵ inflammation,⁶ genetic factors,⁷ and oxidative stress at the maternal-fetal interface.³ Iron is essential for many fundamental biological processes, including energy metabolism, oxygen transport, DNA synthesis and metabolism, et al.⁸ However, excess iron can result in increased oxidative stress by participating in Fenton reactions.⁹ It has been revealed that iron homeostasis is disrupted in patients with PE.¹⁰ In addition, recent studies have shown positive associations of serum levels of ferritin and iron with PE.^{11,12} In particular, as the key hormone regulating systemic iron metabolism, hepcidin was also detected in PE with inconsistent results. Some studies have found higher

serum hepcidin concentrations in PE compared to controls,^{13,14} while other studies have shown no difference or even opposite results.^{15,16} In addition, dietary iron intake of pregnant women have also been linked with PE,^{17,18} which is regarded as a modifiable condition. Most previous studies showed the dietary total iron intake was inversely associated with the development of PE, while the relationship between iron intake from different sources and PE risk was only one study conducted in Iranians.¹⁹ To date, most studies investigating the association of maternal iron status and dietary iron intake with the risk of PE have been conducted in Western countries,^{20,21} and evidence from Chinese population remains limited.

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The placenta is the only link between fetus and mother during gestation. Dysregulated iron metabolism in the placenta is also an important factor in the development of PE.³ Excessive iron accumulation in the placenta can lead to the production of reactive oxygen species (ROS), placental dysfunction and trophoblast injury which are recognized factors involved in the occurrence of PE.²² Generally, multiple transporters participate in regulating the absorption and transplacental transport of iron.²³ Divalent metal transporter (DMT1) is a transmembrane divalent metal transfer protein responsible for transferring ferrous iron from endosomes into the cytoplasm of placental syncytiotrophoblast cells.²⁴ In addition, intracellular iron enters fetal circulation through ferroportin-1 (FPN1) located on the basal membrane of placental syncytiotrophoblasts facing towards the fetus.²⁵ A recent study showed that the expression of FPN1 protein in placenta was downregulated in PE.²⁶ Previous study showed that the mRNA of DMT1 in placenta was down-regulated in gestational diabetes mellitus (GDM) patients.²⁷ The alteration of the expression of placental DMT1 can cause iron metabolism disorder, and then induce the production of ROS which results in endothelial dysfunction in placenta.²⁸ To date, whether the expression of DMT1 in the placenta is altered in PE has never been examined. In addition, the potential relationships between these placental iron transport proteins and maternal iron status remain unexplored in PE.

In view of the limitations and conflicting outcomes of previous studies, this study aimed to examine the expression of placental iron transport proteins and maternal iron status and evaluate their relationships with PE during late pregnancy.

METHODS

Study population

The subjects of study were recruited from the Affiliated Hospital of Qingdao University (Qingdao, China). PE is new-onset hypertension (systolic blood pressure ≥ 140 mmHg or diastolic blood pressure of ≥ 90 mmHg) that develops after 20 weeks of gestation combined with proteinuria (>300 mg/day).¹ All subjects were older than 20 years, ≥ 28 gestational weeks, and had good compliance and communication skills. Exclusion criteria included multiple pregnancies, infectious diseases, immune system diseases, diabetes, heart, liver or renal disease and fetal malformation. Finally, a total of 94 subjects were identified as PE patients and 112 healthy pregnant women with similar age and gestational week hospitalized during the same period (± 1 week) were randomly selected as the corresponding controls.

The study was approved by the Ethics Committee of Medical College of Qingdao University (QDU-HEC-2022287). Informed consents were obtained from all participants.

Sample size calculation

Sample size calculation formula for unmatched case-control study was used to estimate sample size. With reference to relevant publications,^{29,30} the parameters were determined as follows: $P_0 = 0.14$, $OR = 3.8$, $\alpha = 0.05$, $\beta = 0.10$. $P_1 = (OR \times P_0) / (1 - P_0 + OR \times P_0)$, $\bar{P} = (P_1 + P_0) / 2$. The high hemoglobin exposure rate (P_0) in the control group was 14%.

The odds ratio between the high hemoglobin levels and PE was 3.8. The statistical power (β) was set at 0.10 and the test level was set at 0.05. Plug the above parameters into the formula and confirmed that each group need at least 68 cases. Therefore, the sample size of this study can meet the statistical requirements.

$$n = [Z_{1-\alpha/2} \sqrt{2\bar{P}(1-P)} + Z_{\beta} \sqrt{P_1(1-P_1) + P_0(1-P_0)}]^2 / (P_1 - P_0)^2$$

Socio-demographic information survey

Each participant was interviewed using a standardized questionnaire, including age, educational level, gestational weight gain, gravidity, family history of hypertension.

The preconceptional body mass index (BMI) was calculated from self-reported pre-pregnancy body weight and body height. BMI was calculated as a ratio of weight (kg) to height squared (m^2).

Food Frequency Questionnaire (FFQ) survey

Dietary data were collected by a FFQ during face-to-face interviews by an experienced dietitian. The questionnaire was modified on the basis of the questionnaire used in the Chinese nutrition and health surveillance. Participants were asked to report the consumption frequency and the average consumption of food items according to food pictures labeled with standard portion sizes over the past three months. The daily average consumption of each item was calculated by the consumption frequency and the average consumption per time. Finally, the data were imported and computed to obtain the dietary iron intake by using NCCW 12.0 (Qingdao University, China), which is primarily based on China Food Composition 2002 and 2004.

Maternal iron status indicators

Fasting blood samples were collected from the pregnant women before delivery. After centrifuging at $3000 \times g$ at $4^\circ C$ for 5 min, the serum samples were stored at $-80^\circ C$ for further measurement. The concentrations of serum iron were measured using inductively coupled plasma mass spectrometry (ICP-MS). The concentrations of serum ferritin were determined by radioimmunoassay. Serum transferrin levels were measured using an automatic biochemical analyzer (Olympus AU640, Japan). The levels of serum hepcidin were measured using a commercial ELISA kit (Jiancheng Bioengineering Institute, Nanjing, China).

Western blot analysis of placental DMT1 and FPN1

The placenta samples (50 mg) were homogenized in cold RIPA lysis buffer and centrifuged to collect supernatant. The extracted total proteins were qualified by a BCA Protein Quantitation Kit and then loaded onto 12% SDS-PAGE followed by transferring to fluoride (PVDF) membranes. After blocking with skim milk, it was incubated overnight with different primary antibodies (Absin, America): β -actin (1:1000), DMT1 (1:1000), FPN1 (1:1000). Afterwards, the membranes were washed three times with TBST and incubated with a secondary antibody (1:10000) at room temperature for 1 h. The β -actin was utilized as a control. The protein bands were observed by an enhanced chemiluminescence (ECL) localization reagent and quantified by Tanon GIS analysis.

Real-time quantitative PCR analysis of placental DMT1 and FPNI

Trizol reagent and a NanoDrop spectrophotometer (NanoDrop Technologies, Wilmington, DE, USA) were utilized to extract total RNA from the placenta tissues and evaluate the amount and purity of RNA, respectively. Purity and quantity of RNA were assessed and 1 µg RNA was reversely transcribed into cDNA. The real-time quantitative PCR was carried out using SYBR Premix Ex Taq fluorescent quantitative PCR (Eppendorf, Framingham, MA, USA). The PCR cycling conditions were as follows: a cycle of 95°C for 2 min, 40 cycles of 95°C for 5 s and 60°C for 10 s.

Statistical analysis

Baseline characteristics were presented as means ± SDs for normally distributed variables, medians (ranges) for non-parametrically distributed variables, and numbers (percentages) for categorical variables. Student's *t*-test or Mann-Whitney *U*-test was used to compare the mean levels of normal distributions or with non-normal distributions. Chi-square test was used to compare the distribution of categorical variables. Dietary and serum indicators were categorized based on tertiles (Tertile 1: <33.3th percentile, Tertile 2: ≥33.3th to 66.6th percentile, Tertile 3: ≥66.6th percentile). Multivariate logistic regression was used to investigate the associations of the dietary iron intake and serum iron indicators with the risk of PE. Odds ratio (OR) and 95% confidence interval (CI) were calculated in Crude Model and Adjusted Model. Crude Model: no covariates were adjusted. Adjusted Model: age, preconceptional BMI, gestational weight gain, gravidity and family history of hypertension were adjusted. Spearman correlation analysis

was used to evaluate the relationship between maternal iron status and the expression of placental iron transport proteins. GraphPad Prism 8.0 and Figdarw³ were used for drawing of figures. All statistical analyses were conducted in SPSS (version 22; IBM Corp., Armonk, NY, USA). A two-tailed *p*-value <0.05 was considered statistically significant. Bootstrap analysis was used to verify the stability of the results.

RESULTS

Characteristics of participants

In this case-control study, 206 participants were included. The characteristics of PE patients (*n* = 94) and controls (*n* = 112) were presented in Table 1. Participants in the PE group had higher preconceptional BMI levels (*p* < 0.001) than control group. The proportion of first pregnancy was higher in the PE group. There were no significant differences in age, maternal educational level, family history of hypertension, gestational weight gain, history of abortion and dietary intake of energy, carbohydrate, fat and protein between PE and controls (*p* > 0.05).

Comparison of maternal iron status between two groups

As shown in Figure 1, the dietary intake of total iron and non-heme iron in PE group were significantly lower than those in control group (*p* < 0.05). The PE patients had higher levels of serum iron and ferritin and lower levels of serum transferrin and hepcidin than controls (*p* < 0.05).

Table 1. Baseline characteristics of included participants

| Characteristics | Control (<i>n</i> = 112) | PE (<i>n</i> = 94) | <i>t/z/χ²</i> | <i>p</i> |
|--|---------------------------|---------------------|--------------------------|----------|
| Age, years | 28.5 ± 3.76 | 29.6 ± 4.45 | -1.91 | 0.057 |
| Maternal educational level, <i>n</i> (%) | | | 1.24 | 0.537 |
| Junior and below | 31 (27.7%) | 27 (28.7%) | | |
| Senior | 26 (23.2%) | 16 (17.0%) | | |
| Undergraduate and higher | 55 (49.1%) | 51 (54.3%) | | |
| Family history of hypertension, <i>n</i> (%) | | | 2.52 | 0.113 |
| No | 109 (97.3%) | 87 (92.6%) | | |
| Yes | 3 (2.70%) | 7 (7.40%) | | |
| Preconceptional BMI, kg/m ² | 21.3 (19.5, 24.6) | 24.9 (22.6, 27.6) | -5.71 | <0.001 |
| Gestational weight gain, kg | 15.0 (12.0, 17.3) | 16.0 (12.0, 20.0) | -1.26 | 0.207 |
| Gravidity, <i>n</i> (%) | | | 7.19 | 0.006 |
| 1 | 10 (8.93%) | 21 (22.3%) | | |
| 2 | 38 (33.9%) | 33 (35.1%) | | |
| ≥3 | 64 (57.1%) | 40 (42.6%) | | |
| History of abortion, <i>n</i> (%) | | | 0.65 | 0.421 |
| No | 63 (67.00%) | 69 (61.6%) | | |
| Yes | 31 (33.00%) | 43 (38.4%) | | |
| Dietary intake | | | | |
| Energy (kcal/d) | 1557 ± 340 | 1557 ± 316 | 0.01 | 0.991 |
| Carbohydrate (g/d) | 201 ± 64.5 | 201 ± 49.9 | -0.06 | 0.950 |
| Fat (g/d) | 54.9 (47.8, 65.3) | 56.0 (49.2, 65.7) | -0.69 | 0.488 |
| Protein (g/d) | 52.1 (45.0, 66.6) | 51.1 (42.4, 62.54) | -1.26 | 0.209 |
| Serum iron, mg/L | 9.11 (7.18, 12.0) | 10.7 (9.25, 13.2) | -3.14 | 0.002 |
| Ferritin, ng/mL | 14.3 (10.9, 19.7) | 18.6 (11.2, 29.2) | -2.37 | 0.018 |
| Transferrin, ng/mL | 3.74 (3.42, 4.07) | 3.49 (2.89, 3.84) | -2.40 | 0.016 |
| Hepcidin, ng/mL | 41.3 (33.8, 43.6) | 26.0 (24.4, 28.3) | -5.70 | 0.001 |

PE, pre-eclampsia; BMI, body mass index change

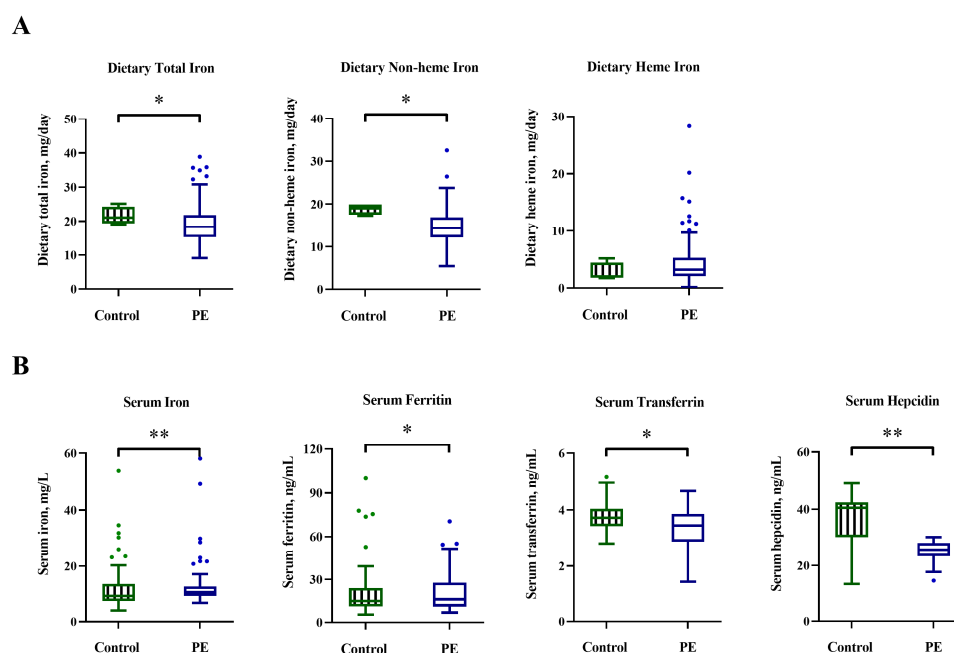


Figure 1. Comparisons of dietary iron intake and maternal iron status between the two groups. (A) Maternal dietary iron intake; (B) Maternal serum iron indicators. Asterisk indicates significant differences between PE and Control group. Data were presented as M (P_{25} , P_{75}). Mann-Whitney U -test were used to investigate the difference between control and PE. PE, Pre-eclampsia; * $p < 0.05$.

Table 2. Association between dietary iron intake and risk of PE in pregnant women

| | Crude Model OR (95% CI) | Adjusted Model OR (95% CI) |
|-----------------------|----------------------------|-------------------------------|
| Total iron, mg/day | | |
| < 16.2 | 1.00 (ref.) | 1.00 (ref.) |
| 16.2 to <20.4 | 0.50 (0.25, 0.99) * | 0.39 (0.17, 0.90) * |
| ≥ 20.4 | 0.45 (0.23, 0.88) * | 0.40 (0.17, 0.91) * |
| Heme iron, mg/day | | |
| < 2.46 | 1.00 (ref.) | 1.00 (ref.) |
| 2.46 to <4.34 | 1.00 (0.51, 1.98) | 0.84 (0.36, 2.00) |
| ≥ 4.34 | 0.78 (0.39, 1.56) | 0.70 (0.29, 1.67) |
| Non-heme iron, mg/day | | |
| <13.0 | 1.00 (ref.) | 1.00 (ref.) |
| 13.0 to <16.2 | 0.35 (0.18, 0.71) * | 0.41 (0.18, 0.94) * |
| ≥ 16.2 | 0.40 (0.20, 0.79) * | 0.42 (0.18, 0.94) * |

Crude Model: no covariates were adjusted.

Adjusted Model: adjusted age and preconceptional BMI, gestational weight gain, gravidity and family history of hypertension. * $p < 0.05$

Table 3. Association between serum iron levels and risk of PE in pregnant women

| | Crude Model OR (95% CI) | Adjusted Model OR (95% CI) |
|--------------------|----------------------------|-------------------------------|
| Serum iron, mg/L | | |
| < 8.87 | 1.00 (ref.) | 1.00 (ref.) |
| 8.87 to <11.5 | 5.06 (2.26, 11.3) ** | 5.22 (2.01, 13.5) ** |
| ≥ 11.5 | 3.64 (1.65, 8.06) ** | 4.08 (1.58, 10.6) * |
| Ferritin, ng/mL | | |
| < 15.0 | 1.00 (ref.) | 1.00 (ref.) |
| 15.0 to <20.2 | 0.56 (0.27, 1.67) | 1.54 (0.59, 4.01) |
| ≥ 20.2 | 2.03 (1.00, 4.12) | 5.61 (2.36, 13.3) ** |
| Transferrin, ng/mL | | |
| < 3.40 | 1.00 (ref.) | 1.00 (ref.) |
| 3.40 to <3.82 | 0.52 (0.18, 1.50) | 0.68 (0.20, 2.32) |
| ≥ 3.82 | 0.31 (0.10, 0.92) * | 0.46 (0.13, 1.65) |
| Hepcidin, ng/mL | | |
| < 25.9 | 1.00 (ref.) | 1.00 (ref.) |
| 25.9 to <36.3 | 0.54 (0.15, 2.04) | 0.44 (0.09, 2.20) |
| ≥ 36.3 | 0.04 (0.001, 0.16) ** | 0.02 (0.002, 0.13) ** |

Adjusted Model: adjusted age and preconceptional BMI, gestational weight gain, gravidity and family history of hypertension were adjusted. * $p < 0.05$; ** $p < 0.01$.

Associations of and maternal iron status with PE risk

The associations of maternal iron status with PE risk were presented in Table 2 and Table 3, respectively. After adjusting for age, preconceptional BMI, gestational weight gain, gravidity and family history of hypertension, the participants with highest tertile of dietary total iron, non-heme iron intake and serum hepcidin had a lower risk of PE compared with the lowest tertile, the corresponding ORs (95% CIs) were 0.40 (0.17, 0.91), 0.42 (0.18, 0.94) and 0.02 (0.002, 0.13), respectively. For the highest tertile versus lowest tertile, the concentrations of serum iron (4.08 (1.58, 10.57)) and ferritin (5.61 (2.36, 13.31)) were positively associated with the risk of PE. There was no association between the level of transferrin and PE risk. The ORs and 95%CI of age, pre-pregnancy BMI and gravidity in multivariate logistic regression analysis were shown in Supplementary Table 1.

Comparisons of mRNA expressions and protein levels of placental iron transport proteins between two groups

The mRNA expressions and protein levels of DMT1 and FPN1 in placenta were detected by RT-qPCR and western blot, respectively (Figure 2). Compared with the control group, the mRNA expressions and protein levels of DMT1 and FPN1 were both higher in the PE group ($p < 0.05$).

Correlation analysis of mRNA expressions of placental iron transport proteins and maternal iron status

The correlations between mRNA expressions of DMT1, FPN1 in placenta and maternal iron status were shown in Figure 3. The mRNA expression of FPN1 was positively correlated with the serum concentrations of ferritin ($r = 0.33$, $p < 0.05$). The mRNA expressions of DMT1 ($r = -0.71$, $p < 0.05$) and FPN1 ($r = -0.49$, $p < 0.05$) were negatively correlated with serum hepcidin.

DISCUSSION

In present case-control study, we investigated the associations of dietary iron intake and maternal iron status with the risk of PE. We focused on the expressions of the DMT1 and FPN1 in placenta, and explored the correlation between placental iron metabolism and maternal iron status. We found that dietary total iron, non-heme iron intake and serum hepcidin were negatively associated with the risk of PE. Serum iron and ferritin were positively associated with the risk of PE. In addition, the mRNA expressions and protein levels of DMT1 and FPN1 in placenta were up-regulated in PE patients compared to controls. The mRNA expressions of FPN1 and DMT1 were negatively correlated with maternal serum hepcidin, while mRNA expression of FPN1 was positively correlated with maternal serum ferritin.

Previous studies have evaluated the associations between dietary iron intake and the risk of PE.^{18,19} Consistent with several previous epidemiological studies, we found dietary total iron intake was inversely associated with the development of PE. Allen demonstrated that insufficient maternal dietary iron intake causes hypoxia, which stimulates the secretion of stress hormones, e.g., norepinephrine and cortisol, which increase the risk of placental oxidative stress.³¹ Therefore, ensuring adequate iron intake during pregnancy may reduce the risk of PE effectively. Further, we analyzed the associations between different sources of dietary iron and PE risk, which remains no evidence in the Chinese population. Results showed that dietary non-heme iron intake was inversely associated with the risk of PE, while heme iron intake was not significantly associated with PE risk. Our results were consistent with the results of a study in an Indian,¹⁹ which was the only study to explore the relationship between different sources of dietary iron and PE. The large-scale cohort study in diverse ethnic populations were needed to verify the associations of dietary iron intake with PE.

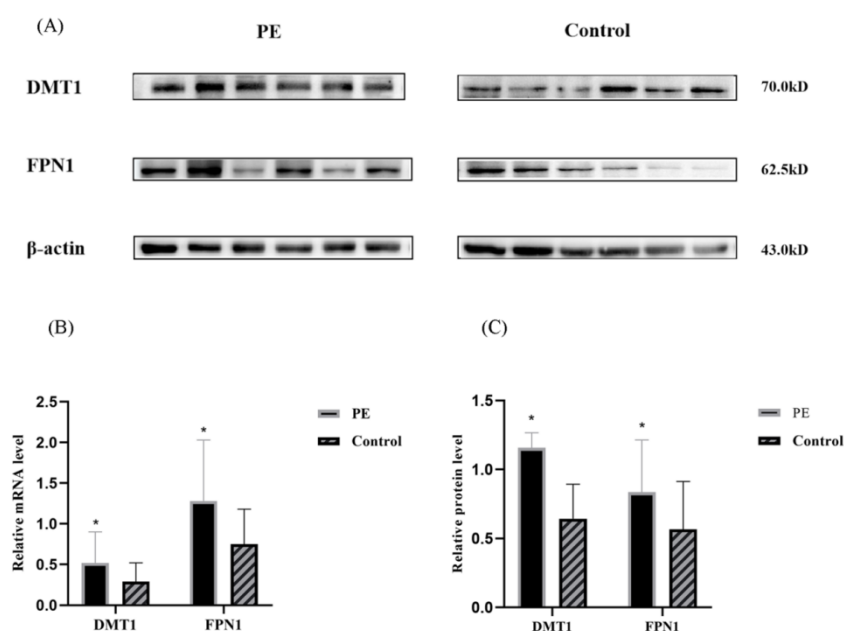


Figure 2. The mRNA expressions and protein levels of DMT1 and FPN1 in the placenta of PE and Control. (A) Western blot assay of proteins in placenta, $n = 6$ each group; (B) The Relative mRNA expression of DMT1 and FPN1, $n = 30$ each group; (C) Quantitative analyses of the DMT1 and FPN1 proteins. Data were presented as means \pm SD. Student's t -test was used to investigate the difference between control and PE. * $p < 0.05$

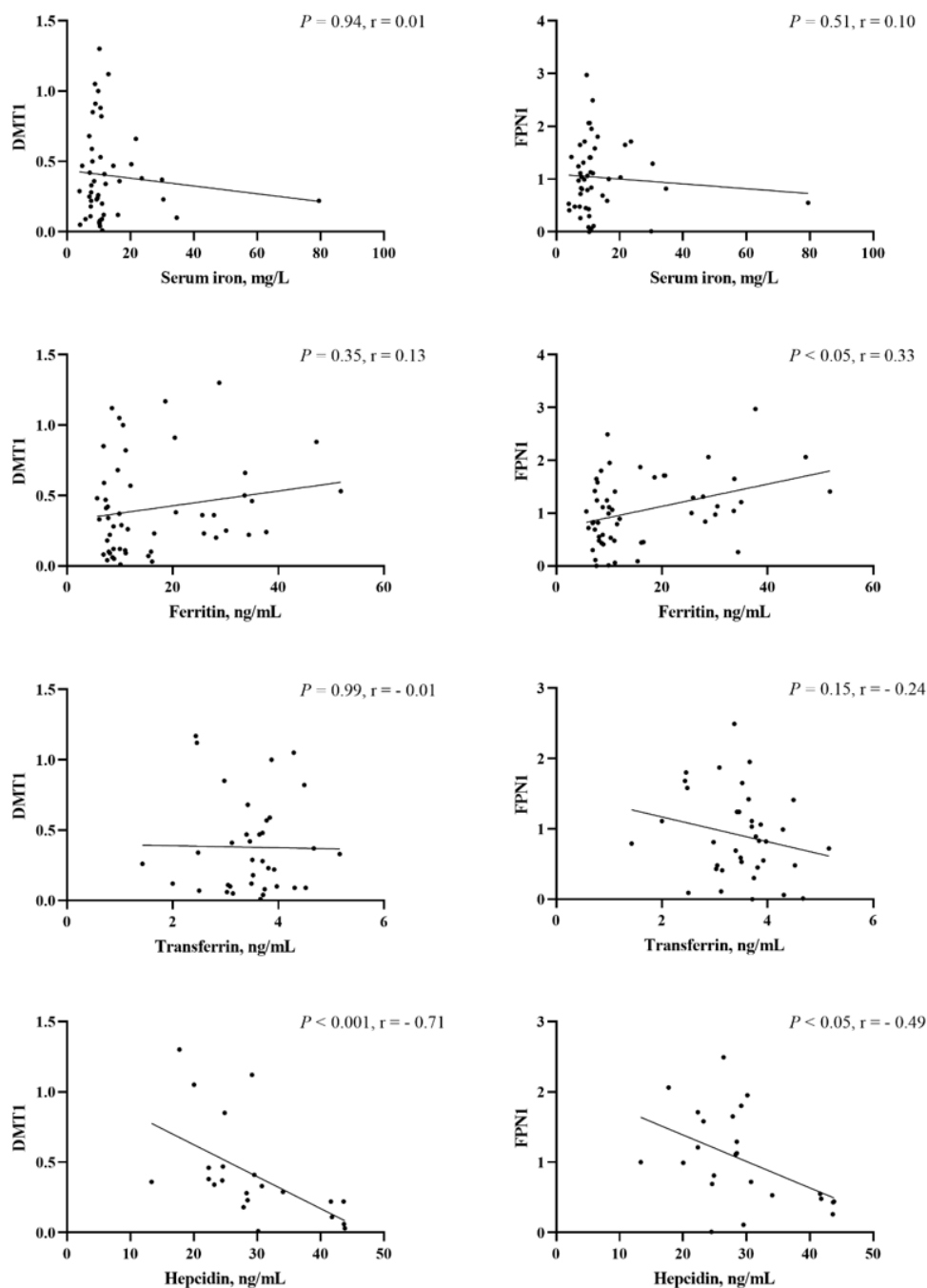


Figure 3. Correlations of placental DMT1 and FPN1 mRNA expressions with maternal serum iron indicators. Spearman correlation analysis was performed on placental DMT1 and FPN1 mRNA expressions and maternal serum iron indicators.

Moreover, the relationship between the maternal iron status and PE risk was analyzed in the study. First, our study showed that higher serum iron was associated with an increased risk of PE, which is consistent with most of previous studies.^{20,32-34} Several existing meta-analyses had also confirmed this relationship.³⁵⁻³⁷ Although the underlying molecular mechanism of the relationship between iron and PE was unclear, excess iron had been shown to be associated with lipid peroxidation which had been shown to be involved in the pathogenesis of PE. Sedaret al. reported that the levels of iron and lipid peroxides both in plasma and placental tissue of PE patients were significantly elevated.²⁰ In addition, our results showed that higher serum ferritin was associated with an increased risk of PE. At present, the evidence of this association is limited and

controversial,^{11,38,39} and it had not been explored in the Chinese pregnant women. As a well-recognized marker of iron storage *in vivo*, serum ferritin plays a key role in the regulation of iron metabolism.^{40,41} The increase of iron content in pregnant women can cause the increase of inflammation, oxidative stress and insulin resistance in the body,⁴² suggesting that the increase of serum ferritin is related to the higher PE risk. It has been suggested that abnormal increase of serum ferritin level can destroy the structure of vascular endothelial cells, affect their function, and then lead to vasoconstriction, endothelial cell injury and systemic inflammation,¹² which were the promoting factors involved in the development of PE. Therefore, our findings suggested that pregnant women should strengthen the

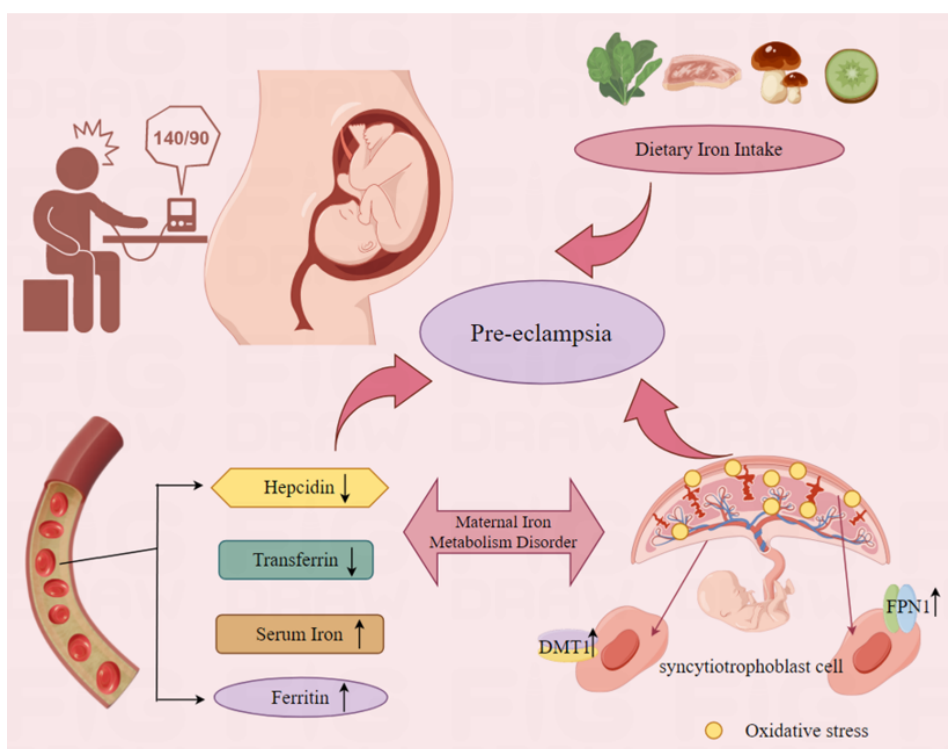


Figure 4. Maternal iron status, placental iron transport and the development of PE

detection of iron status *in vivo* and reasonable iron supplementation is recommended.

Hepcidin had been the focus of many studies on maternal iron regulation in pregnancy and iron transfer to the fetus.^{43,44} The association between hepcidin and PE risk has gained much attention with mixed conclusions.^{8,16,38,45,46} Consistent with studies of Brunacci and Masoumi,^{8,16} our results showed that the serum hepcidin was lower in patients with PE compared with controls. However, Cardaropoli and Ahmed et al. suggested that there was no difference in hepcidin between PE and healthy pregnant women,^{10,38} while several other studies reached the opposite conclusion.^{10,15} Our present findings seem to conflict with the pronounced inflammatory status of the PE. Brunacci et al. found that hepcidin was lower in the PE group, and at the same time, they found a significant positive correlation between serum hepcidin and CRP concentration and suggested that mechanisms involved in stimulating hormone production in the inflammatory response may be preserved.¹⁶ In addition, previous studies had shown that the disturbance of iron metabolism may result in hypoxia *in vivo*,⁴⁷ which may inhibit the expression of hepcidin. Hypoxia inducible factor (HIF) is activated under hypoxia status, which inhibits the promoter region of hepcidin and reduces the expression of hepcidin.⁴⁸ The production of erythropoietin (EPO) increased under hypoxia environment, which further down-regulated the expression of hepcidin.^{49,50} Lower hepcidin levels observed in PE suggested a hormonal drive to increase maternal iron bioavailability in these pregnancies, a phenomenon that may underlie the iron overload previously described in PE pregnancies.³⁶ Taken together, our findings suggest that dysregulation of iron homeostasis in PE might be caused by inadequate production of hepcidin, leading to an increased risk of developing iron overload. However, the

mechanism of decreased hepcidin levels in PE patients is still unclear and needs further study.

The central role of the placenta in the pathogenesis of PE is undisputed. The imbalance of placental iron homeostasis may lead to the accumulation of lipid peroxidation and lipid peroxides, which may ultimately lead to placental dysfunction and the development of PE.^{51,52} As the main iron transport proteins in the placenta, DMT1 and FPN1 play an important role in maintaining the balance of iron intake and release in the placenta. At present, there were few studies focusing on the associations of placental DMT1 and FPN1 with PE development, the present study was the first time to explore it in Chinese population. The results showed that placental protein and mRNA levels of DMT1 and FPN1 were upregulated in PE patients compared to controls, indicating that the uptake and transfer of iron in the placenta of PE patients were increased. This suggests that women with PE transport iron more actively than normal pregnant woman. The upregulation of DMT1 and FPN1 in trophoblasts leads to increased iron uptake, storage, and placental sequestration, contributing to the evoked oxidative stress in placenta.²⁸ At the same time, the mRNA expressions of DMT1 and FPN1 were shown negatively correlated with maternal serum hepcidin in our study. Previous evidence suggested that the regulation of hepcidin on iron transport proteins in placenta may be similar to that in the intestine, and it is a negative regulator of iron transport.⁵³ Hepcidin can directly downregulate the expression of DMT1 and FPN1 in the intestine.^{54,55} Therefore, the upregulated expression of DMT1 and FPN1 may be due to the decrease of maternal serum hepcidin levels. Yang et al. also found a negative correlation between serum hepcidin and placental FPN1 expression.⁵⁶ They found that the level of hepcidin was decreased in gestational diabetes patients, while the mRNA expression and

protein level of FPN1 in placenta was increased, suggesting that lower hepcidin level can up-regulate FPN1 expression, thereby increasing the inflow of iron into fetal circulation. Overall, our findings suggested that placental iron transport proteins were pronouncedly associated with the development of PE, DMT1 and FPN1 may be a promising target for the prevention and treatment of PE. However, the systemic and local mechanisms regulating placental iron transport remain poorly understood, the exact mechanism needs to be further explored *in vivo* and *in vitro* in the future.

This study was the first to comprehensively analyze the relationship between dietary iron intake, maternal iron metabolism status, placental iron transport proteins and PE in Chinese pregnant women. In addition, we focused on the expression changes of placental iron transport proteins in PE patients, and tried to explore the possible mechanism related to the associations between iron metabolism and PE development through a series of correlation analyses. Moreover, this study employed a case-control design to provide a theoretical basis for the exploration of the etiology of PE, but causal inference should be cautious. Our study had some limitations. First, PE patients in this present study were not classified according to the severity of the disease. In the future, it may be necessary to further explore the association between the iron status, iron transport protein of placenta and the severity of the disease. Second, our findings rely on self-reported dietary data, which may be subject to recall bias. Third, this study did not detect the indicators of inflammation and oxidative stress in pregnant women. In addition, the sample size is slightly small. Prospective cohort studies with large sample size are needed to confirm the associations in the future.

Conclusion

In conclusion, the dietary iron intake and maternal iron status were closely associated with PE risk, placental DMT1 and FPN1 were upregulated in PE which may be a promising target for the prevention and treatment of PE.

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AUTHOR DISCLOSURES

No competing interests are reported.

REFERENCES

- Mol BWJ, Roberts CT, Thangaratinam S, Magee LA, de Groot CJM, Hofmeyr GJ. Pre-eclampsia. *Lancet*. 2016; 387: 999-1011. doi: 10.1016/s0140-6736(15)00070-7.
- Yang Y, Le Ray I, Zhu J, Zhang J, Hua J, Reilly M. Preeclampsia Prevalence, Risk Factors, and Pregnancy Outcomes in Sweden and China. *JAMA Netw Open*. 2021; 4: e218401. doi: 10.1001/jamanetworkopen.2021.8401.
- Rana S, Lemoine E, Granger JP, Karumanchi SA. Preeclampsia: Pathophysiology, Challenges, and Perspectives. *Circ Res*. 2019; 124: 1094-112. doi: 10.1161/circresaha.118.313276.
- Phipps EA, Thadhani R, Benzing T, Karumanchi SA. Author Correction: Pre-eclampsia: pathogenesis, novel diagnostics and therapies. *Nat Rev Nephrol*. 2019; 15: 386. doi: 10.1038/s41581-019-0156-1.
- Redman CW, Sargent IL. Latest advances in understanding preeclampsia. *Science*. 2005; 308: 1592-4. doi: 10.1126/science.1111726.
- Palei AC, Spradley FT, Warrington JP, George EM, Granger JP. Pathophysiology of hypertension in pre-eclampsia: a lesson in integrative physiology. *Acta Physiol (Oxf)*. 2013; 208: 224-33. doi: 10.1111/apha.12106.
- Romero R, Chaiworapongsa T. Preeclampsia: a link between trophoblast dysregulation and an antiangiogenic state. *J Clin Invest*. 2013; 123: 2775-7. doi: 10.1172/jci70431.
- Masoumi Z, Hansson LR, Hansson E, Ahlm E, Mezey E, Erlandsson L, Hansson SR. Assessing erythroferrone and iron homeostasis in preeclamptic and normotensive pregnancies: A retrospective study. *Placenta*. 2023; 133: 10-8. doi: 10.1016/j.placenta.2023.01.008.
- Kell DB, Pretorius E. Serum ferritin is an important inflammatory disease marker, as it is mainly a leakage product from damaged cells. *Metallomics*. 2014; 6: 748-73. doi: 10.1039/c3mt00347g.
- Cardaropoli S, Todros T, Nuzzo AM, Rolfo A. Maternal serum levels and placental expression of hepcidin in preeclampsia. *Pregnancy Hypertens*. 2018; 11: 47-53. doi: 10.1016/j.preghy.2017.12.008.
- Siddiqui IA, Jaleel A, Kadri HM, Saeed WA, Tamimi W. Iron status parameters in preeclamptic women. *Arch Gynecol Obstet*. 2011; 284: 587-91. doi: 10.1007/s00404-010-1728-2.
- Rayman MP, Barlis J, Evans RW, Redman CW, King LJ. Abnormal iron parameters in the pregnancy syndrome preeclampsia. *Am J Obstet Gynecol*. 2002; 187: 412-8. doi: 10.1067/mob.2002.123895.
- Shaji Geetha N, Bobby Z, Dorairajan G, Jacob SE. Increased hepcidin levels in preeclampsia: a protective mechanism against iron overload mediated oxidative stress? *J Matern Fetal Neonatal Med*. 2022; 35: 636-41. doi: 10.1080/14767058.2020.1730322.
- Amstad Bencaiova G, Vogt DR, Hoesli I. Serum hepcidin and iron status parameters in pregnant women and the association with adverse maternal and fetal outcomes: a study protocol for a prospective cohort study. *BMJ Open*. 2019; 9: e032280. doi: 10.1136/bmjopen-2019-032280.
- Toldi G, Stenczer B, Molvarec A, Takáts Z, Beko G, Rigó J Jr, Vászárhelyi B. Hepcidin concentrations and iron homeostasis in preeclampsia. *Clin Chem Lab Med*. 2010; 48: 1423-6. doi: 10.1515/CCLM.2010.290.
- Brunacci F, Rocha VS, De Carli E, Esposito BP, Ruano R, Colli C. Increased serum iron in preeclamptic women is likely due to low hepcidin levels. *Nutr Res*. 2018; 53: 32-9. doi: 10.1016/j.nutres.2018.03.005.
- Kinshella MW, Omar S, Scherbinsky K, Vidler M, Magee LA, von Dadelszen P, Moore SE, Elango R; PRECISE Conceptual Framework Working Group. Maternal nutritional risk factors for pre-eclampsia incidence: findings from a narrative scoping review. *Reprod Health*. 2022; 19: 188. doi: 10.1186/s12978-022-01485-9.
- Liu Y, Wang X, Fu W, Cao Y, Dou W, Duan D, Zhao X, Ma S, Lyu Q. The association between dietary mineral intake and the risk of preeclampsia in Chinese pregnant women: a matched case-control study. *Sci Rep*. 2023; 13: 16103. doi: 10.1038/s41598-023-43481-4.
- Hajianfar H, Abbasi K, Azadbakht L, Esmailzadeh A, Mollaghasemi N, Arab A. The Association between Maternal Dietary Iron Intake during the First Trimester of Pregnancy with Pregnancy Outcomes and Pregnancy-Related Complications. *Clin Nutr Res*. 2020; 9: 52-62. doi: 10.7762/cnr.2020.9.1.52.

20. Serdar Z, Gur E, Develioglu O. Serum iron and copper status and oxidative stress in severe and mild preeclampsia. *Cell Biochem Funct.* 2006; 24: 209-15. doi: 10.1002/cbf.1235.
21. Sarwar MS, Ahmed S, Ullah MS, Kabir H, Rahman GK, Hasnat A, Islam MS. Comparative study of serum zinc, copper, manganese, and iron in preeclamptic pregnant women. *Biol Trace Elem Res.* 2013; 154: 14-20. doi: 10.1007/s12011-013-9721-9.
22. Lewandowska M, Sajdak S, Lubiński J. Can Serum Iron Concentrations in Early Healthy Pregnancy Be Risk Marker of Pregnancy-Induced Hypertension? *Nutrients.* 2019; 11. doi: 10.3390/nu11051086.
23. Santhakumar S, Athiyarath R, Cherian AG, Abraham VJ, George B, Lipiński P, Edison ES. Impact of maternal iron deficiency anemia on fetal iron status and placental iron transporters in human pregnancy. *Blood cells Mol Dis.* 2023; 99: 102727. doi: 10.1016/j.bcmd.2023.102727.
24. Li YQ, Bai B, Cao XX, Zhang YH, Yan H, Zheng QQ, Zhuang GH. Divalent metal transporter 1 expression and regulation in human placenta. *Biol Trace Elem Res.* 2012; 146: 6-12. doi: 10.1007/s12011-011-9214-7.
25. Li YQ, Bai B, Cao XX, Yan H, Zhuang GH. Ferroportin 1 and hephaestin expression in BeWo cell line with different iron treatment. *Cell Biochem Funct.* 2012; 30: 249-55. doi: 10.1002/cbf.1843.
26. Zhang H, He Y, Wang JX, Chen MH, Xu JJ, Jiang MH, Feng YL, Gu YF. miR-30-5p-mediated ferroptosis of trophoblasts is implicated in the pathogenesis of preeclampsia. *Redox Biol.* 2020; 29: 101402. doi: 10.1016/j.redox.2019.101402.
27. Zaugg J, Melhem H, Huang X, Wegner M, Baumann M, Surbek D, Körner M, Albrecht C. Gestational diabetes mellitus affects placental iron homeostasis: Mechanism and clinical implications. *FASEB J.* 2020; 34: 7311-29. doi: 10.1096/fj.201903054R.
28. Moreno-Fernandez J, Ochoa JJ, De Paco Matallana C, Caño A, Martín-Alvarez E, Sanchez-Romero J, et al. COVID-19 during Gestation: Maternal Implications of Evoked Oxidative Stress and Iron Metabolism Impairment. *Antioxidants (Basel).* 2022; 11. doi: 10.3390/antiox11020184.
29. Phaloprakarn C, Tangjitgamol S. Impact of high maternal hemoglobin at first antenatal visit on pregnancy outcomes: a cohort study. *J Perinat Med.* 2008; 36: 115-9. doi: 10.1515/jpm.2008.018.
30. Gaillard R, Eilers PH, Yassine S, Hofman A, Steegers EA, Jaddoe VW. Risk factors and consequences of maternal anaemia and elevated haemoglobin levels during pregnancy: a population-based prospective cohort study. *Paediatr Perinat Epidemiol.* 2014; 28: 213-26. doi: 10.1111/ppe.12112.
31. James AH. Iron Deficiency Anemia in Pregnancy. *Obstet Gynecol.* 2021; 138: 663-74. doi: 10.1097/AOG.00000000000004559.
32. Gupta S, Nanda S, Singh U, Bansal S, Lal H. Evaluation of the changes in serum iron levels in pre-eclampsia. *Indian J Clin Biochem.* 1997; 12: 91-4. doi: 10.1007/BF02867964.
33. Fenzl V, Flegar-Meštrić Z, Perkov S, Andrišić L, Tatzber F, Žarković N, Duić Ž. Trace elements and oxidative stress in hypertensive disorders of pregnancy. *Arch Gynecol Obstet.* 2013; 287: 19-24. doi: 10.1007/s00404-012-2502-4.
34. Kolusari A, Kurdoglu M, Yildizhan R, Adali E, Edirne T, Cebi A, Demir H, Yoruk IH. Catalase activity, serum trace element and heavy metal concentrations, and vitamin A, D and E levels in pre-eclampsia. *J Int Med Res.* 2008; 36: 1335-41. doi: 10.1177/147323000803600622.
35. Song QY, Luo WP, Zhang CX. High serum iron level is associated with an increased risk of hypertensive disorders during pregnancy: a meta-analysis of observational studies. *Nutr Res.* 2015; 35: 1060-9. doi: 10.1016/j.nutres.2015.09.021.
36. Kim J, Kim YJ, Lee R, Moon JH, Jo I. Serum levels of zinc, calcium, and iron are associated with the risk of preeclampsia in pregnant women. *Nutr Res.* 2012; 32: 764-9. doi: 10.1016/j.nutres.2012.09.007.
37. Liu JX, Chen D, Li MX, Hua Y. Increased serum iron levels in pregnant women with preeclampsia: a meta-analysis of observational studies. *J Obstet Gynaecol.* 2019; 39: 11-6. doi: 10.1080/01443615.2018.1450368.
38. Ahmed YIB, Yagoub HS, Hassan MA, Adam I, Hamdan HZ. Maternal serum iron status, hepcidin and interleukin-6 levels in women with preeclampsia. *Front Physiol.* 2023; 14: 1049994. doi: 10.3389/fphys.2023.1049994.
39. Gutierrez-Aguirre CH, García-Lozano JA, Treviño-Montemayor OR, Iglesias-Benavides JL, Cantú-Rodríguez OG, González-Llano O, et al. Comparative analysis of iron status and other hematological parameters in preeclampsia. *Hematology.* 2017; 22: 36-40. doi: 10.1080/10245332.2016.1220120.
40. Kalantar-Zadeh K, Kalantar-Zadeh K, Lee GH. The fascinating but deceptive ferritin: to measure it or not to measure it in chronic kidney disease? *Clin J Am Soc Nephrol.* 2006; 1 Suppl 1: S9-18. doi: 10.2215/CJN.01390406.
41. Fisher AL, Nemeth E. Iron homeostasis during pregnancy. *Am J Clin Nutr.* 2017; 106: 1567S-74S. doi: 10.3945/ajcn.117.155812.
42. Gbadegesin A, Sobande A, Agbara JO, Adedeji O, Dosunmu A, Shakunle A. Serum Iron Parameters among Pre-Eclamptic and Normotensive Pregnant Patients in The Lagos State University Teaching Hospital, Ikeja, Lagos, Nigeria. *West Afr J Med.* 2020; 37: 445-9. doi: 10.1182/blood.2020005745.
43. Sangkhae V, Fisher AL, Chua KJ, Ruchala P, Ganz T, Nemeth E. Maternal hepcidin determines embryo iron homeostasis in mice. *Blood.* 2020; 136: 2206-16. doi: 10.1182/blood.2020005745.
44. Sangkhae V, Fisher AL, Wong S, Koenig MD, Tussing-Humphreys L, Chu A, Lelić M, Ganz T, Nemeth E. Effects of maternal iron status on placental and fetal iron homeostasis. *J Clin Invest.* 2020; 130: 625-40. doi: 10.1172/JCI127341.
45. Entman SS, Richardson LD, Killam AP. Altered ferrokinetics in toxemia of pregnancy: a possible indicator of decreased red cell survival. *Clin Exp Hypertens B.* 1983; 2: 171-8. doi: 10.3109/10641958309023469.
46. Samuels P, Main EK, Mennuti MT, Gabbe SG. The origin of increased serum iron in pregnancy-induced hypertension. *Am J Obstet Gynecol.* 1987; 157: 721-5. doi: 10.1016/s0002-9378(87)80037-6.
47. Fuhrmann DC, Brüne B. A graphical journey through iron metabolism, microRNAs, and hypoxia in ferroptosis. *Redox Biol.* 2022; 54: 102365. doi: 10.1016/j.redox.2022.102365.
48. Ravasi G, Pelucchi S, Buoli Comani G, Greni F, Mariani R, Pelloni I, Bombelli S, Perego R, Barisani D, Piperno A. Hepcidin regulation in a mouse model of acute hypoxia. *Eur J Haematol.* 2018; 100: 636-43. doi: 10.1111/ejh.13062.
49. Xu MM, Wang J, Xie JX. Regulation of iron metabolism by hypoxia-inducible factors. *Acta Physiologica Sinica.* 2017; 69: 598-610. doi: 10.13294/j.aps.2017.0054.
50. Liu Q, Davidoff O, Niss K, Haase VH. Hypoxia-inducible factor regulates hepcidin via erythropoietin-induced erythropoiesis. *J Clin Invest.* 2012; 122: 4635-44. doi: 10.1172/JCI63924.
51. Aouache R, Biquard L, Vaiman D, Miralles F. Oxidative Stress in Preeclampsia and Placental Diseases. *Int J Mol Sci.* 2018; 19. doi: 10.3390/ijms19051496.
52. Goulopoulou S, Davidge ST. Molecular mechanisms of maternal vascular dysfunction in preeclampsia. *Trends Mol Med.* 2015; 21: 88-97. doi: 10.1016/j.molmed.2014.11.009.

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53. Cao C, O'Brien KO. Pregnancy and iron homeostasis: an update. *Nutr Rev.* 2013; 71: 35-51. doi: 10.1111/j.1753-4887.2012.00550.x.
54. Frazer DM, Anderson GJ. The orchestration of body iron intake: how and where do enterocytes receive their cues? *Blood Cells Mol Dis.* 2003; 30: 288-97. doi: 10.1016/s1079-9796(03)00039-1.
55. Frazer DM, Wilkins SJ, Becker EM, Vulpe CD, McKie AT, Trinder D, Anderson GJ. Hepcidin expression inversely correlates with the expression of duodenal iron transporters and iron absorption in rats. *Gastroenterology.* 2002; 123: 835-44. doi: 10.1053/gast.2002.35353.
56. Yang A, Zhao J, Lu M, Gu Y, Zhu Y, Chen D, Fu J. Expression of Hepcidin and Ferroportin in the Placenta, and Ferritin and Transferrin Receptor 1 Levels in Maternal and Umbilical Cord Blood in Pregnant Women with and without Gestational Diabetes. *Int J Environ Res Public Health.* 2016; 13. doi: 10.3390/ijerph13080766.